

Original Research Article

THE EFFECT OF AIR DRIED LEAF POWDER AND BURNT LEAF ASH OF DIFFERENT PLANTS ON ROOT-KNOT NEMATODE (*Meloidogyne* spp.) ON OKRA (*Abelmoschus esculentus* L. Moench)

ABSTRACT

The effect of air dried leaf powder and burnt leaf ash of different plants on the control of root-knot nematode (*Meloidogyne* spp.) on Okra was investigated in a pot experiment. The experiment was laid out in a Completely Randomized Design with nine treatments each replicated four times. The treatments included: burnt leaf ashes and air dried leaf powders of *Baphia nitida*, *Pentaclethra macrophylla* and *Delonix regia* respectively, a synthetic nematicide (Carbofuran 3G) and two control which included inoculated but untreated (control 1) and uninoculated and untreated (control 2) pots. The plants were inoculated with 1000 nematode eggs (*Meloidogyne* spp.) two weeks after germination. Three days after, the treatments were applied at the rate of 30 g each while the synthetic nematicide (Carbofuran 3G) at 3 g/per pot. Parameters recorded were: plant height, number of leaves, fresh and dry shoot weights, fresh root weight, number of galls and nematode eggs in roots, number of larva in soil, number and weight of pods per plant. Results obtained indicated significant differences among the treatments in most parameters recorded. Generally, results from air dried leaf powder of *Baphia nitida* and burnt leaf ash of *Delonix regia* were better and compared favorably with the nematicide treated plants.

Key words: Air dried leaf, Burnt leaf ash, Carbofuran, *Meloidogyne* spp., *Abelmoschus esculentus*

INTRODUCTION

Okra as a vegetable crop belongs to the genus *Abelmoschus*, Family *Malvaceae* and has two main species: *Abelmoschus esculentus* (L) Moench and *Abelmoschus caillei* (A. Chev) (Siemonsma, 1982). It is an annual crop mainly grown as a fruit and leafy vegetable in the tropics (Tamiyu *et al.*, 2012). It is widely distributed in the Tropical and Subtropical regions of Asia, Southern Europe, the Mediterranean countries and the Americas (Andres *et al.*, 2005). Nigeria is the second largest producer of Okra in the world after India and largest producer in Africa (5.8 million tonnes), followed by Cote d'Ivoire, Ghana and others (FAOSTAT, 2011). In Nigeria, it is widely distributed and consumed in either fresh or dried forms (Farinde *et al.*, 2007).

Okra is important constituent of most local dishes in West Africa. It is used as a soup thickener and may also be served with rice and other food types (Tamiyu *et al.*, 2012). In Nigeria, okra is grown across different ecological zones because it serves as a source of income to framers as well as a cheap source of protein, vitamins A, B complex and C and mineral (Ca, P, Fe and K) to many households (Adebisi *et al.*, 2007). It is a nutritious vegetable that contains 86.1% of water, 2.2% of protein, 0.2% of fat, 9.7% of carbohydrates, 1.0% of fiber and 0.8% of ash (Saifullah and Rabbani, 2009), hence it plays vital role in human diet. Okra seeds contain greenish yellow edible oil which is also suitable for use as a bio-fuel (Farroq, *et al.*, 2010).

Okra is notorious for its susceptibility to root knot nematodes (Noling, 2012). *Meloidogyne* spp. are responsible for about 70-90% of the yield losses in okra (Saufiuddin *et al.*, 2011). Almost all the vegetables in tropical and warm temperate regions are severely attacked by plant-parasitic nematodes particularly root-knot nematodes (*Meloidogyne* spp.) (Sikora and Fernandez, 2005). *Meloidogyne* spp. cause wilting, chlorosis, stunted growth, formation of galls in roots often leads to destruction of roots, poor growth, yield and crop failure when nematode population exceeds economic threshold level (Fourie *et al.*, 2001; Sikora and Fernandez, 2005). The symptoms for identifying disease caused by *Meloidogyne* spp. is the presence of galls, wilting, loss of vigor, yellowing of leaves (Kayani *et al.*, 2012). They have caused up to 80% yield losses in heavily infested soils (Kaskavanci, 2007). However the extent of damage is influenced by the cultivar, nematode species, level of soil infestation and environment (Ononuju, 1999). It is therefore necessary to control root-knot nematodes in order to avoid or minimize yield losses in okra.

