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2
3 **Endothelium-Independent Vasorelaxant Effects of Anthocyanins-Enriched**
4 **Extract from *Odontonema Strictum* (Nees) Kuntze (Acanthaceae) Flowers:**
5 **Ca²⁺ Channels Involvement**

6
7

8 **ABSTRACT**

9 **Aims:** We aimed in this study to investigate the mechanisms of the vasorelaxation effect caused
10 by the anthocyanins-enriched extract of *Odontonema strictum* flowers.

11 **Study Design:** Anthocyanins-enriched extract of *Odontonema strictum* flowers and
12 vasorelaxantes activities of mice aortic rings.

13 **Place and Duration of Study:** The flowers of *Odontonema strictum* (Nees) Kuntze
14 (Acanthaceae) were collected in January 2015 at the “Institut de Recherche en Sciences de la
15 Santé (IRSS)” experimental station in Ouagadougou. The experiments were conducted in
16 October - November 2018 at the department of Medicine and Traditional Pharmacopeia-
17 Pharmacy (MEPHATRA-PH)/IRSS.

18 **Methodology:** The extract was enriched in anthocyanins using Amberlite XAD-7 non-ionic resin
19 column. The vasorelaxant activity of anthocyanins-enriched extract of *O. strictum* flowers (OSF)
20 was tested using isolated organ-chamber technique with mice aorta rings.

21 **Results:** OSF showed concentration-dependent relaxant effects on mice endothelium intact or
22 denuded aortic rings pre-contracted with U46619 (10⁻⁷ M) and KCl (80 mM). OSF induced
23 relaxation in the mice aortic rings by stimulating smooth muscle cells. The vasorelaxant effect of
24 OSF (10-1000 µg/mL) was similar in endothelium-intact and endothelium-denuded aortic rings.
25 The maximum relaxant effect was 93.78 ± 4.69% and 92.30 ± 3.19% for endothelium-intact and
26 endothelium-denuded aortic rings, respectively. Moreover, after incubation of the aorta rings with
27 OSF (400 µg/mL) or vehicle (0.02% of DMSO) in PSS, OSF blocked the contraction through
28 mechanism involving inhibition of CaCl₂ and U46619 effect.

29 **Conclusions:** The present study provides a pharmacological evidence for the antihypertensive
30 medicinal use of *Odontonema strictum* by highlighting its vasorelaxant activity.

31 **Keywords:** *Odontonema strictum*; flowers; endothelium-independent; vasorelaxant; calcium
32 channels

33

34 1. INTRODUCTION

35 In the past decade, epidemiological studies have shown that cardiovascular diseases are the main
36 cause of death and disability worldwide [1, 2]. In Africa, cardiovascular diseases (CVD) have
37 reached nearly epidemic proportions. Indeed, high blood pressure is the major determinant of
38 mortality related to cardiovascular disease, cerebrovascular disease, and stroke [3]. High blood
39 pressure is a major cause of deaths in developing countries accounting for about 8-9 million
40 deaths compared to about 3-5 million in developed countries [4]. The pathophysiological
41 mechanism behind this disorder is multifactorial including oxidative stress, inflammation, renin-
42 angiotensin system and autoimmune vascular dysfunction [5-7]. Hypertension is characterized
43 by a chronic elevation of arterial blood pressure (superior or equal to 140/90 mmHg), in which
44 abnormally increased vascular tone plays a major role in the maintenance of high blood pressure
45 [2, 8].

46 However, although conventional drugs continue to be developed against hypertension, they do
47 not fully manage this condition. In such context 80% of the population resort to traditional
48 medicine for their health care, including hypertension [9]. In addition, natural drugs are another
49 alternative to synthetic drugs [10]. Natural products represent an extremely valuable source for
50 production of new chemical entities for the treatment of emerging diseases, since they represent
51 structures selected by evolutionary mechanisms over a period of millions of years through an
52 adaptation according to time and climate [1, 11]. Previous studies reported the use of plants in
53 traditional medicine to treat various diseases including cardiovascular disease. It is very
54 important to screen plants or plants extracts for the treatment of diseases such as hypertension
55 [7]. Among the available plants, *Odontonema strictum* (Nees) Kuntze (Acanthaceae) a decorative
56 plant of Latin America is known to be traditionally used for the treatment of arterial
57 hypertension [12]. Authors have reported its antihypertensive/hypotensive and vasorelaxation
58 effects on the rat and pig heart coronary arteries respectively [13]. These authors have shown
59 through pharmacological tests that the aqueous, alcoholic and ethyl acetate extracts of the leaves
60 of the plant possess antihypertensive and vasorelaxant properties. Following this work, C-

