

4 **Effect of ethephon and storage temperature on physico-chemical changes during**
5 **ripening of mango (*Mangifera indica* L.) Cv. Neelum**

6 **Abstract**

7 Mango fruits Cv. Neelum treated with ethephon of different concentrations i.e. 250,
8 500, 750 and 1000 ppm for five minutes at different temperature conditions viz., 16, 20, 24
9 and 28 °C with 80% RH ripened in ripening chamber and untreated fruits kept at ambient
10 temperature (30-34 °C). Then the mango fruits were analyzed for physico-chemical changes
11 and sensory qualities. It was found that mango Cv. Neelum ethephon dip treatment placed in
12 the ripening chamber triggered the ripening process and showed that the significant
13 increasing trends in L*, a*, b* values of colour, TSS (°Brix), PLW (%), reducing sugars
14 (%), pH and decreasing trends in firmness (N), acidity during ripening in all the treatment
15 combinations during advancement of storage period in ripening chamber. It was observed
16 that mango fruits Cv. Neelum ripened by ethephon dip treatment of 750 ppm for 5 minutes at
17 28 °C, 80% RH showed better results in respect of a high overall acceptability score of 8.66.

18
19 **Keywords:** Mango, Ethephon, Ripening, Physico-chemical changes

20
21 **Introduction**

22
23 Mango (*Mangifera indica* L.) belongs to the family *Anacardiaceae*, is considered as
24 one of the choicest fruits of the world because of its attractive colour, delicious taste, and
25 excellent nutritional value. Mango is the most important tropical fruit in India accounting for
26 2.29 million ha of an area with a production of 15.19 million tons and a productivity of 6.63 t
27 ha⁻¹. India is estimated to account for about 40 per cent of the world's mango production.
28 Mango is one of the most extensively exploited fruit because of its flavour, fragrance and
29 juice content. It is a good source of fiber, sugars, vitamins, and minerals along with anti-
30 oxidants.

31 Fruit ripening is a genetically programmed stage of development overlapping with
32 senescence (Watada *et al.*, 1984). Mango fruits ripen unevenly on the tree and the natural
33 ripening process can be very slow and unpredictable. To overcome this problem, fruits can be
34 ripened artificially by exposing the fruits to certain chemicals, which initiate the ripening
35 process. (Campbell and Malo, 1969) found that the ripening of mature-green mangos was
36 accelerated in response to ethylene released from 2- chloroethylphosphonic acid (Ethephon).
37 Ethephon/ethrel is an ethylene-releasing chemical, which can be used to improve fruit colour
38 development and stimulates ripening process of the fruit.

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40 However, ripening of fruit generally is not completed on the tree because natural
41 ripening is a long process. Ethylene is one of the natural ripening hormones which is
42 responsible for accelerating the normal process of maturation, senescence and ripening. The
43 use of ethylene gas helps in achieving faster and more uniform ripening of fruits (Kader,
44 2002). Ethylene gas is commercially applied in the form of liquid i.e. Ethephon. In the case of
45 ethephon, the ripening is slightly cumbersome, the fruit sellers have to either dip the fruits in
46 a solution or pass through fumes of this chemical (Siddiqui and Dhua, 2009; Kulkarni *et al.*,
47 2004). In alkaline medium, ethylene is evolved from ethephon (Thompson and Seymour,
48 1982). Mohamed and Abu Goukh (2003) reported that ethylene released from ethephon was
49 more effective in triggering ripening of fruits. The ripening of green mangoes was accelerated
50 by 2-chloroethyl phosphonic acid (ethephon). Ethephon releases ethylene gas, which
51 naturally facilitates the ripening of fruits without any harmful effects (Sudhakar, 2006).

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52
53 The ripening with ethephon treatment seems to hold promise to get good and uniform
54 quality. In this technique, the fruits are dipped in different concentrations and placed in a
55 ripening chamber for 24 h to induce the fruits to ripen. The most important thing in this
56 technique is temperature and relative humidity control inside the ripening chamber which
57 should range between 16-25 °C and 90-95% RH, depending upon the fruit type (Mahajan and
58 Ghuman, 2010). And also, the ripening technology can be used at wholesale markets before
59 distribution to retailers. Therefore, the present investigation was carried out to study the
60 effect of different concentrations of ethephon at different temperatures on physico-chemical
61 changes during ripening behaviour of mango Cv. Neelum.