Management of root-knot nematodes with synthetic nematodes can be very effective (Sikora and Fernandez, 2005; Adegbite and Agbaje, 2007). However, there is public outcry against synthetic nematicides due to their toxicity, persistence and hazards posed to non-target species and agriculturists (Oka *et al.*, 2014). As a result there is a growing preference for plant products which are less harmful, effective, easily degraded, pollution free, leave no harmful residues, cheaper and not toxic to host plants and humans (Amadioha, 2003). These plants if explored for nematicidal activity might be alternatives to synthetic nematicides (Siji *et al.*, 2010). [Have these products been safety tested? Plants also make toxic products and burning can create more. This should be addressed, either way!](#)

The objective of this study is to determine the effect of air dried leaf powder and burnt leaf ash of *Baphia nitida*, *Pentaclethra macrophylla* and *Delonix regia* on the population of root-knot nematode (*Meloidogyne* spp.) on the growth and yield of okra, gall formation and also to compare the effect of these plant materials with the synthetic nematicide (Carbofuran 3G).

MATERIALS AND METHODS

LOCATION OF EXPERIMENTAL SITE

The experiment was carried out in the Department of Plant Health Management, Michael Okpara University of Agriculture, Umudike, Abia State, Nigeria located on latitude 5⁰2¹N and longitude 7⁰33¹E. It lies in the humid tropical rain forest zone with annual rainfall of 1916 mm per annum, altitude of 112m above sea level and relative humidity of 76% with temperature range of 19-35⁰C (N.R.C.R.I, 2010).

EXPERIMENTAL DESIGN

The experiment was laid out in a Completely Randomized Design (CRD) on a platform in an open field using plastic pots. Okra variety *Pusa suwaan* was planted. There were nine treatments replicated four times including the control, giving a total of 36 pots.

EXPERIMENTAL MATERIALS

Source of seed: Seeds of okra variety *Pusa suwaan* were obtained from National Root Crop Research Institute Umudike, Abia State, Nigeria.

Treatments: The materials used were: burnt leaf ash and air dried leaf powder of *Baphia nitida*, *Pentaclethra macrophylla* and *Delonix regia* and a synthetic nematicide (Carbofuran 3G).

82 | **Treatments' sources and preparation:** The plant materials? Not plants?, leaves? or roots ?
 83 | were obtained within the school premises while the synthetic nematicide (Carbofuran 3G) was
 84 | obtained from a chemical store. The plant materials were collected and washed under running tap
 85 | to remove soil particles and reduce contamination by microorganisms. They were then spread on
 86 | a clean platform for air drying. This was done to reduce the moisture content after which each
 87 | material was divided into two parts: one part was ground into powder using an electric grinder
 88 | while the other part was burnt to ash in a controlled environment and each part were put in a
 89 | clean plastic container ready for application.

90 | **Soil preparation and sterilization:** Top soil was collected and sifted to remove large soil
 91 | particles, stones and plants debris. The soil was poured into a cut metal drum. It was moistened
 92 | and a source of heat applied underneath. It was sterilized at 80⁰C for 20 minutes after which it
 93 | was left to cool (Ononuju *et al.*, 2014).

94 | **Extraction of nematode eggs from inoculum:** Eggs of root-knot nematodes were extracted
 95 | from the heavily galled roots of *Basella alba* (Ceylon spinach). The galled roots were washed in
 96 | tap water and cut into pieces of 2cm long and placed in a beaker containing 200_mls of 0.5%
 97 | NaOCl (Sodium hypochloride) solution and ~~was~~ shaken vigorously for about 5minutes (Hussey
 98 | and Barker, 1973). This was done to prevent egg damage while the gelatinous matrix is being
 99 | removed. The solution was poured over two nested sieves of 75µm mesh 25µm mesh to collect
 100 | the eggs. Eggs in the 25µm mesh sieve were rinsed with cold water and washed into a beaker.
 101 | The cut roots in the original beaker were washed twice with water to obtain additional eggs. The
 102 | number of eggs in 1ml of water was estimated by counting four samples of a milliliter each using
 103 | Doncaster's counting dish under a stereomicroscope and the average taken (Hussey and Barker,
 104 | 1973).