61 heteroside flavonoids and O-heteroside flavonoids were isolated in the leaves of *Odontonema*
62 *strictum* [12]. These extracts of leaves have shown antioxidant properties [9, 14]. The leaves and
63 flowers of *Odontonema strictum* contain carbohydrates, saponins of flavonoids, glycosides,
64 tannins, steroids and terpenoids as well as Stigmasterol and β -Sitosterol [15].

65 All these pharmacological investigations were mainly focused on the leaves of *Odontonema*
66 *strictum*. In addition, anthocyanins are the main phenolic compounds involved in the color of
67 flowers and they also possess physiological activities such as antioxidative, antimutagenic and
68 antihypertensive potential [16]. No data is available about the others parts of the plants such as
69 the flowers. Therefore, this study has been undertaken to assess the antihypertensive efficacy of
70 *Odontonema strictum* flowers and to characterize the vasorelaxant activity, as potential mode of
71 action.

72 **2. MATERIAL AND METHODS**

73 **2.1. Plants Material**

74 *Odontonema strictum* Flowers were collected in January 2015 at “Institut de Recherche en
75 Sciences de la Santé” experimental station in Ouagadougou (GPS coordinates N 12°22.161’, W
76 001°29.088’). Plant was properly identified, and a voucher specimen (HNBU 8702) was
77 deposited in the herbarium of the “Département Environnement et Forêt / Centre National de la
78 Recherche Scientifique et Technologique” (DEF/CNRST), Ouagadougou, Burkina Faso [13].
79 The plant material (flowers) was washed carefully before drying at lyophilization and powdered
80 into a fine powder in a blender.

81 **2.2. Extract enriched with anthocyanins**

82 We mixed 100 mL of n-hexane ($\geq 99.7\%$, Sigma-Aldrich) with 10 grams of *Odontonema*
83 *strictum* powder to remove fats and lipid compounds; then the solid phase was macerated with
84 100 mL of methanol ($\geq 99.9\%$, Sigma-Aldrich) for 24 hours at 4 °C. The maceration process with
85 methanol was repeated twice and the filtrated extracts were pooled and concentrated under
86 vacuum to dryness under 40 °C. The dried extract was solubilized in 5 mL of methanol (0.5%
87 HCl) for further purification.

88 Further enrichment of anthocyanins was obtained with 5 mL of methanol (0.5% HCl) using
89 Amberlite XAD-7 (Sigma Life Sciences), non-ionic resin column. Amberlite XAD-7 column
90 was initially washed with 0.5% HCl to remove free sugars and non-aromatic compounds. This
91 resin adsorbed the aromatic compounds including anthocyanins, whereas sugars and non-
92 aromatic compounds were eluted by washing with acidified water (0.5% HCl). The adsorbed
93 anthocyanins were eluted by acidified methanol (0.5% HCl). The pooled methanolic was
94 concentrated on rotavapor under vacuum at 40 °C to obtain dried powder.

95 **2.3. Animals**

96 Male 6-8 weeks old mice Naval Medical Research Institute (NMRI) were obtained from the pet
97 Shop of IRSS, Ouagadougou and exposed to daily light-dark 12 hours cycle with free access to
98 proteins enriched pellet (29%) and water. They were maintained in controlled temperature room
99 of 22-25°C.

100 **2.4. Preparation of mice thoracic aortic rings for isometric tension recording**

