

Original Research Article

THE EFFECT OF AIR DRIED LEAF POWDER AND BURNT LEAF ASH OF DIFFERENT PLANTS ON ROOT-KNOT NEMATODE (*Meloidogyne* spp.) ON OKRA (*Abelmoschus esculentus* L. Moench)

ABSTRACT

The effect of air dried leaf powder and burnt leaf ash of different plants on the control of root-knot nematode (*Meloidogyne* spp.) on Okra was investigated in a pot experiment. The experiment was laid out in a Completely Randomized Design with nine treatments each replicated four times. The treatments included: burnt leaf ashes and air dried leaf powders of *Baphia nitida*, *Pentaclethra macrophylla* and *Delonix regia* respectively, a synthetic nematicide (Carbofuran 3G) and two control which included inoculated but untreated (control 1) and uninoculated and untreated (control 2) pots. The plants were inoculated with 1000 nematode eggs (*Meloidogyne* spp.) two weeks after germination. Three days after, the treatments were applied at the rate of 30 g each while the synthetic nematicide (Carbofuran 3G) at 3 g/per pot. Parameters recorded were: plant height, number of leaves, fresh and dry shoot weights, fresh root weight, number of galls and nematode eggs in roots, number of larva in soil, number and weight of pods per plant. Results obtained indicated significant differences among the treatments in most parameters recorded. Generally, results from air dried leaf powder of *Baphia nitida* and burnt leaf ash of *Delonix regia* were better and compared favorably with the nematicide treated plants.

Key words: Air dried leaf, Burnt leaf ash, Carbofuran, *Meloidogyne* spp., *Abelmoschus esculentus*

INTRODUCTION

Okra as a vegetable crop belongs to the genus *Abelmoschus*, Family *Malvaceae* and has two main species: *Abelmoschus esculentus* (L) Moench and *Abelmoschus caillei* (A. Chev) (Siemonsma, 1982). It is an annual crop mainly grown as a fruit and leafy vegetable in the tropics (Tamiyu *et al.*, 2012). It is widely distributed in the Tropical and Subtropical regions of Asia, Southern Europe, the Mediterranean countries and the Americas (Andres *et al.*, 2005). Nigeria is the second largest producer of Okra in the world after India and largest producer in Africa (5.8 million tonnes), followed by Cote d'Ivoire, Ghana and others (FAOSTAT, 2011). In Nigeria, it is widely distributed and consumed in either fresh or dried forms (Farinde *et al.*, 2007).

Okra is important constituent of most local dishes in West Africa. It is used as a soup thickener and may also be served with rice and other food types (Tamiyu *et al.*, 2012). In Nigeria, okra is grown across different ecological zones because it serves as a source of income to framers as well as a cheap source of protein, vitamins A, B complex and C and mineral (Ca, P, Fe and K) to many households (Adebisi *et al.*, 2007). It is a nutritious vegetable that contains 86.1% of water, 2.2% of protein, 0.2% of fat, 9.7% of carbohydrate, 1.0% of fiber and 0.8% of ash (Saifullah and Rabbani, 2009), hence it plays vital role in human diet. Okra seeds contain greenish yellow edible oil which is suitable for use as a bio-fuel (Farroq, *et al.*, 2010).

41 Okra is notorious for its susceptibility to root knot nematodes (Noling, 2012). *Meloidogyne* spp.
42 are responsible for about 70-90% yield losses in okra (Saufiuddin *et al.*, 2011). Almost all the
43 vegetables in tropical and warm temperate regions are severely attacked by plant-parasitic
44 nematodes particularly root-knot nematodes (*Meloidogyne* spp.) (Sikora and Fernandez, 2005).
45 *Meloidogyne* spp. cause wilting, chlorosis, stunted growth, formation of galls in roots often leads
46 to destruction of roots, poor growth, yield and crop failure when nematode population exceeds
47 economic threshold level (Fourie *et al.*, 2001; Sikora and Fernandez, 2005). The symptoms for
48 identifying disease caused by *Meloidogyne* spp. is the presence of galls, wilting, loss of vigor,
49 yellowing of leaves (Kayani *et al.*, 2012). They have caused up to 80% yield losses in heavily
50 infested soils (Kaskavalci, 2007). However the extent of damage is influenced by the cultivar,
51 nematode species, level of soil infestation and environment (Ononuju, 1999). It is therefore
52 necessary to control root-knot nematodes in order to avoid or minimize yield losses in okra.