62 63 **Materials and Methods**

64 The present research entitled “Effect of ethylene on physico-chemical changes during
65 ripening of mango (*Mangifera indica* L.) Cv. Neelum.” was carried out in the Process
66 Engineering Laboratory, Department of Processing and Food Engineering, Dr. NTR College
67 of Agricultural Engineering, Bapatla during the year 2018 - 2019. Physiologically mature,
68 healthy green fruits at an optimum maturity of mango Cv. Neelum was collected from
69 Agricultural Research Farm, Bapatla. Manually harvested fruits were washed with
70 chlorinated water for further investigation. The ripening chamber made up of polyurethane
71 foam panels having a thickness of 60 mm with a dimension of 3.6 x 3.0 x 3.6 m having the
72 capacity to hold 3500-4000 kg of mango fruits.

73

74 The selected mango fruits Cv. Neelum was kept in the ripening chamber at Dr. NTR
75 College of Agricultural Engineering, Bapatla. For the preparation of 250 ppm, 500 ppm, 750
76 ppm and 1000 ppm of ethephon solutions, 0.64 ml, 1.28 ml, 1.92 ml and 2.56 ml of ethrel (2,
77 Chloro ethyl phosphonic acid 39% aqueous solution under the brand name ethefol)
78 respectively were dissolved in 1litre of distilled water. Uniform sized fruits were dipped in
79 ethephon solution for 5 min and air-dried to remove excess moisture. Fruits treated with
80 different concentrations such as 250, 500, 750 and 1000 ppm at different temperatures of 16,
81 20, 24 and 28 °C with relative humidity 80% in ripening chamber and control sample under
82 ambient conditions (30 °C, 55-65% RH). Further, physico-chemical analysis of mango fruits
83 was recorded at every two days interval.

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84
85 **Experimental design?, treatment design? how will**
86 **the results be tested?**

87
88 The physiological loss in weight (PLW) after each interval of storage was calculated
89 by subtracting the final weight from the initial weight of the fruits and ex-pressed in per cent.
90 The fruit firmness was measured with the help of a fruit penetrometer (Model FT- 327, USA)
91 using an 8 mm stainless steel probe. Total soluble solids (TSS) was determined by Hand
92 refractometer and expressed in °brix, the acidity of fruits by AOAC method (Anon, 1985),
93 reducing and acidity of fruits were recorded by a method as suggested by Ranganna (1997).
94 Data were recorded after 2, 4, 6 and 8 days of storage. Specific attributes and acceptability
95 were evaluated by the six untrained panelists based on acceptability, using the 9-point
96 hedonic scale with a panel of ten judges.

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97 98 **Results and Discussion**

99 **Weight loss**

100 It was observed from Fig. 1 that the physiological loss in weight was significantly
101 increased with the increase of ethephon concentrations. The maximum weight loss (19.0%)
102 was observed in T15 whereas it was only 10.5% in the control sample on the 11th day of
103 storage. Similarly, the loss in weight of fruit during storage both at ambient and in the
104 ripening chamber increased with the enhancement of storage days and was also observed in
105 pear (Dhillon and Mahajan, 2011) and papaya (Singh *et al.*, 2012). Continuous processes of