101 The method used has been previously described [17, 18]. Briefly, mice were euthanized and
102 thoracic aortas were excised, cleaned from fat tissue and cut into 2 mm length-rings. The aorta
103 was immersed in physiological salt solution (PSS, mM: 130 NaCl, 14.9 NaHCO₃, 3.7 KCl, 1.2
104 MgSO₄ 7H₂O, 1.6 CaCl₂ 2H₂O, 1.2 KH₂PO₄, and 11 glucose), pH 7.4. The PSS was continuously
105 kept at 37 °C and aerated with a pneumatic bubbling. Isolated mouse aortic rings were suspended
106 in organ chambers containing PSS and placed between 2 tungsten stirrups. Aortic rings were
107 stretched with a passive wall tension of one (01) g and an equilibrium period of 60 min was
108 allowed during which period it was washed every 20 min. After stabilization, the aorta was
109 contracted by addition of KCl (bath concentration of 80 mM). After washing, increasing
110 concentrations (10⁻⁹ - 3.10⁻⁷ M) of the thromboxane A₂ analogue agonist (U46619) were
111 cumulatively added and maximal tension of the tissue was recorded. Changes in tension were
112 recorded via isometric force transducers connected to a data acquisition system. The
113 endothelium-intact was checked by the ability of ACh (10⁻⁵ M) to induce more than 80%
114 relaxation in U46619-contracted aorta rings. When necessary, the endothelium layer was
115 removed by gently rubbing inside the lumen using forceps. Endothelium-denudation was
116 considered effectively removed when ACh (10⁻⁵ M) caused less than 10% relaxation. Ca²⁺-free

117 PSS was prepared by removing CaCl_2 . The thromboxane A2 analogue, 9,11-dideoxy-9 α ,11 α -
118 methanoepoxy $\text{PGF}_{2\alpha}$ (U46619, Merck Chemicals Ltd, Nottingham, UK) and the NO• synthase
119 inhibitor, N(ω)-nitro-L-arginine methyl ester (L-NAME, Sigma-Aldrich, 100 μM) and the non-
120 specific cyclooxygenase inhibitor, indomethacin (Indo, Sigma-Aldrich, 10 μM) were selected for
121 the experiments. The enriched anthocyanin extract of *Odontonema strictum* (OSF) flowers
122 induced vasorelaxant effects in a concentration-dependent manner (10 - 1000 $\mu\text{g}/\text{mL}$) on mice
123 aortic rings. These aortic rings intact and denuded of the endothelium are previously contracted
124 with U46619 (10^{-7} M) or KCl (80 mM) in PSS.

125 The effect of OSF (400 $\mu\text{g}/\text{mL}$) on extracellular Ca^{2+} influx was studied in Ca^{2+} -free PSS. After
126 equilibration of rings in Ca^{2+} -free PSS containing 80 mM KCl, cumulative concentrations of
127 CaCl_2 were added (10^{-5} - 10^{-2} M, respectively) with preincubation of OSF in organ bath. The
128 CaCl_2 concentration-dependent maximum contraction of the endothelium denuded aortic rings
129 with KCl (80 mM) in Ca^{2+} -free PSS was expressed as 100% for the curve constructions. The
130 aortic rings were measured after each experiments.

131 To study the relationship between the intracellular Ca^{2+} release inhibition and the OSF-induced
132 relaxation, endothelium denuded aortic rings were incubated with OSF (400 $\mu\text{g}/\text{mL}$) or vehicle
133 (0.02% of DMSO) in PSS, then the aortic rings were measured after experiment to report each
134 maximum contraction to the size of the ring.

135 **2.5. Statistical analysis**

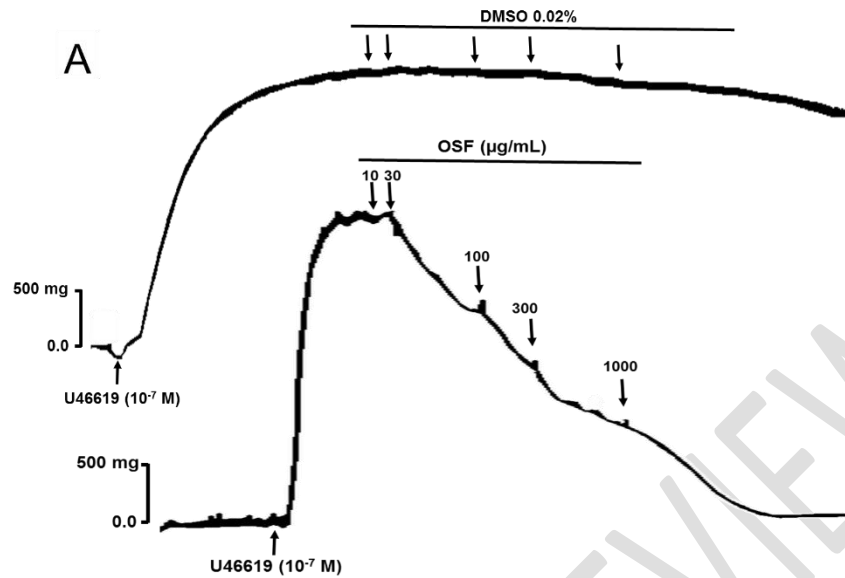
136 The experimental values were calculated by considering the maximum contraction produced by
137 U46619 of each segment equal to 100%. The baseline tension before addition of U46619 was
138 considered as 0%. The raw data have been normalized to the control (vehicle). Concentration-
139 response curves were fitted using a nonlinear interactive fitting program (Graph Pad Prism 5.0;
140 GraphPad Software, San Diego, CA), and two pharmacological parameters were obtained: the
141 maximal effect generated by the agonist (E_{max}) and pD_2 ($-\log EC_{50}$) [EC_{50} is a concentration of
142 agonist producing 50% of the maximum response]. Statistical comparisons were performed using
143 one-way ANOVA or two-way ANOVA. Post hoc test was performed using Bonferroni's test
144 analysis to compare all the groups. A p-value less than 0.05 was considered as statistically
145 significant.