105 | **Sowing of seeds:** Okra seeds were sown at two seeds per hole in plastic pots with diameter
 106 | 22.5cm containing 6kg sandy loam soil mixture that has been sterilized. Two weeks after, the
 107 | seedlings were thinned down to a healthy plant per pot.

108 | **Inoculation of plants with nematode eggs:** Three weeks after emergence, the plants were
 109 | inoculated with a calculated volume of 1000 eggs of *Meloidogyne* spp. ~~extracted by Hussey and~~
 110 | ~~Barker (1973) method.~~ This was Eggs were applied near the base of plant by making holes
 111 | around the Okra plants.

112 | **Application of treatments:** Thirty (30g) each of the air dried leaf powder and burnt leaf ash of
 113 | *B. nitida*, *P. macrophylla* and *D. regia* were applied evenly on the surface of the soil in each
 114 | bucket according to the treatments and their replications three days after inoculation. A synthetic
 115 | nematicide (Carbofuran 3G) at 3.0g ai per plant was applied. The controls include plants whose
 116 | soils were inoculated but untreated (control 1) and un-inoculated and untreated (control 2)
 117 | respectively.

118 | The treatment combinations were as follows:

- 119 | 1. T1 Nematode + Burnt leaf ash of *Baphia nitida*
- 120 | 2. T2 Nematode + Air dried leaf ash of *Baphia nitida*
- 121 | 3. T3 Nematode + Burnt leaf ash of *Pentaclethra macrophylla*
- 122 | 4. T4 Nematode + Air dried leaf ash of *Pentaclethra macrophylla*
- 123 | 5. T5 Nematode + Burnt leaf ash of *Delonix regia*
- 124 | 6. T6 Nematode + Air dried leaf ash of *Delonix regia*
- 125 | 7. T7 Nematode + synthetic nematicide (Carbofuran 3G).

- 126 8. T8 Inoculated but untreated (Control 1)
 127 9. T9 Un-inoculated and untreated (Control 2)

128 (NPK 15:15:15) was applied at the rate of 100kg/ha to give a rate of 0.4g per bucket. Water was
 129 applied to the plant as ~~at when~~ required.

130

131 DATA COLLECTION

132 Data collected at the end of the experiment after twelve weeks included:

- 133 • Plant height- measured using a meter rule in centimeter.
- 134 • Number of leaves-~~counted with hand (cm)~~.
- 135 • Fresh and dry shoot and fresh root weights- measured using a digital laboratory weighing
 136 balance in grams (g).
- 137 • Number of eggs in root and larva in soil- extracted using the modified Baermann
 138 technique (Hooper, 1969), and counted using Domncaster's counting dish under an
 139 electronic stereomicroscope.
- 140 • Number of galls in roots-~~counted with hand~~.
- 141 • Number of pods-~~counted with hand~~.
- 142 • Weight of pods- measured in grams (g) using a digital laboratory weighing balance-~~in~~
 143 grams (g).

144 STATISTICAL ANALYSIS

145 The data collected were subjected to Analysis of Variance (ANOVA) and means were separated
 146 using Least Significant Difference (LSD) at 5% probability level ($P < 0.05$) by using computer
 147 software "Genstat Discovery Edition 4".

148 RESULTS AND DISCUSSION

149 RESULTS

150 The effect of treatments on Plant height and Number of leaves are shown in Table 1. ~~On- For~~
 151 plant height, significant difference ($P < 0.05$) was observed between control 2 (un-inoculated and
 152 untreated) (27.17cm) and all the treatments. There was also a significant difference between the
 153 plants treated with air dried leaf powder of *B. nitida* (18.77cm) and control 1 (6.65cm). ~~On- For~~
 154 Number of leaves, there was no significant difference recorded among the treatments including
 155 the controls. Although, the highest number of leaves (15.00) was recorded for the plants treated
 156 with burnt leaf ash of *B. nitida* while the lowest (6.00cm) was recorded for control 1 (inoculated
 157 but untreated).

