Determination of Acetylcholinesterase and Butylcholinesterase inhibitory activities of *G. asiatica* and *G. tiliaefolia* (family Tiliaceae) leaves..

ABSTRACT

Aims: Our study was carried out to appraise the acetylcholinestrase (AChE) and butyrylcholinestrase (BChE) inhibitory activities of *Grewia asiatica* and *Grewia tiliaefolia* leaves extracts.

Study Design: For the purpose of these experiments the extracts were subjected to an in-vivo study.

Place and Duration of Study: The study was carried out between June 2016 to December 2016 in the Department of Pharmacy, Southeast University, Dhaka, Bangladesh.

Methodology: In this study, cholinesterase inhibitory effect of different fraction of crude ethanolic extract of both plants were examined using swiss albino mice at 300 mg/kg b.w. dose. We determined acetylcholinestrase (AChE) and butyrylcholinestrase (BChE) inhibitory activities using slightly modified Elman coupled enzyme assay.

Results: The highest inhibition of bovine brain acetylcholinesterase and human blood butyrylcholinesterase were exhibited by PEF and CLF of *G.asiatica* with the IC_{50} values were found to be 55.88µg/ml and 26.14 µg/ml respectively whereas the highest inhibition of bovine brain acetylcholinesterase and human blood butyrylcholinesterasewere exhibited by CLF of *G.tiliaefolia*.

Conclusion: The result of the present study of various fractions of this plants have considerable amount of anti-acetylcholinesterase and anti-butyrylcholinesterase activity which suggest its effectiveness against various neurodegenerative disorders.

Keywords: Free radicals, Acetylcholinesterase, Butyrylcholinesterase, *G.asiatica, G.tiliaefolia.*

ABBREVIATIONS

IC_{50}: Concentration of an Inhibitor

PEFGA: Petroleum ether fraction of *G. asiatica*

CEEGA: Crude ethanolic extract of *G. asitica.*

CLFGA=Chloroform fraction of *G.asiatica*

EAFGA=Ethyl acetate fraction of *G. asiatica,*

AEFGA=Aqueous Ethanolic fraction of *G. asiatica.*

AD: Alzheimer's Disease

AChE: Anti-acetylcholinesterase

BchE: Anti-butyrylcholinesterase

1. INTRODUCTION

Plants serve various purposes and their usefulness to man is not limited to their role as sources of raw materials for industries; they are also consumed as food and sometimes used as medication. For ages, plants have provided man with diverse means of healing. In fact, many parts of plants such as fruits, seeds, barks, roots, and flowers have been used as medication to provide alternative therapies for various diseases that affect man and animals [1]. Medicinal plants contain potentially useful chemicals that are currently used for the manufacturing of modern therapeutic agents [2]. The evaluation of medicinal plants, used traditionally in treating Alzheimer's disease (AD) is of growing interest. Alzheimer's disease (AD) is a silent killer, one of the major leading causes of mortality after heart disease, cancer and stroke. AD associated with memory impairment that progressively declines in cognitive abilities and behaviors, which lead to the complete functional dependency that defines the dementia phase of the illness [3]. Therefore, inhibition of acetylcholinesterase (AChE), butyrylcholinesterase (BChE) enzyme and oxidation are the two promising strategies in the development of drug for neurological diseases like Alzheimer's and as well as in the treatment of Parkinson's disease, ataxia and dementia [4]. *Grewia asiatica* and *Grewia tiliaefolia* belonging to the family Tiliaceae [5] is a tree and has been used in traditional medicine to relief of various health problems such as cold, hepatitis, diarrhoea, heat stroke, dyspepsia, tuberculosis, sexual debility troubles, rheumatism and also important to promote intellect and enhancing memory, thus supporting its possible anti-Alzheimer's properties [6-7]. Several literature reviews demonstrated that the plant *G. asiatica* and *G. tiliaefolia* possess analgesic, anti-inflammatory, antioxidant, antimalarial, antidiabetic, antiemetic, antipyretic, antifungal, antiviral, antiplatelate, anticancer and immune-modulatory activities [8-16]. Thus, our main intention of the present study is to investigate the neuroprotective effect of different extracts of two medicinal plants *i.e. G. asiatica* and *G. tiliaefolia* (Family-Tiliaceae) available in Bangladesh on animal model.

2. MATERIALS AND METHODS

2.1 Collection of the plant materials and preparation of extracts:

For this present investigation whole plant of *Grewia asiatica* & *Grewia tilieafolia* were collected from Moulovibazar, Bangladesh, in July 2016. After collection the plant was thoroughly washed with water and dried. The plants were identified by expert of Bangladesh National Herbarium, Mirpur, and DACB Accession number 73883 for *Grewia asiatica* & DACB Accession number 73882 for *Grewia tilieafoilia.* For whole plant part of the dried and powdered materials (100 g for) were soaked in 500 ml of 90% ethanol

for about 14 days at room temperature with occasional shaking. After 14 days the solution was filtered using cotton filter and Whitman's filter paper.