53 Management of root-knot nematodes with synthetic nematodes can be very effective (Sikora and
54 Fernandez, 2005; Adegbite and Agbaje, 2007). However, there is public outcry against synthetic
55 nematicides due to their toxicity, persistence and hazards posed to non-target species and
56 agriculturists (Oka *et al.*, 2014). As a result there is a growing preference for plant products
57 which are less harmful, effective, easily degraded, pollution free, leave no harmful residues,
58 cheaper and not toxic to host plants and humans (Amadioha, 2003). These plants if explored for
59 nematicidal activity might be alternatives to synthetic nematicides (Siji *et al.*, 2010).

60 The objective of this study is to determine the effect of air dried leaf powder and burnt leaf ash
61 of *Baphia nitida*, *Pentaclethra macrophylla* and *Delonix regia* on the population of root-knot
62 nematode (*Meloidogyne* spp.) on the growth and yield of okra, gall formation and also to
63 compare the effect of these plant materials with the synthetic nematicide (Carbofuran 3G).

64 MATERIALS AND METHODS

65 LOCATION OF EXPERIMENTAL SITE

66 The experiment was carried out in the Department of Plant Health Management, Michael Okpara
67 University of Agriculture, Umudike, Abia State, Nigeria located on latitude 5⁰2¹N and longitude
68 7⁰33⁴E. It lies in the humid tropical rain forest zone with annual rainfall of 1916mm per annum,
69 altitude of 112m above sea level and relative humidity of 76% with temperature range of 19-
70 35⁰C (N.R.C.R.I, 2010).

71 EXPERIMENTAL DESIGN

72 The experiment was laid out in a Completely Randomized Design (CRD) on a platform in an
73 open field using plastic pots. Okra variety *Pusa suwaan* was planted. There were nine treatments
74 replicated four times including the control, giving a total of 36 pots.

75 EXPERIMENTAL MATERIALS

76 **Source of seed:** Seeds of okra variety *Pusa suwaan* were obtained from National Root Crop
77 Research Institute Umudike, Abia State, Nigeria.

78 **Treatments:** The materials used were: burnt leaf ash and air dried leaf powder of *Baphia nitida*,
79 *Pentaclethra macrophylla* and *Delonix regia* and a synthetic nematicide (Carbofuran 3G).

80 **Treatments' sources and preparation:** The plant materials were obtained within the school
81 premises while the synthetic nematicide (Carbofuran 3G) was obtained from a chemical store.

82 The plant materials were collected and washed under running tap to remove soil particles and
 83 reduce contamination by microorganisms. They were then spread on a clean platform for air
 84 drying. This was done to reduce the moisture content after which each material was divided into
 85 two parts: one part was ground into powder using an electric grinder while the other part was
 86 burnt to ash in a controlled environment and each part were put in a clean plastic container ready
 87 for application.

88 **Soil preparation and sterilization:** Top soil was collected and sifted to remove large soil
 89 particles, stones and plants debris. The soil was poured into a cut metal drum. It was moistened
 90 and a source of heat applied underneath. It was sterilized at 80⁰C for 20 minutes after which it
 91 was left to cool (Ononuju *et al.*, 2014).

92 **Extraction of nematode eggs from inoculum:** Eggs of root-knot nematodes were extracted
 93 from the heavily galled roots of *Basella alba* (Ceylon spinach). The gall roots were washed in
 94 tap water and cut into pieces of 2cm long and placed in a beaker containing 200mls of 0.5%
 95 NaOCl (Sodium hypochloride) solution and was shaken vigorously for about 5minutes (Hussey
 96 and Barker, 1973). This was done to prevent egg damage while the gelatinous matrix is being
 97 removed. The solution was poured over two nested sieves of 75µm mesh 25µm mesh to collect
 98 the eggs. Eggs in the 25µm mesh sieve were rinsed with cold water and washed into beaker. The
 99 cut roots in the original beaker were washed twice with water to obtain additional eggs. The
 100 number of eggs in 1ml of water was estimated by counting four samples of a milliliter each using
 101 Doncaster's counting dish under a stereomicroscope and the average taken (Hussey and Barker,
 102 1973).

103 **Sowing of seeds:** Okra seeds were sown at two seeds per hole in plastic pots with diameter
 104 22.5cm containing 6kg sandy loam soil mixture that has been sterilized. Two weeks after, the
 105 seedlings were thinned down to a healthy plant per pot.

106 **Inoculation of plants with nematode eggs:** Three weeks after emergence, the plants were
 107 inoculated with a calculated volume of 1000 eggs of *Meloidogyne* spp. extracted by Hussey and
 108 Barker (1973) method. This was applied near the base of plant by making holes around the Okra
 109 plants.