158 **Table 1: Effect of treatments on the plant height and number of leaves of okra infested**
 159 **with root-knot nematode (*Meloidogyne* spp.).**

Treatments	Plant height(cm)	No. of leaves per plant
Burnt leaf ash of <i>Baphia nitida</i>	11.20	15.00
Air dried leaf powder of <i>Baphia nitida</i>	18.77	13.00
Burnt leaf ash of <i>Pentaclethra macrophylla</i>	12.62	8.00

Air dried leaf powder of <i>Pentaclethra macrophylla</i>	12.00	10.00
Burnt leaf ash of <i>Delonix regia</i>	11.07	13.00
Air dried leaf powder of <i>Delonix regia</i>	7.25	10.00
Synthetic nematicide (Carbofuran 3G)	10.25	12.00
Inoculated but untreated (control 1)	6.65	6.00
Un-inoculated and untreated (control 2)	27.17	11.00
LSD _(0.05)	6.25	NS

160 Where, NS = No Significant Difference [show +/- values for each measure; provides idea of](#)
 161 [variability between samples](#)

162

163 Table 2 shows the effect of treatments on number and weight of pods. There was no significant
 164 difference ($P < 0.05$) between the treatments and the controls on the number of pods. Although the
 165 highest number of pods was recorded by control 2 (11) and plants treated with synthetic
 166 nematicide (11) respectively, with the least number of pods (5.0) recorded for plants that were
 167 inoculated but untreated (control 1). Significant differences ($P < 0.05$) were recorded on the
 168 weight of pods. All the treatments differed significantly from control 2 (34.9g), except the plants
 169 treated with synthetic nematicide (28.3g). Nevertheless plants treated with air dried leaf powder
 170 of *B. nitida* (23.4g) compared favorably with the synthetic nematicide.

171 **Table 2: Effect of treatments on the number and weight of Okra pods.**

Treatments	No. of pods per plant	Weight of pods (g)
Burnt leaf ash of <i>Baphia nitida</i>	7.0	21.3
Air dried leaf powder of <i>Baphia nitida</i>	10.0	23.4
Burnt leaf ash of <i>Pentaclethra macrophylla</i>	7.0	15.6
Air dried leaf powder of <i>Pentaclethra macrophylla</i>	8.0	21.4
Burnt leaf ash of <i>Delonix regia</i>	10.0	22.0
Air dried leaf powder of <i>Delonix regia</i>	6.0	16.1
Synthetic nematicide (Carbofuran 3G)	11.0	28.3
Inoculated but untreated (control 1)	5.0	6.1
Un-inoculated and untreated (control 2)	11.0	34.9
LSD _(0.05)	NS	9.75

172 Where, NS = No Significant Difference [Would Duncan's multiple range test be better?](#)

173 The effect of treatments on fresh shoot, dry shoot and fresh root weights [are shown in](#) (Table 3).
 174 On fresh shoot weight, air dried leaf powder of *B. nitida* (480g) compared favourably with
 175 control 1 (205g) but did not with control 2 and other treatments. On dry shoot weight, there was
 176 no significant difference recorded among the treatments including the controls. Although the
 177 highest dry shoot weight (85.2g) was recorded for the plants treated with burnt leaf ash of *P.*
 178 *macrophylla* while the lowest (30g) was recorded for the inoculated but untreated plants (control
 179 1). On fresh root weight all the treatments significantly differed from control 2 (371g). However,
 180 the other treatments did not differ significantly from each other.

181 **Table 3: Effect of treatments on fresh shoot, dry shoot and fresh root weights of Okra**
 182 **infested with root-knot nematode (*Meloidogyne* spp.)**

Treatments	Fresh shoot	Dry shoot	Fresh root
------------	-------------	-----------	------------

	weight (g)	weight (g)	weight (g)
Burnt leaf ash of <i>Baphia nitida</i>	133.0	32.8	66
Air dried leaf powder of <i>Baphia nitida</i>	480.0	76.8	187
Burnt leaf ash of <i>Pentaclethra macrophylla</i>	335.0	85.2	118
Air dried leaf powder of <i>Pentaclethra macrophylla</i>	326.0	46.2	57
Burnt leaf ash of <i>Delonix regia</i>	308.0	64.0	136
Air dried leaf powder of <i>Delonix regia</i>	389.0	63.5	165
Synthetic nematicide (Carbofuran 3G)	362.0	70.0	132
Inoculated but untreated (control 1)	205.0	30.0	122
Un-inoculated and untreated (control 2)	629.0	68.2	371
LSD _(0.05)	267.1	NS	133.2

183 Where, NS = No Significant Difference

184 Effect of treatments on the number of nematode eggs in roots, galls in roots and larva in soil are
 185 shown in Table 4. On number of galls in roots, all the treatments compared favourably with
 186 control 1, but the treatments did not differ significantly from each other. Similar observations
 187 were made on the number of eggs in root. On number of larva in soil, the treatments and the
 188 controls did not differ significantly from each other. However apart from control 2 (000), plants
 189 treated with nematicide had the least number of larva (126) while the highest number (475) was
 190 recorded for plants treated with burnt leaf ash of *P. macrophylla*.