Fig 1: Accession number -DACB 73883 for *G. asiatica*

Fig 2: Accession number -DACB 73882 for *G. tiliaefolia*

2.1.1 Drugs and chemicals

5, 5´-dithio-bis-(2-nitro) benzoic acid (DTNB) (Sigma-Aldrich, Japan), Acetylcholine iodide (Sigma-Aldrich, Japan), Brain homogenate (Crude enzyme), Tris-Hcl buffer (Merck, Germany), Triton X-100 (Sigma chemical company, USA), BCA kit (bicinchoninic acid; Sigma Co., USA), Bovine serum albumin (Merck, India), Donepezil (Sigma-Aldrich, Japan),

2.1.2 Acetyl cholinesterase inhibitory activity assay

The anti-acetylcholinesterase (AChE) assay was performed according to the colourimetric method of Ellman et al. [17], using acetylthiocholine iodide as a substrate. For the enzyme source, the rat brains were homogenised in a homogeniser with 5 volumes of a homogenisation buffer 910 mM Tris-HCl (pH

7.2), which contained 1M NaCl, 50mM MgCl2 and 1% Triton X-1000 [18], centrifuged at 10,000 rpm for 15 min. The resulting supernatant was used as an enzyme source. All of the extraction steps were carried out at 4°C. Protein concentration was determined using the BCA kit (bicinchoninic acid) with bovine serum albumin (BSA) as a protein standard. The rates of hydrolysis by acetylcholinesterase were monitored spectrophotometrically. Each extract or standard (500µl) was mixed with an enzyme solution (500µl) and incubated at 37°C for 15 min. Absorbance at 405 nm was read immediately after adding an Ellman's reaction mixture [3.5 ml; 0.5 mM acetylthiocholine, 1 mM 5, 5´-dithio-bis (2-nitro benzoic acid)] in a 50 mM sodium phosphate buffer (pH 8.0) to the above reaction mixture. Reading was repeated for 10 min at 2 min intervals to verify that the reaction occurred linearly. The blank reaction was measured by substituting saline for the enzyme.

Acetylcholine →Thiocholine + Acetate; Thiocholine + dithiobisnitro→ Benzoate yellow color

2.1.3 Anti-butyrylcholinestrase activity assay

The anti-butyrylcholinesterase (BchE) assay was performed according to the colourimetric method of Doctor et al. [19], using butyrylthiocholine iodide as a substrate. For the enzyme source, the human blood was homogenised in a homogeniser with 5 volumes of a homogenisation buffer [10 mM Tris-HCl (pH 7.2), which contained 1M NaCl, 50mM MgCl2 and 1% Triton X-100] [17], centrifuged at 10,000 ´ rpm for 15 min. The resulting supernatant was used as an enzyme source. All of the extraction steps were carried out at 4°C. Protein concentration was determined using the BCA kit (bicinchoninic acid) with bovine serum albumin (BSA) as a protein standard. The rates of hydrolysis by butyrylcholinesterase were monitored spectrophotometrically. Each extract or standard (500 µl) was mixed with an enzyme solution (500 µl) and incubated at 37°C for 15 min. Absorbance at 405 nm was read immediately after adding an Ellman's reaction mixture [3.5 ml; 0.5mM acetylthiocholine, 1mM 5, 5´-dithio-bis (2nitro benzoic acid)] in a 50 mM sodium phosphate buffer (pH 8.0) to the above reaction mixture. Reading was repeated for 10 min at 2 min intervals to verify that the reaction occurred linearly. The blank reaction was measured by substituting saline for the enzyme.

Butyrylcholine →Thiocholine + Acetate; Thiocholine + dithiobisnitro→ Benzoate yellow Color

3. RESULTS

3.1 Acetyl cholinesterase inhibitory activity assay of *G. asiatica*

The acetylcholinesterase inhibitory activity of different extractives was determined by Ellman's method. This method estimates acetylcholinesterase (AchE) using acetylcholine iodide (substrate) and dithiobisnitro benzoic acid (DTNB). The enzymatic activity was measured by the yellow color compound produced by thiocholine when it reacts with dithiobisnitro benzoate ion. The results for anticholinesterase activity of crude ethanolic extracts different fraction of *G. asiatica* are given in Table 1.

Table 1: Acetylcholinesterase inhibitory activity of the crude ethanol extract of *G. asiatica* and Donepezil (standard) at different concentration.