110 **Application of treatments:** Thirty (30g) each of the air dried leaf powder and burnt leaf ash of
 111 *B. nitida*, *P. macrophylla* and *D. regia* were applied evenly on the surface of the soil in each
 112 bucket according to the treatments and their replications three days after inoculation. A synthetic
 113 nematicide (Carbofuran 3G) at 3.0g ai per plant was applied. The controls include plants whose
 114 soils were inoculated but untreated (control 1) and un-inoculated and untreated (control 2)
 115 respectively.

116 The treatment combinations were as follows:

- 117 1. T1 Nematode + Burnt leaf ash of *Baphia nitida*
- 118 2. T2 Nematode + Air dried leaf ash of *Baphia nitida*
- 119 3. T3 Nematode + Burnt leaf ash of *Pentaclethra macrophylla*
- 120 4. T4 Nematode + Air dried leaf ash of *Pentaclethra macrophylla*
- 121 5. T5 Nematode + Burnt leaf ash of *Delonix regia*
- 122 6. T6 Nematode + Air dried leaf ash of *Delonix regia*
- 123 7. T7 Nematode + synthetic nematicide (Carbofuran 3G).
- 124 8. T8 Inoculated but untreated (Control 1)
- 125 9. T9 Un-inoculated and untreated (Control 2)

126 (NPK 15:15:15) was applied at the rate of 100kg/ha to give a rate of 0.4g per bucket. Water was
 127 applied to the plant as at when required.

128

129 DATA COLLECTION

130 Data collected at the end of the experiment after twelve weeks included:

- 131 • Plant height- measured using a meter rule in centimeter.
- 132 • Number of leaves- counted with hand (cm).
- 133 • Fresh and dry shoot and fresh root weights- measured using a digital laboratory weighing
 134 balance in grams (g).
- 135 • Number of eggs in root and larva in soil- extracted using the modified Baermann
 136 technique (Hooper, 1969), and counted using Domncaster's counting dish under an
 137 electronic stereomicroscope.
- 138 • Number of galls in roots- counted with hand.
- 139 • Number of pods- counted with hand.
- 140 • Weight of pods- measured using a digital laboratory weighing balance in grams (g).

141 STATISTICAL ANALYSIS

142 The data collected were subjected to Analysis of Variance (ANOVA) and means were separated
 143 using Least Significant Difference (LSD) at 5% probability level ($P<0.05$) by using computer
 144 software "Genstat Discovery Edition 4".

145 RESULTS AND DISCUSSION

146 RESULTS

147 The effect of treatments on Plant height and Number of leaves are shown in Table 1. On plant
 148 height, significant difference ($P<0.05$) was observed between control 2 (un-inoculated and
 149 untreated) (27.17cm) and all the treatments. There was also a significant difference between the
 150 plants treated with air dried leaf powder of *B. nitida* (18.77cm) and control 1 (6.65cm). On
 151 Number of leaves, there was no significant difference recorded among the treatments including
 152 the controls. Although the highest number of leaves (15.00) was recorded for the plants treated
 153 with burnt leaf ash of *B. nitida* while the lowest (6.00cm) was recorded for control 1 (inoculated
 154 but untreated).

155 **Table 1: Effect of treatments on the plant height and number of leaves of okra infested**
 156 **with root-knot nematode (*Meloidogyne* spp.).**

Treatments	Plant height(cm)	No. of leaves per plant
Burnt leaf ash of <i>Baphia nitida</i>	11.20	15.00
Air dried leaf powder of <i>Baphia nitida</i>	18.77	13.00
Burnt leaf ash of <i>Pentaclethra macrophylla</i>	12.62	8.00
Air dried leaf powder of <i>Pentaclethra macrophylla</i>	12.00	10.00
Burnt leaf ash of <i>Delonix regia</i>	11.07	13.00
Air dried leaf powder of <i>Delonix regia</i>	7.25	10.00
Synthetic nematicide (Carbofuran 3G)	10.25	12.00

Inoculated but untreated (control 1)	6.65	6.00
Un-inoculated and untreated (control 2)	27.17	11.00
LSD _(0.05)	6.25	NS

157 Where, NS = No Significant Difference

158

159 Table 2 shows the effect of treatments on number and weight of pods. There was no significant
 160 difference ($P < 0.05$) between the treatments and the controls on the number of pods. Although the
 161 highest number of pods was recorded by control 2 (11) and plants treated with synthetic
 162 nematicide (11) respectively, with the least number of pods (5.0) recorded for plants that were
 163 inoculated but untreated (control 1). Significant differences ($P < 0.05$) were recorded on the
 164 weight of pods. All the treatments differed significantly from control 2 (34.9g), except the plants
 165 treated with synthetic nematicide (28.3g). Nevertheless plants treated with air dried leaf powder
 166 of *B. nitida* (23.4g) compared favorably with the synthetic nematicide.