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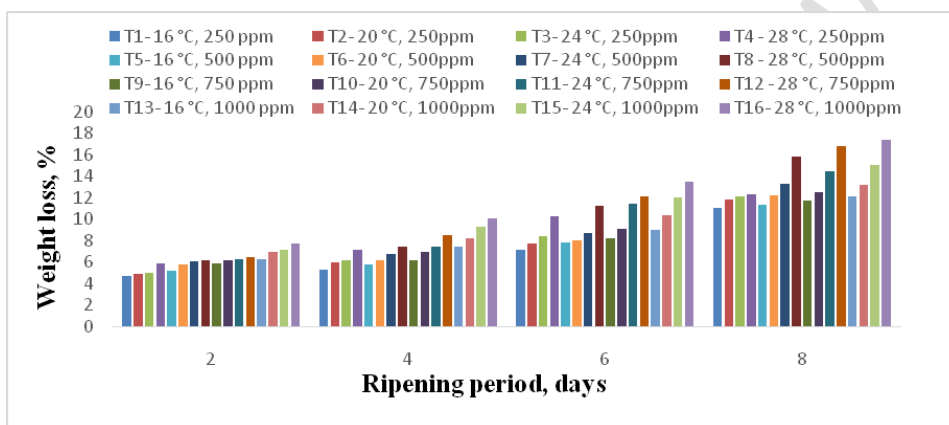
106 respiration and transpiration have resulted in weight loss. Mangoes kept at higher
 107 temperatures showed significantly greater weight loss than those at lower temperatures. A
 108 significant difference was also found in mangoes due to the interaction of ethephon and
 109 temperature. These differences indicated that the difference in total weight loss between
 110 ethephon treated and control sample increased with increased temperature. The higher weight
 111 loss at higher temperatures could be related to the higher evapo-transpiration rate and
 112 respiration rate at the higher temperatures as previously reported by Lebibet *et al.* (1995).
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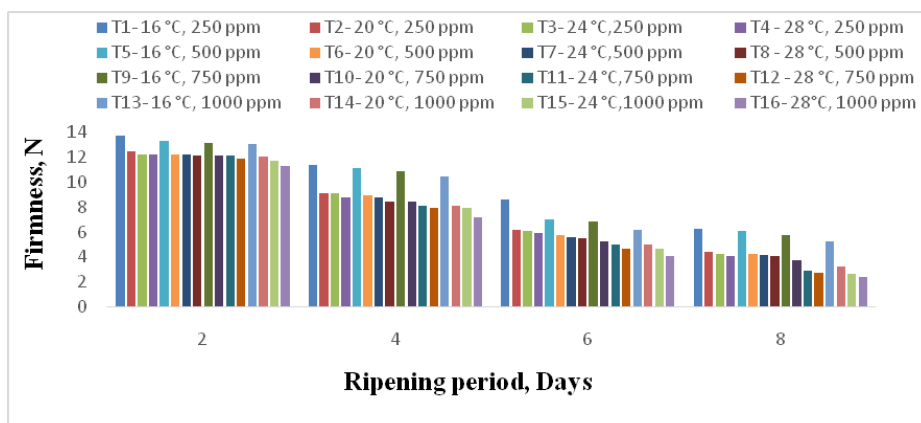


114
 115 **Figure 1. Effect of different concentrations of ethephon at different temperatures on**
 116 **weight loss during ripening of mango Cv. Neelum**

117 **Firmness**

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 119 It was observed from Fig.2 fruit firmness decreases with an increase in temperature
 120 and concentration. At the initial stage, the firmness of mango fruit was found to be 15.83 N.
 121 Fruit firmness decreased with an increase in ethephon concentration and also decreased with
 122 days of storage. The maximum fruit firmness was observed in the control sample and
 123 minimum in treatment, T15 ethephon.

124
 125 The decrease in firmness, during ripening, may be due to the breakdown of insoluble
 126 protopectin into soluble pectin or by cellular disintegration leading to membrane permeability
 127 (Brinston *et al.*, 1988). The interaction between temperature and ethephon in firmness
 128 showed that differences in firmness between ethephon treated and control mango fruits were
 129 less at higher temperatures than at lower temperatures. The reason could be starch hydrolysis
 130 or conversion of sugars.