Name of Sample	Conc. (µg/ml)	% of Scavenging			% of Scavenging Mean ± STD	IC_{50} (µg/ml)±STD
		a	b	C		
CEEGA	31.25	22.22	22.79	21.65	22.22±0.465	286.78±1.77
	62.5	24.50	25.64	25.07	25.07±0.465	
	125	30.77	30.20	84.64	30.77±0.465	
	250	43.87	43.30	43.87	43.48±0.269	
PEFGA	31.25	57.55	58.12	56.98	57.55±0.465	55.88±0.889
	62.5	56.41	54.70	56.70	55.94±0.881	
	125	69.52	68.95	70.09	69.52±0.465	
	250	71.22	70.66	70.94	70.94±0.232	
CLFGA	31.25	55.27	55.84	54.70	55.27±0.465	59.62±0.529
	62.5	51.85	52.42	52.99	52.42±0.465	
	125	62.39	62.68	61.54	62.20±0.484	
	250	63.82	64.39	63.53	63.91±0.355	
EAFGA	31.25	27.92	28.49	28.77	23.08±0.355	250.29±3.58
	62.5	34.19	34.76	33.90	34.28±0.355	
	125	43.30	42.17	43.87	43.11±0.711	
	250	49.00	50.71	50.14	49.95±0.711	
AEFGA	31.25	23.36	22.79	23.07	23.08±0.232	268.66±0.3.37
	62.5	37.04	37.61	36.47	37.07±0.465	
	125	40.46	42.17	41.60	41.41±0.711	
	250	45.87	46.44	47.29	46.53±0.585	
Donepezil	31.25	54.13	54.70	55.84	54.89±0.711	28.47±0.150
	62.5	71.23	71.23	71.79	71.41±0.269	
	125	73.50	73.79	74.07	73.79±0.232	
	250	75.21	75.78	76.07	75.68±0.355	

Further, the anticholinesterase activity of all the fractions of crude ethanol extract such as CEEGA, PEFGA, CLFGA, EAFGA and AEFGA have been investigated at 250 µg/ml concentration. Among the fractions the highest activity was found in PEFGA (55.88 % inhibition).

Fig 3: Acetylcholinesterase inhibitory activity of different fractions of crude ethanol extract different fraction of *G. asiatica and donepezil at* different concentration.

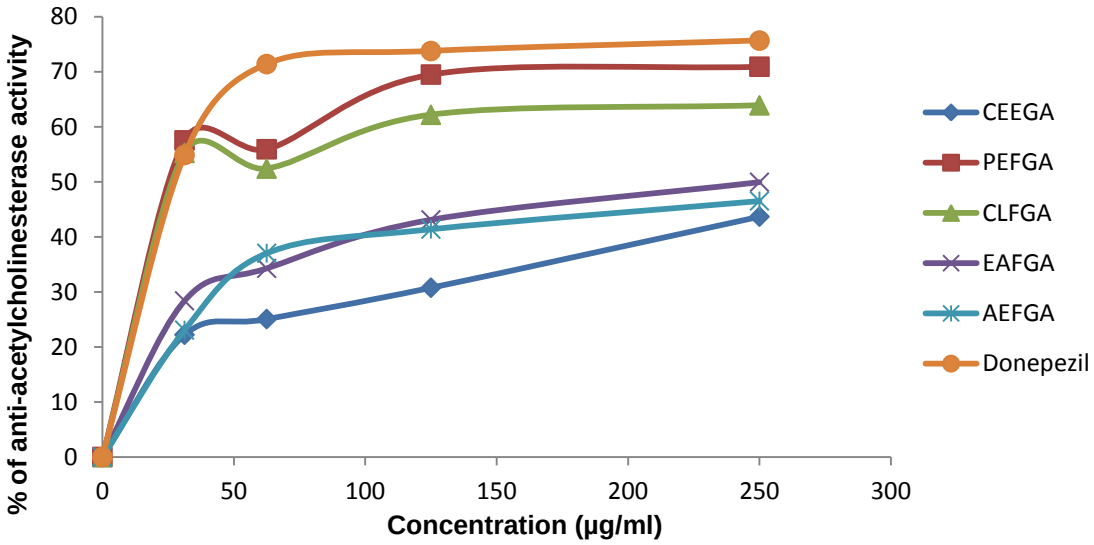
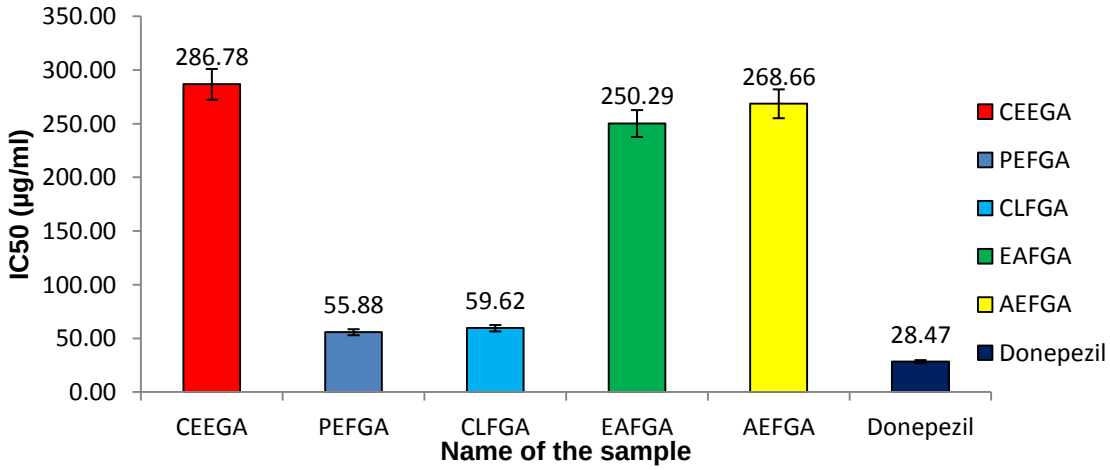