146 **3. RESULTS**

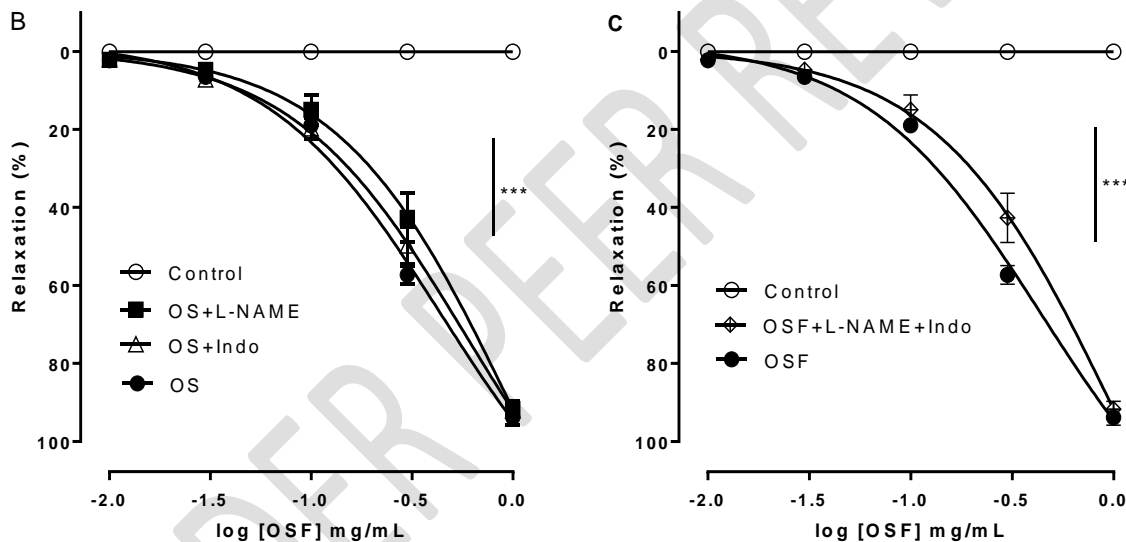
147 **3.1. Effect of anthocyanins extract of *Odontonema Strictum* flowers (OSF) on U46619-**
148 **induced contraction of endothelium-intact mice aortic rings**

149 The effect of OSF (10 - 1000 µg/mL) extract on endothelium-intact aortic rings contraction is
150 concentration dependent. In Fig. 1A, OSF effect was compared to the control while Figs. 1B, C
151 shows OSF vasorelaxant effect on the nitric oxide (NO[•]) synthesis pathway and prostacyclin
152 (PGI₂) pathway in endothelium-intact aortic rings (Figs. 1B, C). Incubation with L-NAME (100
153 µM, an eNOS inhibitor) and the combination of L-NAME and indomethacin (10 µM, a non
154 selective COX inhibitor), did not affect OSF relaxation effect on endothelium-intact aortic rings
155 pre-contracted by U46619. In the absence and presence of L-NAME or indomethacin or both of
156 them, the maximal relaxant effects (E_{max}) of OSF on endothelium-intact aortic rings were
157 respectively 93.78 ± 4.69%, 91.67 ± 4.31%, 91.99 ± 2.18% and 89.45 ± 5.99%. The pD₂ of the
158 different relaxation conditions are presented in the table.