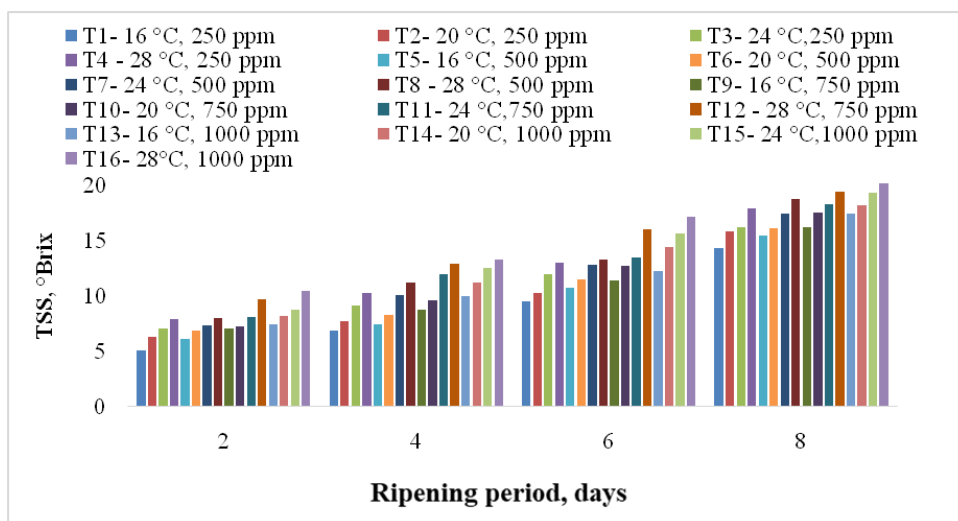


131
132 **Figure 2. Effect of different concentrations of ethephon at different temperatures on**
133 **firmness during ripening of mango Cv. Neelum**

134 **Total Soluble Solids**

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136
137 It was observed from Fig. 3 that the total soluble solids increased with an increase in
138 ethephon concentrations and increased with days of storage. The TSS might be increased due
139 to hydrolysis of starch, cellulose and pectin substances into simpler substances or might be
140 due to a decrease in moisture content. Similar results were also reported by Sakhale et al.,
141 (2006).

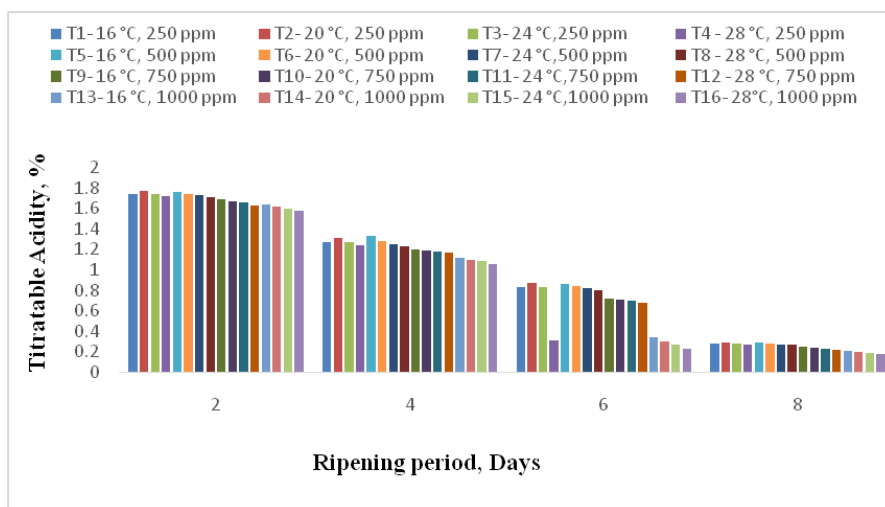
142
143 Ethephon treated mangoes possessed significantly greater TSS than the control
144 sample. Mangoes kept at higher temperatures showed greater TSS than those at lower
145 temperatures. The increase in TSS was the outcome of the conversion of carbohydrates into
146 simple sugars through a complex mechanism during the storage and the conversion rate was
147 increased with the increase in temperature. This conversion is also considered to be one of the
148 important indexes of the ripening process in mango and other climacteric fruit (Doreyappi-
149 Gowda and Huddar, 2001; Kays, 1991; Kittur et al., 2001).