191 **Table 4: Effect of treatments on the population of *Meloidogyne* spp. in root and soil.**

Treatments	No. of galls in roots	No. of eggs in roots	No. of larva in soil
Burnt leaf ash of <i>Baphia nitida</i>	0.75	126.00	350
Air dried leaf ash of <i>Baphia nitida</i>	0.25	102.00	250
Burnt leaf ash of <i>Pentaclethra macrophylla</i>	0.50	102.00	475
Air dried leaf ash of <i>Pentaclethra macrophylla</i>	1.75	202.00	351
Burnt leaf ash of <i>Delonix regia</i>	1.00	158.00	425
Air dried leaf ash of <i>Delonix regia</i>	0.75	126.00	300
Synthetic nematicide (Carbofuran 3G)	0.00	78.00	126
Inoculated but untreated (control 1)	5.00	575.00	425
Un-inoculated and untreated (control 2)	0.00	000.00	000
LSD _(0.05)	2.23	272.5	NS

192 Where, NS = No Significant Difference

193 DISCUSSION

194 The results on the effect of the treatments on plant height agreed with the findings of Bawa *et al.*,
 195 (2014) in which the plant extracts used had a significant effect on the height of tomato. They are
 196 still in agreement with the findings of Kayani *et al.*, (2012) who confirmed that the symptoms for
 197 identifying diseases caused by *Meloidogyne* spp. is the presence of galls, wilting, loss of vigor,
 198 yellowing of leaves. Also Perry *et al.*, (2009) confirmed that the symptoms also include,
 199 yellowing of leaves and stunting, loss of vigor, wilting due to lack of moisture, decay of tissue
 200 due to secondary infection and yield loss. The growth reduction in crops such as Okra due to
 201 | nematode [investigation](#) varies depending on population density of nematode, level of cultivar

202 susceptibility and environmental condition to which the host is subjected to (Ononuju and
203 Fawole, 2000).

204 The reduction in the fresh shoot and root weight could be due to slow rate of action of the active
205 ingredients in the plant materials (Barman and Das, 1996), or due to poor nutrients and water
206 flow as a result of ~~formed~~-giant galls ~~formed~~ on the plant roots (Ploeg, 2001) which inhibit the
207 normal growths and development of shoots and roots of plants affected.

208 The reduced number of nematode population and number of galls compared to control 2 agrees
209 with the observations of Adegbite and Adesiyani (2001) who indicated that the higher the
210 nematode population the higher the number of galls and the higher the ~~number-amount~~ of
211 damage done to plants which hinders the transportation of water and mineral nutrients from roots
212 to aerial part of the plant. The results obtained confirmed the findings of Ozores-Hampton (2002)
213 that the use of organic amendments suppressed soil phyto-parasitic nematode populations.
214 Abulusoro *et al.*, (2004) reported that ~~the susceptibility-susceptible~~of tomato plants infected with
215 root-knot nematodes (*Meloidogyne* spp.) show stunted growth, yield loss and conspicuous root
216 galls, but ~~that~~ a number of plants are thought to contain biologically active ingredients which
217 when applied in the soil reduced the incidence of plant-parasitic nematodes. Sikora and
218 Fernandez (2005) also found that application of sesame seed extract reduced the incidence of
219 root-knot nematodes and the severity of galling on okra roots. According to Akhtar and
220 Mahmood (1993), sesame seed extracts have a systemic activity against nematodes which may
221 have accounted for the lower number of galls and ~~lower~~ mean population in treated plants. Also
222 Vats *et al.*, (1996) reported reduction of galls and egg masses when some *M. javanica* infected
223 tomato plants were treated with leaf extracts of *Azadirachta indica* and *Eucalyptus tereticornis*.