167 **Table 2: Effect of treatments on the number and weight of Okra pods.**

Treatments	No. of pods per plant	Weight of pods (g)
Burnt leaf ash of <i>Baphia nitida</i>	7.0	21.3
Air dried leaf powder of <i>Baphia nitida</i>	10.0	23.4
Burnt leaf ash of <i>Pentaclethra macrophylla</i>	7.0	15.6
Air dried leaf powder of <i>Pentaclethra macrophylla</i>	8.0	21.4
Burnt leaf ash of <i>Delonix regia</i>	10.0	22.0
Air dried leaf powder of <i>Delonix regia</i>	6.0	16.1
Synthetic nematicide (Carbofuran 3G)	11.0	28.3
Inoculated but untreated (control 1)	5.0	6.1
Un-inoculated and untreated (control 2)	11.0	34.9
LSD _(0.05)	NS	9.75

168 Where, NS = No Significant Difference

169 The effect of treatments on fresh shoot, dry shoot and fresh root weights (Table 3). On fresh
 170 shoot weight, air dried leaf powder of *B. nitida* (480g) compared favourably with control 1
 171 (205g) but did not with control 2 and other treatments. On dry shoot weight, there was no
 172 significant difference recorded among the treatments including the controls. Although the highest
 173 dry shoot weight (85.2g) was recorded for the plants treated with burnt leaf ash of *P.*
 174 *macrophylla* while the lowest (30g) was recorded for the inoculated but untreated plants (control
 175 1). On fresh root weight all the treatments significantly differed from control 2 (371g). However,
 176 the other treatments did not differ significantly from each other.

177 **Table 3: Effect of treatments on fresh shoot, dry shoot and fresh root weights of Okra**
 178 **infested with root-knot nematode (*Meloidogyne* spp.)**

Treatments	Fresh shoot weight (g)	Dry shoot weight (g)	Fresh root weight (g)
Burnt leaf ash of <i>Baphia nitida</i>	133.0	32.8	66
Air dried leaf powder of <i>Baphia nitida</i>	480.0	76.8	187
Burnt leaf ash of <i>Pentaclethra macrophylla</i>	335.0	85.2	118
Air dried leaf powder of <i>Pentaclethra</i>	326.0	46.2	57

macrophylla

Burnt leaf ash of <i>Delonix regia</i>	308.0	64.0	136
Air dried leaf powder of <i>Delonix regia</i>	389.0	63.5	165
Synthetic nematicide (Carbofuran 3G)	362.0	70.0	132
Inoculated but untreated (control 1)	205.0	30.0	122
Un-inoculated and untreated (control 2)	629.0	68.2	371
LSD _(0.05)	267.1	NS	133.2

179 Where, NS = No Significant Difference

180 Effect of treatments on the number of nematode eggs in roots, galls in roots and larva in soil are
 181 shown in Table 4. On number of galls in roots, all the treatments compared favourably with
 182 control 1, but the treatments did not differ significantly from each other. Similar observations
 183 were made on the number of eggs in root. On number of larva in soil, the treatments and the
 184 controls did not differ significantly from each other. However apart from control 2 (000), plants
 185 treated with nematicide had the least number of larva (126) while the highest number (475) was
 186 recorded for plants treated with burnt leaf ash of *P. macrophylla*.

187 **Table 4: Effect of treatments on the population of *Meloidogyne* spp. in root and soil.**

Treatments	No. of galls in roots	No. of eggs in roots	No. of larva in soil
Burnt leaf ash of <i>Baphia nitida</i>	0.75	126.00	350
Air dried leaf ash of <i>Baphia nitida</i>	0.25	102.00	250
Burnt leaf ash of <i>Pentaclethra macrophylla</i>	0.50	102.00	475
Air dried leaf ash of <i>Pentaclethra macrophylla</i>	1.75	202.00	351
Burnt leaf ash of <i>Delonix regia</i>	1.00	158.00	425
Air dried leaf ash of <i>Delonix regia</i>	0.75	126.00	300
Synthetic nematicide (Carbofuran 3G)	0.00	78.00	126
Inoculated but untreated (control 1)	5.00	575.00	425
Un-inoculated and untreated (control 2)	0.00	000.00	000
LSD _(0.05)	2.23	272.5	NS