150
 151 **Figure 3. Effect of different concentrations of ethephon at different temperatures on**
 152 **total soluble solids during ripening of mango Cv. Neelum**

153
 154 **Titrateable acidity**

155
 156 It is observed from the Fig.4 that acidity of the mango fruit was decreased by post
 157 harvest application of ethrel and the response varied within the concentrations. A maximum
 158 decrease (0.18%) in total acidity was found in T16 on 8th day of the storage treatment. The
 159 result indicates that the acidity content declined significantly with the increase in days of
 160 storage. Riberau-Gayon (1968) suggested that the transformation of organic acids into sugars
 161 may be the reason for decreasing organic acids during fruit ripening. The titrateable acidity
 162 was also decreased with the increase in storage temperature. The decrease in acidity was
 163 attributed to the conversion of citric acid into sugars and their further utilization in the
 164 metabolic process of the fruit (Doreyappy-Gowda and Huddar, 2001; Mizrach *et al.*, 1997;
 165 Rathore *et al.*, 2007; Srinivasa *et al.*, 2002).

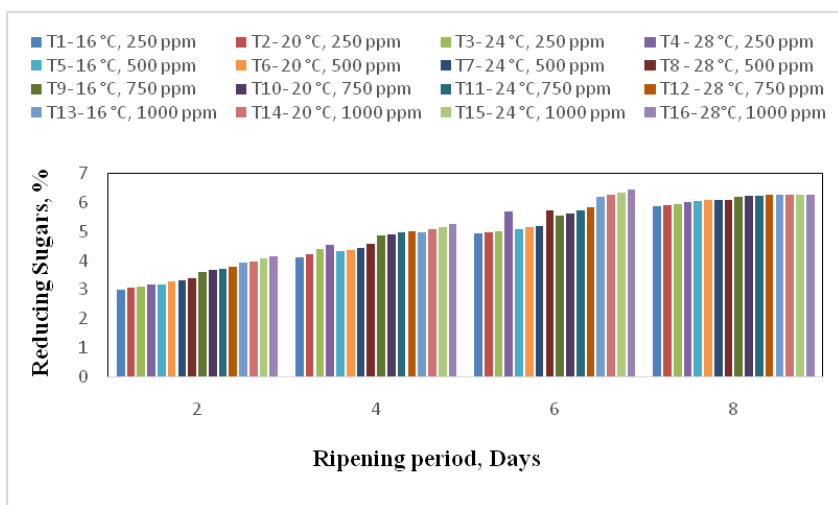


166
 167 **Figure 4. Effect of different concentrations of ethephon at different temperatures on**
 168 **titrateable acidity during ripening of mango Cv. Neelum**

169 **Reducing sugars**

170
 171 The results revealed that the sugar content increased with increasing ethephon
 172 concentration and storage temperatures as shown in Fig. 5. The maximum reducing sugar
 173 content (6.26%) was observed in T16 on 8th day of storage. The values of sugar content
 174 increased up to 8 days. In the present experiment, ethephon enhanced the rate of
 175 accumulation of reducing sugars in mango fruits. Similarly, a high percentage of sugar with
 176 ethephon application was observed by Singh *et al.* (2012) in mango. It was due to the faster
 177 rate of respiration and the formation of sugar content with the oxidation of carbohydrates.

178
 179 From the Figure, it was also observed that the reducing sugar content was lowest for
 180 samples stored at 16 °C and highest for samples stored at 28 °C. The increasing trend may be
 181 the polysaccharides were converted into soluble sugar through a hydrolytic conversion
 182 process, which was sensitive to temperature and/or to sunlight exposure for climacteric fruits
 183 during the ripening process (Campestre *et al.*, 2002; Kays, 1991; Martinez *et al.*, 1997).



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Figure 5. Effect of different concentrations of ethephon at different temperatures on reducing sugars during ripening of mango Cv. Neelum