224 The reduced number and weight of pods agrees with Bolles *et al.*, (2012) who reported that
225 nematode (*Meloidogyne* spp.) ~~are found to be serious pest of Okra, damaging stands and~~ greatly
226 ~~decreasing-decreased~~ and ~~delaying-delayed~~ the production of pods. Also, Adesiyani, (1990)
227 reported that some crop varieties could be effective hosts of root-knot nematode ~~that-but~~ suffer
228 no statistically significant damage. However the extent of damage is ~~generally~~ influenced by the
229 cultivar, nematode species, level of soil infestation and environment (Ononuju, 1999). According
230 to Gommers (1981) active compounds with nematicide activity which have been found to limit
231 the activities of plant-parasitic nematodes in plants includes alkaloids, diterpenes, fatty acids,
232 glucosinolates, isothiocyanates, phenols, polyacetylenes, sesquiterpenes and thienyls. These
233 compounds were extracted from different plants part such as roots, leaves and so on which could
234 be responsible for the positive effects of air dried leaf ash and burnt leaf ash of these plants on
235 the population of nematode in the soil and root as well as in pod formation.

236 CONCLUSION AND RECOMMENDATION

237 The different treatments significantly decreased the population of root-knot nematode
238 (*Meloidogyne* spp.). There were significant differences between the controls and the treatments
239 in some of the parameters measured, while there were no significant differences in others.
240 Synthetic nematicide (Carbofuran 3G) appeared to be more effective in the control of root-knot
241 nematode. However, the performance of the synthetic nematicide was not significantly different
242 from the results obtained from air dried powder of *Baphia nitida* and burnt leaf ash of *Delonix*
243 *regia* at the rate applied. From the findings of this study, air dried powder of *B. nitida* and burnt
244 leaf ash of *D. regia* appeared to have significantly performed better than other treatments and is
245 being recommended as an alternative for the synthetic nematicide (Carbofuran 3G) in the control
246 of root-knot nematode. Notwithstanding, further research work both in green house and field

247 | trials at different rates are needed in order to authenticate this findings. [Is there any practical](#)
 248 [way this could be developed for use in growers plots?](#)

249 **REFERENCES**

- 250 Abulusoro, S.A., Oyedunmade, E.A and Olabiyi, T.I. (2004). Screen House and Laboratory
 251 Assessment of Toxic Effect of Brimstone (*Morinda lucida*) leaf of the Rook-knot
 252 Nematode, *Meloidogyne incognita*. *Plant Pathology Journal* 2004; 1:45 – 49.
 253
- 254 Adebisi, M. A., Akintobi, D. C. and Oyekale, K. O. (2007). Preservation of Okra seed vigor by
 255 seed treatment and storage containers. *Nigerian Journal of Horticultural Science (NJHS)*,
 256 Vol. 12, 1-7.
 257
- 258 Adesiyon, S. O., Caveness, F. E., Adeniji, M. O. and Fawole, B. (1990). The Root-knot
 259 Nematodes, *Meloidogyne* spp.. In *Nematode Pests of Tropical Crops*. Heinemann Books
 260 (Nigeria) Ltd. Chapter 3 pp 19 -34
 261
- 262 Amadioha, A.C. (2003). Evaluation of some plant leaf extracts against *Colletotrichum*
 263 *lindemuthianum* in cowpea. *Acta Phytopathologica et Entomologica Hungarica* 3(3-4),
 264 259-265.
 265
- 266 Andres, C.D., Simandi, B., Orsi, F., Lambrou, C., Tatla, D.M and Panayiotou, C. (2005).
 267 Supercritical carbon dioxide extraction of okra (*Hibiscus esculentus* L.) seeds. *Journal of*
 268 *the Science of Food and Agriculture* 85, 1415–9.
 269
- 270 Asawalam, E. F. and S. O. Adesiyon. (2001). Comparison of Nematicidal Potentials of
 271 *Azadirachta indica* and Carbofuran (Furadan) on the growth and yield of root-knot
 272 nematode-infested okra. *Journal of Sustainable Agriculture and the Environment*, 3(1): 85-
 273 92.
 274
- 275 Akhtar, M. and Mahmood, I. (1993). Control of plant parasitic nematodes with “Nimin” and
 276 some plant oils by bare-root dip treatment. *Nematologia Mediterranean* 21, 89-92.
 277
- 278 Barman, M. and Das, P. (1996). Effect of chemical seed dressing and organic amendments alone
 279 and in combination for the management of root-knot nematode on green gram. *Indian J.*
 280 *Nematol.*, 26: 72-76.
 281
- 282 Bawa, J.A., Mohammed, I. and Liadi, S. (2014). Nematicidal effect of some plant extracts on
 283 root-knot nematodes (*Meloidogyne incognita*) of tomato (*Lycopersicon esculentum*). *World*
 284 *Journal Life Sci. and Medical Res.* 3(3):4-7.
 285
- 286 Bolles, B. and Johnson, L. (2012). Nematode damage of Okra. University of Florida. IFAS
 287 extension. <http://Washington.ifas.ufl.edu/> Retrieved 23/07/2013.
 288
- 289 Djian-Caporalino, C., Fazari, A., Arguel, M.J., Vernie T, VandeCastele, C., Faure, I., Brunoud,
 290 G., Pijarowski, L., Palloix, A and Lefebvre, V. (2007). Root-knot nematode (*Meloidogyne*
 291 spp..) resistant genes in pepper (*Capsicum annum* L.) are clustered on the P9
 292 chromosome. *Theoretical and Applied Genetics* 114: 473–486.
 293