Fig 4: IC_{50} (µg/ml) values of different extractives of *G. asiatica* for Anti-acetylcholinesterase activity Assay.



3.2 Acetyl cholinesterase inhibitory activity assay of *G. tiliaefolia*

The acetylcholinesterase inhibitory activity of different extractives was determined by Ellman's method. This method estimates acetylcholinesterase (AchE) using acetylcholine iodide (substrate) and dithiobisnitro benzoic acid (DTNB). The enzymatic activity was measured by the yellow color compound produced by thiocholine when it reacts with dithiobisnitro benzoate ion. The results for anticholinesterase activity of crude ethanolic extracts different fractions of *G. tiliaefolia* are given in Table 2.

Table 2: Acetylcholinesterase inhibitory activity of the crude ethanol extract of *G.tiliaefolia* and Donepezil (standard) at different concentration.

Name of Sample	Conc. (µg/ml)	% of Scavenging			% of Scavenging Mean ± STD	IC_{50} (µg/ml)±STD
		a	b	C		
CEEGT	31.25	31.91	32.48	33.05	32.48±0.465	272.53±2.73
	62.5	38.18	38.46	37.61	38.08±0.355	
	125	39.32	38.75	39.89	39.31±0.465	
	250	45.58	46.15	46.72	46.15±0.465	
PEFGT	31.25	56.98	57.83	58.12	57.64±0.484	59.61±0.529
	62.5	52.42	51.85	52.99	52.42±0.465	
	125	64.10	63.25	63.82	63.72±0.355	
	250	68.95	68.38	69.23	68.85±0.355	
CLFGT	31.25	46.15	46.72	48.43	47.10±0.968	57.13±0.485
	62.5	54.13	55.27	54.70	54.70±0.465	
	125	56.69	57.83	58.40	57.64±0.711	
	250	60.11	58.12	59.54	59.26±0.839	
EAFGT	31.25	29.06	29.63	28.77	29.15±0.355	277.20±5.01
	62.5	36.75	37.03	34.19	35.99±1.28	
	125	40.17	40.46	42.16	40.93±0.880	
	250	44.16	46.15	45.01	45.10±0.816	
AEFGT	31.25	26.78	26.21	26.78	26.59±0.268	268.66±3.37
	62.5	31.91	32.19	32.48	32.19±0.232	
	125	39.60	39.03	40.46	39.69±0.585	
	250	45.87	46.44	47.29	46.53±0.585	
Donepezil	31.25	54.13	54.70	55.84	54.89±0.711	28.47±0.150
	62.5	71.23	71.23	71.79	71.41±0.269	
	125	73.50	73.79	74.07	73.79±0.232	
	250	75.21	75.78	76.07	75.68±0.355	

Further, the anticholinesterase activity of all the fractions of crude ethanol extract such as CEEGT, PEFGT, CLFGT, EAFGT and AEFGT have been investigated at 250 µg/ml concentration. Among the fractions the highest activity was found in CLFGT (57.17 % inhibition).

Fig. 5: Acetylcholinesterase inhibitory activity of different fractions of crude methanol extract different fraction of *G. tiliaefolia and donepezil at* different concentration.