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Ascorbic acid

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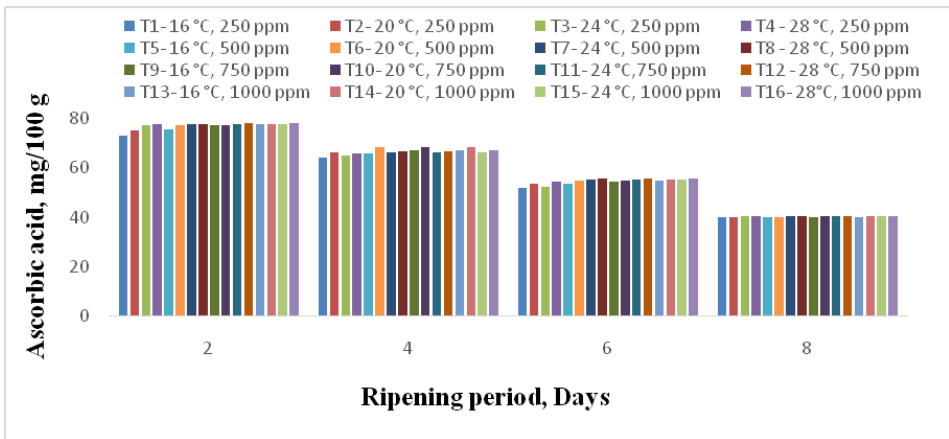
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It was observed from Fig. 6 that the ascorbic acid increased with an increase in ethephon concentrations and decreased with days of storage. The minimum value of ascorbic acid content (40.08 mg/100 g) was observed in T1 on 8th day of storage. The values of ascorbic acid content decreased up to 8 days. A decline in the ascorbic acid content of the mango fruits might be due to the utilization of ascorbic acid in the respiration process during ripening at ambient conditions. A similar trend was also observed by Sakhale *et al.*, (2006), William *et al.*, (2009), Pandarinathan and Sivakumar (2010), in mango fruits.

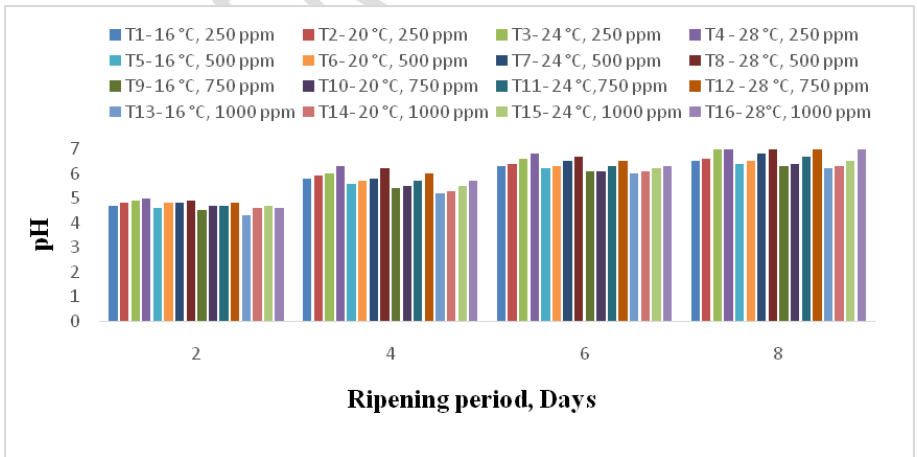
Further, it is also observed that the ascorbic acid was decreased with the ripening of the fruit or with the increase in storage temperature. This trend was due to conversion of acid into sugars and their further utilization in metabolic process of the fruit and that the chemical and biological process was increased with the increase in storage temperature (Doreyappu-Gowda and Huddar, 2001; Mizrach *et al.*, 1997; Rathore *et al.*, 2007; Srinivasa *et al.*, 2002).



202
 203 **Figure 6. Effect of different concentrations of ethephon at different temperatures on**
 204 **ascorbic acid during ripening of mango Cv. Neelum**