- 294 Ekwurube, C. S., Osakwe, J. A., Chukwu, E.C. and Epidi, T.T. (2008). Effect of oil palm sludge
295 on cowpea nodulation and weed control in the humid forest zone of Nigeria. *African*
296 *Journal of Biotechnology* Vol. 7 (16), pp.2869-2873.
297
- 298 FAOSTAT. (2011). Food and Agricultural Organization of the United Nations: Okra production
299 statistic 2011. Retrieved on 31 January, 2014 from <http://faostat.fao.org/foostat/>
300
- 301 Farinde, A.J., Owolarafe, O.K and Ogungbemi, O.I (2007). “An overview of Production,
302 Processing, Marketing and Utilisation of Okra in Egbedore Local Government Area of
303 Osun State, Nigeria ” Agricultural Engineering International: Manuscript. No. MES 07 002
304 *the CIGR E-journal*, 9: 1-17.
305
- 306 Farrooq, A., Umer, R., Muhammad A. and Muhammad, N. (2010). Okra (*Hibiscus esculentus.*)
307 Seed Oil for biodiesel production. *Applied Energy* 87 (3): 779-785.
308
- 309 Fourie, H., McDonald, A.L and Loots, G.C (2001). Plant parasitic nematodes in field crops in
310 South Africa 6, Soybean. *Nematology.*, 3: 447-454.
311
- 312 Gommers, F. J. (1981). Biochemical interaction between nematodes and plants and their
313 relevance to control. *Helminthological Abstract Series B Plant Nematology* 50: 9-24.
314
- 315 Hooper, D. J. (1969). Extraction and handling of plant and soil nematode. *Nematodes of Tropical*
316 *Crops*, 21-22.
- 317 Hussey, R. S. and K. R. Barker (1973). A comparison of methods of collecting inoculum of
318 *Meloidogyne* spp. including a new technique. *Plant Disease Reporter*, 57: 1025-1028.
319
- 320 Iannotti, M. (2013). Okra – Growing Okra in the Backyard. Vegetable Garden.
321 <http://about.com/homegarden/Retrieved 23/07/2013>.
322
- 323 Inra, P. (2013). *Meloidogyne incognita* resources: <http://inra.fr/Retrieved 12/01/2014>.
324
- 325 Kaskavalci, G. (2007). Effect of soil solarization and organic amendment treatment for
326 controlling *Meloidogyne incognita* in tomato cultivars in Western Anatolia. *Turkish. J.*
327 *Agric.* 31:156-167.
328
- 329 Kayani, M. Z., Mukhtar, T. and Hussian, M. A. (2012). Association of Root-Knot Nematodes
330 (*Meloidogyne* spp.) with cucumber in the Pothowar region of the Punjab province of
331 Pakistan. *Int. J. Biotech*, a(1-2):23-29.
332
- 333 Ngele, K.K. (2010). Introduction to Entomology and pest management. Good Tidings Publishers
334 Limited, Abakaliki, Nigeria. pp.107-112.
335
- 336 Noling, J. W. (2012). *Nematode Management in Okra*. Entomology and Nematology
337 Department, Citrus Research Center, Cooperative Extension Service, Institute of Food and
338 Agricultural Sciences, University of Florida, Lake Alfred, FL. <http://ed.is.ifas.ufl.edu/>
339 Retrieved 23/07/2013.
340