188 Where, NS = No Significant Difference

189 DISCUSSION

190 The results on the effect of the treatments on plant height agreed with the findings of Bawa *et al.*,
 191 (2014) in which the plant extracts used had a significant effect on the height of tomato. They are
 192 still in agreement with the findings of Kayani *et al.*, (2012) who confirmed that the symptoms for
 193 identifying diseases caused by *Meloidogyne* spp. is the presence of galls, wilting, loss of vigor,
 194 yellowing of leaves. Also Perry *et al.*, (2009) confirmed that the symptoms also include,
 195 yellowing of leaves and stunting, loss of vigor, wilting due to lack of moisture, decay of tissue
 196 due to secondary infection and yield loss. The growth reduction in crops such as Okra due to
 197 nematode varies depending on population density of nematode, level of cultivar susceptibility
 198 and environmental condition to which the host is subjected to (Ononuju and Fawole, 2000).

199 The reduction in the fresh shoot and root weight could be due to slow rate of action of the active
 200 ingredients in the plant materials (Barman and Das, 1996), or due to poor nutrients and water
 201 flow as a result of formed giant galls on the plant roots (Ploeg, 2001) which inhibit the normal
 202 growths and development of shoots and roots of plants affected.

203 The reduced number of nematode population and number of gall compared to control 2 agrees
 204 with the observations of Adegbite and Adesiyani (2001) who indicated that the higher the
 205 nematode population the higher the number of galls and the higher the number of damage done
 206 to plant which hinders the transportation of water and mineral nutrients from roots to aerial part
 207 of the plant. The results obtained confirmed the findings of Ozores-Hampton (2002) that the use
 208 of organic amendments suppressed soil phyto-parasitic nematode populations. Abulusoro *et al.*,
 209 (2004) reported that the susceptibility of tomato plants infected with root-knot nematodes
 210 (*Meloidogyne* spp.) show stunted growth, yield loss and conspicuous root galls, but a number of
 211 plants are thought to contain biologically active ingredients which when applied in the soil
 212 reduced the incidence of plant-parasitic nematodes. Sikora and Fernandez (2005) also found that
 213 application of sesame seed extract reduced the incidence of root-knot nematodes and the severity
 214 of galling on okra roots. According to Akhtar and Mahmood (1993), sesame seed extracts have a
 215 systemic activity against nematodes which may have accounted for the lower number of galls
 216 and mean population in treated plants. Also Vats *et al.*, (1996) reported reduction of galls and
 217 egg masses when some *M. javanica* infected tomato plants were treated with leaf extracts of
 218 *Azadirachta indica* and *Eucalyptus tereticornis*.

219 The reduced number and weight of pods agrees with Bolles *et al.*, (2012) who reported that
 220 nematode (*Meloidogyne* spp.) are found to be serious pest of Okra, damaging stands and greatly
 221 decreasing and delaying the production of pods. Also, Adesiyani, (1990) reported that some crop
 222 varieties could be effective host of root-knot nematode that suffer no statistically significant
 223 damage. However the extent of damage is influenced by the cultivar, nematode species, level of
 224 soil infestation and environment (Ononuju, 1999). According to Gommers (1981) active
 225 compounds with nematicide activity which have been found to limit the activities of plant-
 226 parasitic nematodes in plants includes alkaloids, diterpenes, fatty acids, glucosinolates,
 227 isothiocyanates, phenols, polyacetylenes, sesquiterpenes and thienyls. These compounds were
 228 extracted from different plants part such as roots, leaves and so on which could be responsible
 229 for the positive effects of air dried leaf ash and burnt leaf ash of these plants on the population of
 230 nematode in the soil and root as well as in pod formation.

231 CONCLUSION AND RECOMMENDATION

232 The different treatments significantly decreased the population of root-knot nematode
 233 (*Meloidogyne* spp.). There were significant differences between the controls and the treatments
 234 in some of the parameters measured, while there were no significant differences in others.
 235 Synthetic nematicide (Carbofuran 3G) appeared to be more effective in the control of root-knot
 236 nematode. However, the performance of the synthetic nematicide was not significantly different
 237 from the results obtained from air dried powder of *Baphia nitida* and burnt leaf ash of *Delonix*
 238 *regia* at the rate applied. From the findings of this study, air dried powder of *B. nitida* and burnt
 239 leaf ash of *D. regia* appeared to have significantly performed better than other treatments and is
 240 being recommended as an alternative for the synthetic nematicide (Carbofuran 3G) in the control
 241 of root-knot nematode. Notwithstanding, further research work both in green house and field
 242 trials at different rates are needed in order to authenticate this findings.

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