205 **pH**

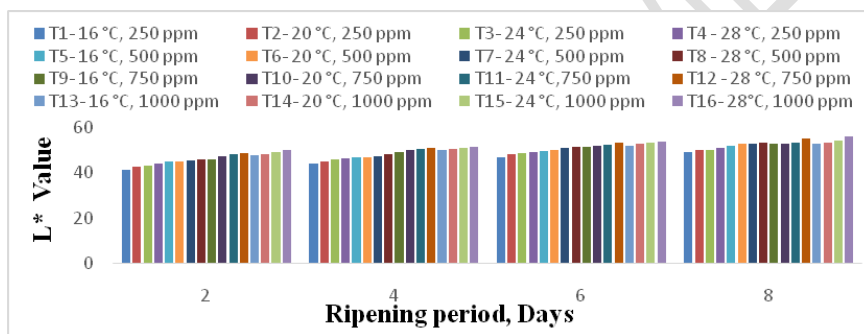
206 The results revealed that the pH increased during ripening and decreased with an
 207 increase in ethephon concentrations. as shown in Fig.7. The pH value of the control sample is
 208 4.1. The pH value of mango Cv. Neelum fruits are in the range of 4.3-7.3. Also, results
 209 indicated that the pH increased with an increase in storage temperatures during the ripening
 210 of mango fruits. These observations were attributed to the conversion of citric acid and
 211 ascorbic acid into sugar and other products with the ripening process and whose rate of
 212 conversion was increased with the temperature (Absar et al., 1993; Kumar and Singh, 1993;
 213 Rathore *et al.*, 2007; Yuniarti, 1980).



214
 215 **Figure 7. Effect of different concentrations of ethephon at different temperatures on pH**
 216 **during ripening of mango Cv. Neelum**

217 **L* value for colour (Lightness)**

218
 219 It was observed from Fig. 8 that the L* value for colour (Lightness) increases with
 220 increase in ethephon concentration as well as storage temperature during the enhancement of
 221 storage period in mango fruits. At the initial stage, the L* value of mango fruit was found to
 222 be 36.43. At the end of the 2nd, 4th, 6th and 8th day of storage, treatment T15 recorded the
 223 highest L* value (51.47, 52.72, 56.04). This is because of the fact that ethylene effects on the
 224 tissue and degreening of fruits. As the ethylene or ethephon triggered the ripening process,
 225 there is a rapid change in the colour from dark (green) to lightness (yellow) and it increased
 226 during the period of storage. Similar findings were reported by Daware (2012), Deepa and
 227 Preetha (2014) and Gill *et al.* (2015) in Dusehari mango fruits. Mangoes ripened at higher
 228 temperatures were significantly less green than those at lower temperatures.

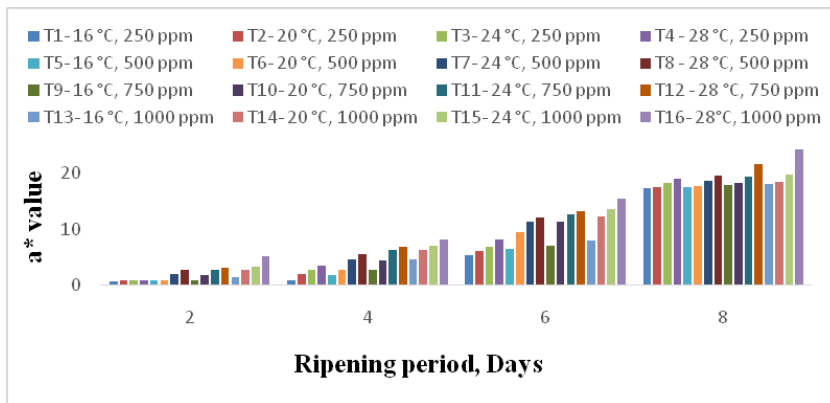


229
 230 **Figure 8. Effect of different concentrations of ethephon at different temperatures on L***
 231 **during ripening of mango Cv. Neelum**

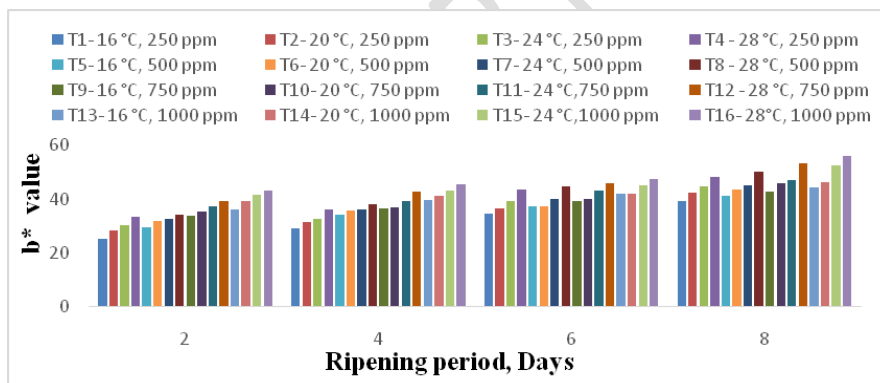
232 **a* and b* value (Redness/Greenness and Yellowness)**