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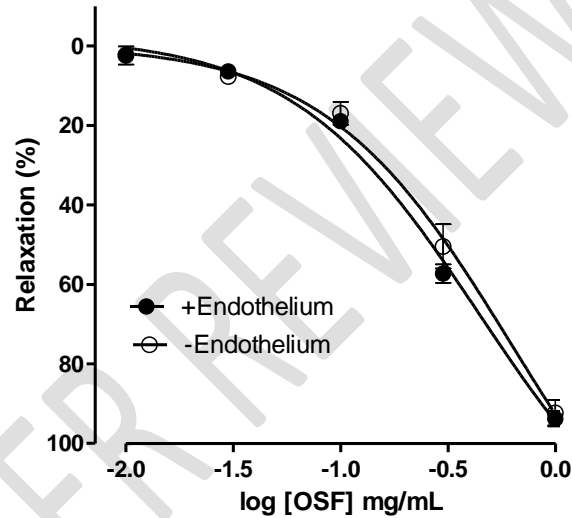
163 **Fig. 1. Cumulative concentration-response curves of OSF on endothelium-intact aortic**
 164 **rings pre-contracted with U46619. OSF effect (illustration, A) in the absence (control) or**
 165 **presence of L-NAME, Indomethacin (A), or both (B). Values are expressed as mean \pm SEM**
 166 **(n = 5-6). *** P < 0.001 vs. Control**

167

168 **3.2. Effect of anthocyanins-enriched extract of *Odontonema strictum* flowers on u46619-**
 169 **induced contraction of endothelium-intact or endothelium-denuded aortic rings**

170 The ability of OSF extract to relax vascular tone was assessed using mice artery rings contracted
 171 submaximally with thromboxane A₂ receptor agonist, U46619. We investigated the
 172 concentration-dependent vasorelaxant effect of OSF (10 - 1000 μ g/mL) on endothelium-intact

173 and endothelium-denuded aortic rings. The magnitude of endothelium (intact or denuded)
 174 relaxation is a function of OSF concentration (Fig. 2). However, the functional removal of
 175 endothelium did not modify OSF-induced relaxation in U46619-precontracted rat thoracic aorta
 176 rings. The pD2 of OSF in presence and in absence of endothelium are recorded in the table. The
 177 maximal relaxant effect was $93.78 \pm 4.69\%$ and $92.30 \pm 3.19\%$ for endothelium-intact and
 178 endothelium-denuded aortic rings, respectively.



179
 180 **Fig. 2. Vasorelaxant response induced by OSF on U46619-induced pre-contractions in**
 181 **endothelium-intact or -denuded rat aortic artery isolated rings. Values are expressed as**
 182 **mean \pm SEM (n = 5-6)**

183
 184 **Table: Vasorelaxant parameters of OSF (pD2) during exposure of mice aortic rings**
 185 **contracted with U46619 in the absence and presence of L-NAME and indomethacin**

Substance administered	pD2 (mg/mL)
U46619+OSF	$0.24 \pm 0.03^{***}$
L-NAME+U46619+OSF	$0.22 \pm 0.04^{***}$
Indomethacin+U46619+OSF	$0.21 \pm 0.04^{***}$
L-NAME+ Indomethacin+U46619+OSF	$0.19 \pm 0.05^{***}$
Denuded endothelium+U46619+OSF	$0.22 \pm 0.04^{***}$
Control (0.02% of DMSO)	0.0

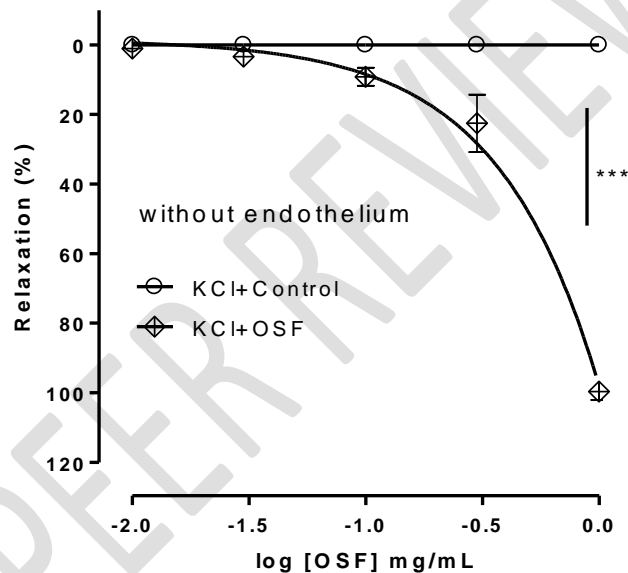
*** P < 0.001 vs. Control

186
 187
 188

189 **3.3. Effect of OSF on KCl (80 mM)-induced contraction of endothelium denuded mice**
190 **aortic rings**