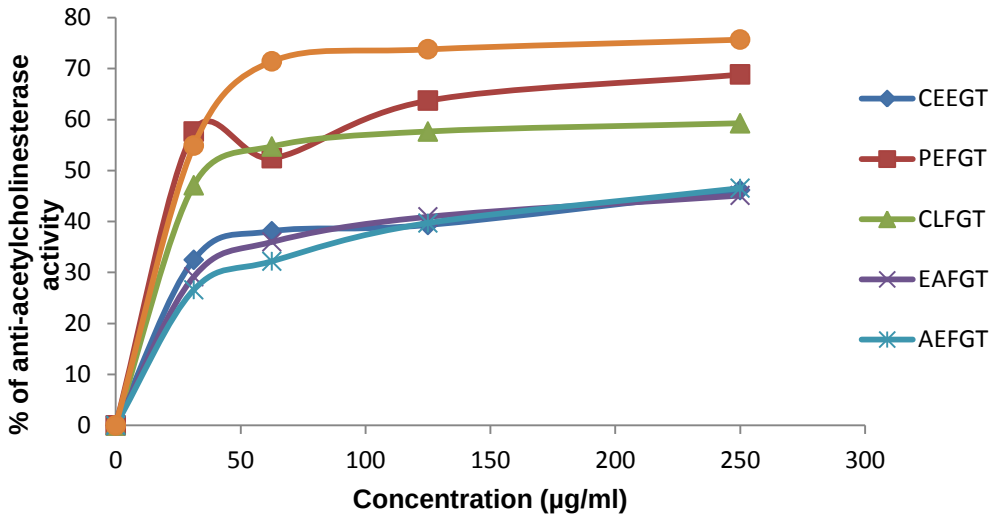
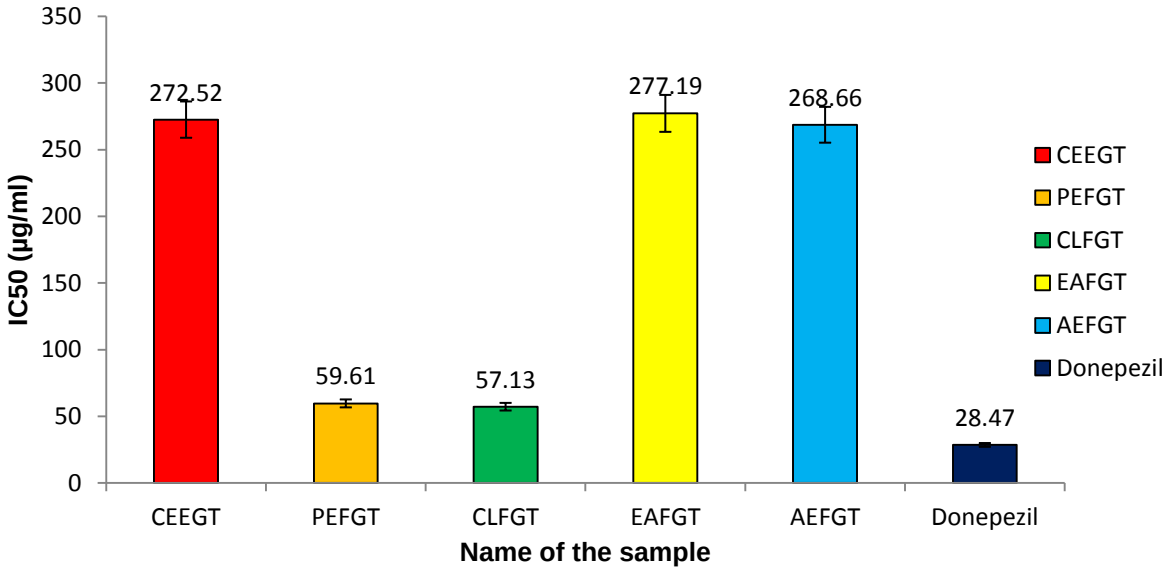


Fig.6: Acetylcholinesterase inhibitory activity of different fractions of crude methanol extract different fraction of *G. tiliaefolia and donepezil* at different concentration.



3.3 Butylcholinesterase inhibitory (BchE) activity assay of *G. asiatica*

The Butyrylcholinesterase inhibitory activity of extractives was determined by Ellman's method. This method estimates Butyrylcholinesterase (BchE) activity using butyrylythiocholine iodide (substrate) and dithiobisnitro benzoic acid (DTNB). The enzymatic activity was measured by the yellow color compound produced by thiocholine when it reacts with dithiobisnitro benzoate ion. The results for anticholinesterase activity of crude methanol extracts different fraction of *G. asiatica* are given in Table 3.

Table 3: Anti-butylcholinesterase activity assay of the crude ethanol extracts different fraction of *G. asiatica* and Donepezil (standard) at different concentrations.

Name of Sample	Conc. (µg/ml)	% of Scavenging			% of Scavenging Mean ± STD	IC_{50} (µg/ml) ±STD
		a	b	C		
CEEGA	31.25	50.33	51.00	52.67	51.33±0.981	30.84±0.577
	62.5	53.00	53.67	54.33	53.67±0.544	
	125	55.67	57.00	57.67	56.77±0.831	
	250	58.33	59.67	60.33	59.44±0.831	
PEFGA	31.25	52.33	53.33	53.00	52.89±0.416	29.54±0.233
	62.5	55.33	56.33	57.00	56.22±0.685	
	125	60.33	61.00	61.67	61.00±0.544	
	250	64.33	63.67	64.00	64.00±0.272	
CLFGA	31.25	59.67	60.00	59.67	59.78±0.157	26.14±0.069
	62.5	61.33	62.33	61.67	61.78±0.416	
	125	66.67	69.00	69.67	68.44±1.29	
	250	75.67	75.67	76.33	75.89±0.314	
EAFGA	31.25	56.33	57.00	57.33	56.89±0.416	27.47±0.201
	62.5	58.33	59.00	60.00	59.11±0.685	
	125	61.33	62.33	63.00	62.22±0.685	
	250	65.00	64.67	65.67	65.11±0.415	
AEFGA	31.25	34.33	33.67	34.33	34.11±0.314	129.33±1.47
	62.5	42.67	43.00	43.67	43.11±0.415	
	125	47.67	48.33	49.00	48.33±0.544	
	250	56.33	56.67	57.00	56.67±0.272	
Donepezil	31.25	55.00	55.67	56.00	55.56±0.416	28.13±0.170
	62.5	60.00	60.33	61.00	60.44±0.416	
	125	70.33	71.00	71.67	71.00±0.544	
	250	84.33	85.33	85.00	84.89±0.416	