- 341 N.R.C.R.I. (2010). *National Root Crops Research Institute*. NRCRI, Umudike, Meteorological
342 Station Data, 2010.
- 343
- 344 Oka, Y., Shuker, S., Tkachi, N., Trabelcy, B and Gerchman, Y. (2014). Nematicidal activity of
345 *Ochradenus baccatus* against the root-knot nematode, *Meloidogyne javanica*. *Plant*
346 *Pathology* 63(1), 221-231.
- 347
- 348 Ononuju, C. C. (1999). *Management of Plant Parasitic Nematode problems in commercial*
349 *plantain and banana productions*. A paper presented at the third Annual Conference of
350 Agricultural Society Institute Baddaggi, Nigeria October 18 1999.
- 351
- 352 Ononuju, C. C. and Fawole, B. (2000). Evaluation of Ethoprophos Isazophos, Carbofuran for the
353 control of plant parasitic nematodes and their effects on plantain growth and yield.
354 *Nigerian J. Sci.*, 34: 81-89.
- 355
- 356 Ononuju, C. C., Ikwunagu, E. A., Okorochoa, A. D. and Okorie, C. C. (2014). Effects of different
357 Agricultural wastes and botanical on root knot nematode (*Meloidogyne* spp) on Okra
358 (*Abelmoschus esculentus* L. Moench). *Journal of Entomology and Nematology*, 6 (5): 56-
359 61.
- 360
- 361 Ozores-Hampton, M. (2002): Organic materials in horticulture: An industry perspective. Intro.
362 *Hor. Tech.* 12(3): 8-9.
- 363
- 364 Perry, R. N., Moens, M. and Starr, J. L. (2009). Root-knot nematodes. Wallingford, UK, CAB
365 International.
- 366
- 367 Ploeg, A. (2001). When nematodes attack is important. *California Grower*. October. 12-13 p.
- 368
- 369 Saifullah, M and Rabbani, M. G. (2009). Evaluation and Characterization of Okra (*Abelmoschus*
370 *esculentus* (L) Moench). Genotypes. *J. Agric* 7:92-99.
- 371
- 372 Saufiuddin, S., Sheila, S. and Shweta, S. (2011). Pathogenicity of root-knot nematode,
373 *Meloidogyne incognita* and root-rot fungus, *Rhizoctonia solani* on okra (*Ablemoschus*
374 *esculentus* L.). *E-Journal of Science and Technology*, 3(6)
- 375
- 376 Shurtleff, M. C and Averre, C. W. (2000). *Diagnosing plant disease caused by plant parasitic*
377 *nematodes*. The American Phytopathological Society, pp. 187.
- 378
- 379 Siemonsma, J.S. (1982): The cultivation of Okra (*Abelmoschus* spp.), tropical fruit-vegetable
380 (with special reference to the Ivory Coast) D.H.O., thesis, Wageningen Agricultural,
381 Wageningen, the Netherland. 297pp
- 382
- 383 Sikora, R.A and Fernández, E. (2005). Nematode parasites of vegetables. In: Luc M, Sikora R.A,
384 Bridge J, eds. *Plant Parasitic Nematodes in Subtropical and Tropical Agriculture*. CAB
385 International, Wallingford, 319-390.
- 386
- 387 Siji, J.V., Jayaprakas., C.A., Sheila, M.S and C. Mohandas, C. (2010). Efficacy of *Cleome*
388 *viscosa* L. against *Meloidogyne incognita* infestation in okra (*Abelmoschus esculentus* L.)
389 *Thai Journal of Agricultural Science* 43 (3): 151-156.

- 390
391 Tamiyu, R.A., Ahmed, H.G and Muhammad A.S. (2012). Effect of sources of organic manure on
392 growth and yield of okra (*Abelmoschus esculentus* (L.) Moench) in Sokoto, Nigeria.
393 *Nigerian Journal of Basic and Applied Science* 20(3), 213-216.
394
- 395 Vats, R., Nandal, S.N and Dalal, M.R. (1996). Comparative efficacy of different preparations of
396 leaf extracts of *Azadirachta indica* and *Eucalyptus tereticornis* against *M. javanica* in
397 tomato nursery. *Haryana Agricultural University journal of Research*, 26(2): 99-102.

UNDER PEER REVIEW