191
192 We have also investigated the effect of OSF on aortic rings without endothelium precontracted by
193 depolarization with high concentration of K^+ . The results have shown that OSF had relaxed KCl (80
194 mM)-precontracted aortic rings in a concentration dependent manner (Fig. 3). The maximal
195 relaxant effect was $99.72 \pm 2.32\%$ and $pD_2 = 0.12 \pm 0.06$ mg/mL (Table).

196



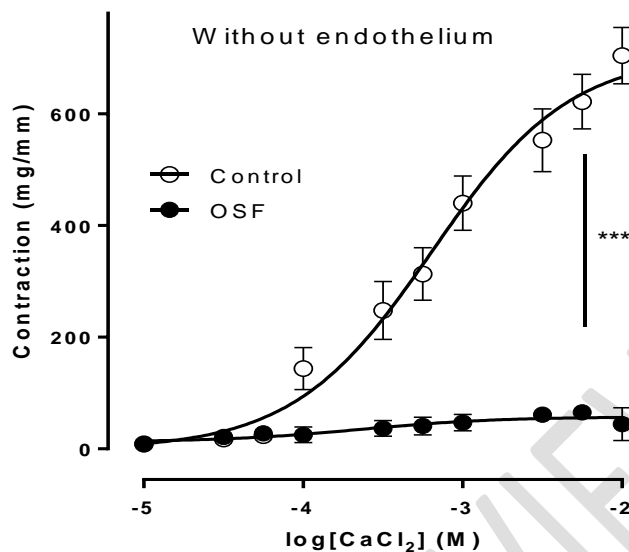
197

198 **Fig. 3. Concentration-response curves of OSF on KCl (80 mM) induced contraction in mice**
199 **endothelium denuded aortic artery rings. Values are expressed as mean \pm SEM (n = 4). *****
200 **P < 0.001 vs. Control**

201

202 **3.4. Effect of anthocyanins-enriched extract of *Odontonema Strictum* flowers on**
203 **extracellular Ca^{2+} -induced contraction**

204 To investigate the role of extracellular Ca^{2+} influx, cumulative addition of $CaCl_2$ (10^{-5} - 10^{-2} M)
205 in Ca^{2+} -free PSS medium containing KCl (80 mM) have been realized with endothelium-denuded
206 aortic rings. As compared to control (0.02% of DMSO), preincubation rings with OSF (400
207 μ g/mL) strongly inhibited Ca^{2+} -induced contraction in aorta rings ($p < 0.001$). The maximal
208 contraction induced by $CaCl_2$ (10^{-2} M) was 704.32 ± 50.43 mg/mm and 44.24 ± 29.46 mg/mm in
209 the absence and presence of OSF (400 μ g/mL), respectively (Fig. 4).

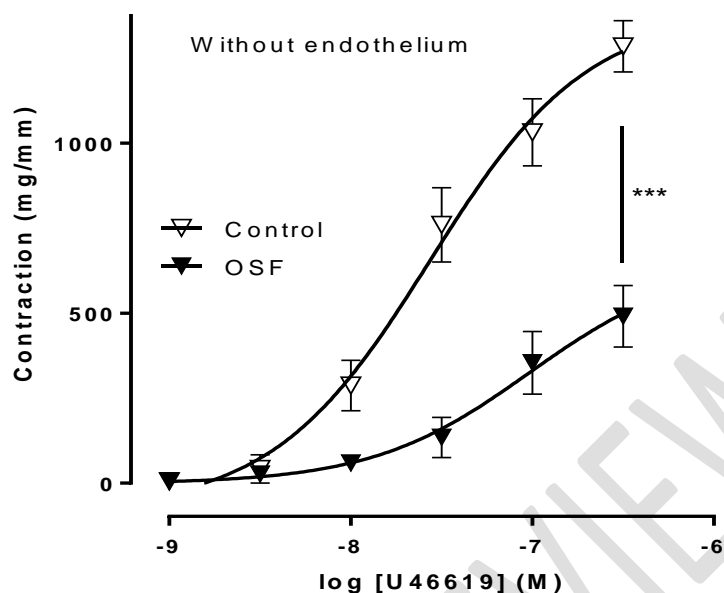


210
 211 **Fig. 4. Inhibitory effect of OSF on contraction induced by extracellular Ca²⁺ in endothelium**
 212 **denuded of mice thoracic aortic rings in PSS containing KCl 80 mM. Values are expressed**
 213 **as mean ± SEM (n = 5). *** p < 0.001 vs. control.**