Among the fractions of crude ethanol extract, CLFGA and EAFGA showed the most potent activity with IC_{50} value of 26.14µg/ml and 27.47 µg/ml which is higher than that of DON (Standard) With IC_{50} value of 28.13 µg/ml. On the other hand, (PEFGA), (CEEGA) and (AEFGA) fraction showed free radical scavenging activity with IC_{50} value of 29.54 µg/ml, 30.84µg/ml and 129.33 µg/ml respectively. Our results clearly demonstrate that the extractives of *G.asiatica* possess antiradical activity.

Fig.7: Anti-Butyrylcholinesterase activity of different fractions of crude ethanol extract of *G. asiatica* and donepezil at different concentrations.

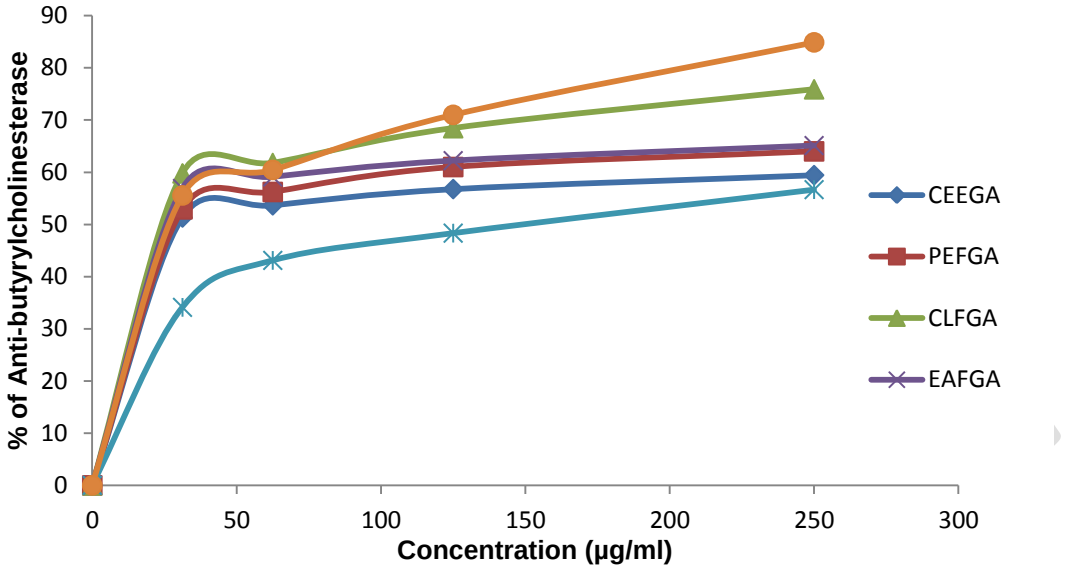
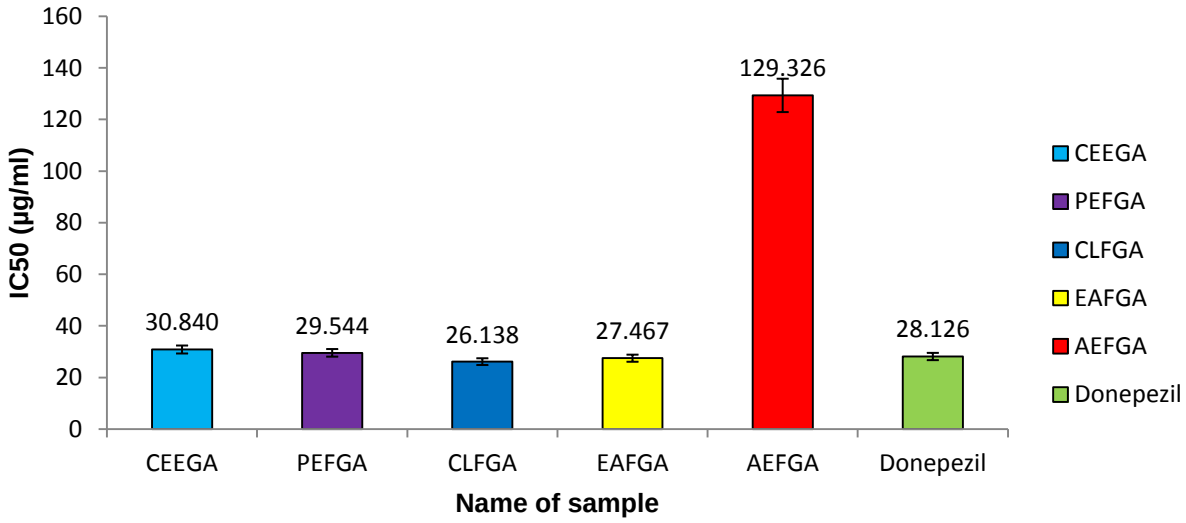