233
 234 The results revealed that the a* and b* value increased during ripening with the
 235 increase in ethephon concentrations and storage temperatures as shown in Fig. 9 & 10. At the
 236 initial stage, a* and b* value of mango fruit was found to be -11.12 and 25.51. It was also
 237 observed that a* values for colour were less in untreated fruits in all days. This might be due
 238 to the breakdown of chlorophyll leading to the disappearance of green colour. During
 239 ripening, the peel colour changed from dark green to bright yellow and this is due to the
 240 change in chlorophyll which gradually unmasked the carotenoid pigments present in unripe
 241 mango fruits. As the ethylene triggered the ripening process, there was a rapid change in the
 242 colour from dark (green) to redness and it increased during the period of storage. Similarly,
 243 the b* values for colour were lower in untreated fruits i.e. control sample at all days. This
 244 might be due to changes during ripening period (loss of greenness, increase in redness and

245 yellowness) occurred as a result of the breakdown of the chlorophyll in the peel. Similar
 246 findings were also reported by Daware (2012), Deepa and Preetha (2014), Gill *et al.* (2015) in
 247 mango fruit.



248
 249 **Figure 9. Effect of different concentrations of ethephon at different temperatures on a***
 250 **value during ripening of mango Cv. Neelum**



252
 253 **Figure 10. Effect of different concentrations of ethephon at different temperatures on**
 254 **b* value during ripening of mango Cv. Neelum**

255 **Organoleptic evaluation at ripening**

256 The data (scores) on the organoleptic evaluation of Neelum mango fruits are
 257 presented in Table 1. It could be revealed from the data that fruits ripened at 28 °C
 258 259 presented in Table 1. It could be revealed from the data that fruits ripened at 28 °C
 260 temperature, 80-90% RH (T12) recorded maximum (8.50) sensory score in overall
 261 acceptability but were at par with those stored at 24 °C (T15). The mango fruits ripened at
 262 ambient temperature (control) recorded the lowest 6.25 as overall acceptability.

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Table 1 Effect of various treatments on organoleptic evaluation during ripening of mango Cv. Neelum fruits

Treatments	Colour	Flavour	Taste	Texture	Overall acceptability
Control	6.00	7.00	6.00	6.00	6.25
T1	7.50	7.50	7.25	8.00	7.56
T2	7.50	7.50	8.00	8.00	7.75
T3	8.00	7.50	7.50	8.00	7.75
T4	8.00	8.00	7.50	8.00	7.88
T5	7.50	8.00	7.50	8.00	7.75
T6	7.50	7.50	7.50	8.00	7.63
T7	8.25	8.00	7.50	7.65	7.85
T8	8.25	8.00	8.25	8.00	8.13
T9	7.50	8.25	7.50	8.00	7.81
T10	7.50	8.00	7.50	8.25	7.81
T11	8.45	8.00	8.00	8.00	8.11
T12	8.50	8.75	8.50	8.25	8.50
T13	7.65	8.45	8.00	8.00	8.03
T14	7.65	8.25	8.00	8.00	7.98
T15	8.50	8.00	8.50	8.25	8.31
T16	8.50	8.25	8.00	8.25	8.25

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Conclusion

It was found that mango Cv. Neelum ethephon dip treatment triggered the ripening process. It was also noticed that more the concentration of ethephon, faster was the ripening process and showed the significant increasing trends in L*, a*, b* values of colour and PLW (%) and decreasing trends in firmness (N) in all the treatments during advancement of the storage period in ripening chamber. It was observed that mango fruits Cv. Neelum ripened in ripening chamber in 8 days whereas fruits ripened at ambient conditions in 12 days. Similarly, mango fruits Cv. Neelum ripened by ethephon dip treatment of 750 ppm at 28 °C, 80% RH for 5 minutes showed better results in respect of high overall acceptability score of 8.50.

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