214
 215 **3.5. Effects of anthocyanins-enriched extract of *Odontonema strictum* flowers on mice aortic**
 216 **rings by cumulative U46619**

217
 218 The preincubation of OSF (400 µg/mL) for 20 min was able to inhibit a concentration dependent
 219 manner the contractile response induced by U46619 on denuded-endothelium rings. The
 220 concentration response curves was significantly rightward with OSF inhibition effect (Fig. 5).
 221 The maximal contraction induced by U46619 (3.10⁻⁷ M) was 1284.34 ± 75.17 mg/mm and
 222 490.88 ± 90.53 mg/mm in the absence and presence of OSF (400 µg/mL), respectively.

223



224
 225 **Fig. 5. Inhibitory effect of OSF on contraction induced U46619 in endothelium-denuded of**
 226 **mice thoracic aortic rings in PSS. Values are expressed as mean \pm SEM (n = 5). *** p <**
 227 **0.001 vs. Control.**

228
 229 **4. DISCUSSION**

230
 231 Vascular smooth muscle vasoactivity is regulated by circulating factors from the endothelium,
 232 neurotransmitters and hormones [19]. The vasorelaxing effect is generally classified as
 233 endothelium-dependent or independent depending on the function of the endothelium.
 234 Endothelial cells secrete vasorelaxant substances such as nitric oxide (NO•), prostacyclin (PGI₂)
 235 and endothelium-derived hyperpolarizing factor (EDHF) to regulate vascular smooth muscle tone
 236 and endothelial-derived contracting factors such as endothelins, angiotensin II, prostanoids
 237 derived from cyclooxygenase and superoxide anions [18, 20]. The present study showed that the
 238 anthocyanins-enriched extract (prepared from *Odontonema strictum* flowers (OSF) inhibited
 239 U46619, KCl, CaCl₂ contractile responses in mice aorta rings ; the response is dose dependente.
 240 Indeed, many studies have reported that plant extracts exert vasculo-protection via their ability to
 241 induce the relaxation of blood vessels by a mechanism dependent on NO• and PGI₂ release [21-
 242 23]. Thus, we aimed at checking the first hypothesis. The inhibitory effect of OSF on U46619-
 243 induced contraction was not affected either in the presence of L-NAME or indomethacin or in the
 244 combinaison of L-NAME plus Indomethacin. Relaxant effect of NO• is mainly due to an increase
 245 in cyclic guanosine monophosphate (cGMP) [24, 25]. Blocking this NO•/cGMP pathway with L-
 246 NAME and indomethacin as a nonselective inhibitor of COX had no effects on the vasorelaxant

247 effect of the extract on U46619-induced contractions. Thus, the relaxant effect of the OSF is
248 independent of production NO• or prostacyclin. OSF also decreased U46619-induced
249 contractions in endothelial denuded aortic rings, as it mediated the same relaxation at similar
250 concentration as in the intact-endothelium. These results suggest that the relaxant effect has been
251 exerted on the vascular smooth muscle cells and not a lesser involvement of EDHF. Interestingly,
252 the endothelium-independent mechanism of OSF could help alleviate high blood pressure
253 associated with endothelial dysfunction by acting directly on smooth muscle. Indeed, authors
254 have shown that plant extracts could act directly on the vascular smooth muscle to induce
255 vasodilation [20, 26, 27].