Fig.8: IC_{50} (µg/ml) values of different extractives of *G. asiatica* for Anti-Butyrylcholinesterase activity Assay.



3.4 Butylcholinesterase inhibitory (BchE) activity assay of *G. titiaefolia*

The Butyrylcholinesterase inhibitory activity of extractives was determined by Ellman's method. This method estimates Butyrylcholinesterase (BchE) activity using butyrylythiocholine iodide (substrate) and dithiobisnitro benzoic acid (DTNB). The enzymatic activity was measured by the yellow color compound produced by thiocholine when it reacts with dithiobisnitro benzoate ion. The results for anticholinesterase activity of crude methanol extracts different fraction of *G.tiliaefolia* are given in Table 4.

Table 4: Anti-butylcholinesterase activity assay of the crude ethanol extracts different fraction of *G. tiliaefolia* and Donepezil (standard) at different concentrations.

Name of Sample	Conc. (µg/ml)	% of Scavenging			% of Scavenging Mean ± STD	IC_{50} (µg/ml)±STD
		a	b	C		
CEEGT	31.25	47.67	49.00	49.67	48.78±0.831	32.33±0.550
	62.5	52.33	53.00	53.67	53.00±0.544	
	125	57.33	57.67	58.67	57.89±0.567	
	250	61.00	61.67	59.67	60.78±0.831	
PEFGT	31.25	54.67	56.00	56.33	55.67±0.720	28.07±0.366
	62.5	60.33	61.67	62.00	61.33±0.720	
	125	62.33	63.00	63.33	62.89±0.415	
	250	67.00	67.67	68.33	67.67±0.544	
CLFGT	31.25	57.67	59.00	59.67	58.78±0.831	26.59±0.378
	62.5	63.33	64.33	64.67	64.11±0.567	
	125	66.33	67.67	68.33	67.44±0.831	
	250	73.00	71.67	73.33	72.67±0.720	
EAFGT	31.25	53.00	54.00	54.33	53.78±0.567	29.06±0.308
	62.5	61.67	61.00	60.00	60.89±0.685	
	125	63.67	63.33	64.00	63.67±0.272	
	250	68.00	67.00	65.67	66.89±0.956	
AEFGT	31.25	32.00	33.00	31.67	32.22±0.567	134.25±0.92
	62.5	40.33	39.67	41.00	40.33±0.544	
	125	46.33	47.00	46.33	46.56±0.314	
	250	58.67	57.67	57.00	57.78±0.685	
Donepezil	31.25	55.00	55.67	56.00	55.56±0.416	28.13±0.170
	62.5	60.00	60.33	61.00	60.44±0.416	
	125	70.33	71.00	71.67	71.00±0.544	
	250	84.33	85.33	85.00	84.89±0.416	

Fig.9: Anti-Butyrylcholinesterase activity of different fractions of crude ethanol extract of *G.tiliaefolia* and donepezil at different concentrations.