256 In addition, contraction and relaxation of vascular smooth muscle cells is regulated by Ca²⁺ entry
257 from the extracellular space through Receptor-Operative Ca²⁺ Channels (ROCCs) or Voltage-
258 Dependent Ca²⁺ Channels (VDCCs) in the cytoplasmic membrane, through Ca²⁺ release from
259 intracellular Ca²⁺ stores (sarcoplasmic reticulum) by activation of 1,4,5 triphosphate inositol (IP₃)
260 and ryanodine receptors (RyR), protein kinase C (PKC) activation, and a Ca²⁺ sensitization
261 mechanism [28]. Moreover, previous reports have shown that the smooth muscle cells
262 contraction elicited by KCl (high K⁺, >30 mM) mainly results from the influx of extracellular
263 Ca²⁺ induced by depolarization of the cells membrane and subsequent opening of the voltage-
264 dependent slow Ca²⁺ channels (VDCCs) [29]. We therefore assessed whether OSF extract could
265 inhibit calcium entry activated by the VDCCs. We looked at OSF effect on the contraction in
266 response to KCl depolarization. As a result, OSF relaxed vasoconstriction induced by KCl in
267 rings. We also demonstrated OSF caused vasorelaxation of KCl-induced contraction in mice
268 isolated aortic ring through possible inhibition of VDCCs. Interestingly, OSF inhibited also
269 dramatically the contraction of endothelium- denuded aortic rings induced by Ca²⁺
270 supplementation in in Ca²⁺ free - PSS containing KCl (80 mM). These results suggested that OSF
271 have blocked both ROCCs and VDCCs involved in the vasodilatation activity. Indeed, the influx
272 of extracellular Ca²⁺ is mainly regulated by receptor operated calcium channels (ROCCs) or
273 VDCCs [25, 29-31].

274 The thromboxane A₂ analogue agonist (U46619) acts by stimulating the production of
275 phospholipase C (PLC). Afterwards, PLC produce diacylglycerol (DG) and IP₃, and
276 subsequently DG activates the light chain of myosin through activation of protein kinase C
277 (PKC), and IP₃ induces Ca²⁺ release from the sarcoplasmic reticulum by opening IP₃ receptors

278 and by Ca^{2+} influx through ROCCs [3, 20, 21, 32, 33]. To verify the involvement of this
279 pathways in the OSF vasodilation effect, the rings were preincubated with OSF before the
280 cumulative of U46619 (10^{-9} - $3 \cdot 10^{-7}$ M). The results showed that OSF significantly reduces this
281 agonist-induced contraction in mice aorta. Thus, OSF effect could be due to the inhibited the IP_3
282 and/or ryanodine receptor-dependent release of intracellular Ca^{2+} . It can also reduce DG-PKC
283 dependent myosin light chain kinase activity, and/or block ROCCs to decrease intracellular Ca^{2+}
284 and relax the mice aorta as reported previously [20, 25, 31]. In order to better determine the
285 mechanisms of action on the muscle cell, the use of specific inhibitors would be necessary for
286 future investigations.

287 According to the published literature, the presence of phytochemical components in *Odontonema*
288 *strictum* flowers such as saponins of flavonoids, tannins, steroids and terpenoids could explain
289 the vasodilation effect of OSF [15]. Indeed, many authors have demonstrated that flavonoids
290 (saponins) [34], tannins [35], steroids and terpenoids [36] have endowed vasodilator property.
291 Furthermore, these phytochemical groups have antioxidant properties [9, 14, 37] that could
292 provide a vascular protection effect by neutralizing reactive oxygen species which are known to
293 have constrictive effects. Oxidative damage can cause endothelial cells injuries and deleterious
294 vasodilator effects. It has been shown that antioxidant compounds could modify molecular events
295 towards an improvement of the endothelium function, and therefore play an important role in the
296 prevention of CVD [38, 39].

297 **5. CONCLUSION**

298 The present study has provided a pharmacological evidence for the vasorelaxant activity of
299 *Odontonema strictum*. This effect is mediated by endothelium-independent pathways including
300 the blockade of extracellular calcium influx and intracellular calcium-release. Therefore this
301 study supports the evidence that extract of OSF could be valuable alternative for the treatment of
302 hypertension in Burkina Faso.

303 **CONSENT**

304 It is not applicable.

306

307

308 **ETHICAL APPROVAL**

309 The laboratory experimentation was carried out according to the experimental protocols approved
310 and validated by the MEPHATRA-PH/IRSS laboratories and meeting the international standards
311 in this field. The protocol was conducted in accordance with the institutional Ethics Committee
312 for Animals protection regulations (directive 2010/63/EU on protection of animals used for
313 scientific purposes). Ethical approval code: 2010/63/EU, Date of approval: 20 October 2010.

314

315 **COMPETING INTERESTS**

316 The authors declare that there is no conflict of interests to disclose regarding the publication of
317 this paper.

318

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