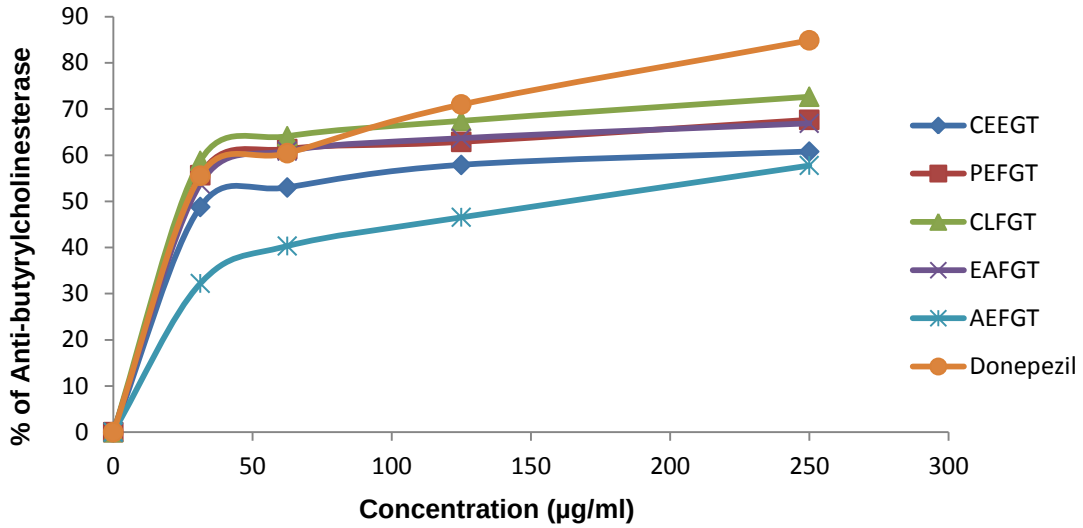
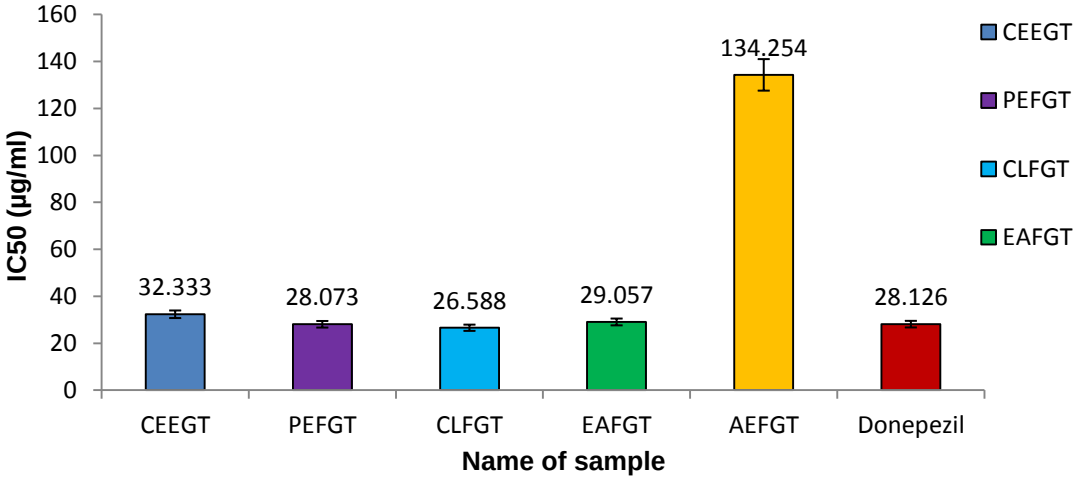


Fig.10: IC_{50} (µg/ml) values of different extractives of *G.tiliaefolia* for Anti-Butyrylcholinesterase activity Assay.



4. DISCUSSIONS

Alzheimer's disease (AD) is a progressive neurodegenerative disorder characterised by loss of memory and cognition. The AD has revealed the development of new therapeutic drugs for the AD is underway. Currently, only five medications are approved by the Food and Drug Administration to treat AD. Four of them are acetylcholinesterase inhibitors such as donepezil, galantamine, rivastigmine, tacrine and the fifth is the N-methyl-d-aspartate antagonist memantine [19]. The history of drug discovery showed that plants are highly rich sources of bioactive compounds and lead to the development of drugs for the treatment of neurological diseases including Alzheimer's disease [20]. In traditional practices of medicine

plants have been used to enhance cognitive function and to alleviate other symptoms associated with the AD. Inhibition of acetylcholinesterase enhances cholinergic transmission by reducing enzymatic degradation of acetylcholine is a promising strategy for the development of AD-drug. Acetylcholinesterase inhibitors are the only source of the compound that is currently approved for the treatment of Alzheimer's disease.

However, our results revealed moderate anti-acetylcholinesterase effect of *G.asiatica* and *G. tiliaefolia.* Also, the activity, the different fractions of the crude extract such as petroleum ether, crude ethanol extract, chloroform, ethyl acetate and aqueous fractions were examined similarly at a concentration of 250 µg/ml. Among the fractions, the CLFGA and CLFGT had the highest % of inhibition on Butyrylcholinesterase at a concentration of 250 µg/ml. However, our results revealed significant anti-butyryllcholinesterase effect of *G. asiatica* and *G. tiliaefolia.*

5 CONCLUSIONS

The present study was undertaken to investigate the *in vivo* anticholinesterase and anti-butyryllcholinesterase effects of *G. asiatica* and *G. tiliaefolia.* Inhibition of acetylcholinesterase, which enhances cholinergic transmission by reducing the enzymatic degradation of acetylcholine, is a promising strategy for the development of neurodegenarative disorders like Alzheimer's diseases drug. In vivo effectiveness of Grewia nervosa and its components remains to be investigated. The results indicate that *G. asiatica* and *G. tiliaefolia.* may be of value for an effective treatment for Alzheimer's disease.

CONSENT

It is not applicable

ETHICAL APPROVAL

The protocol of the experiment was approved by the animal ethics committee of the Department of Pharmacy, Southeast University, Dhaka, Bangladesh. The animals care and health were maintained according to the guidelines of the National Institutes of Health.

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