

## Original Research Article

### THE EFFECT OF AIR DRIED LEAF POWDER AND BURNT LEAF ASH OF DIFFERENT PLANTS ON ROOT-KNOT NEMATODE (*Meloidogyne* spp.) ON OKRA (*Abelmoschus esculentus* L. Moench)

#### ABSTRACT

The effect of air dried leaf powder and burnt leaf ash of different plants on the control of root-knot nematode (*Meloidogyne* spp.) on Okra was investigated in a pot experiment. The experiment was laid out in a Completely Randomized Design with nine treatments each replicated four times. The treatments included: burnt leaf ashes and air dried leaf powders of *Baphia nitida*, *Pentaclethra macrophylla* and *Delonix regia*, respectively, a synthetic nematicide (Carbofuran 3G) and two control which included inoculated but untreated (control 1) and uninoculated and untreated (control 2) pots. The plants were inoculated with 1000 nematode eggs (*Meloidogyne* spp.), two weeks after germination. Three days after, the treatments were applied at the rate of 30 g each, while the synthetic nematicide (Carbofuran 3G) at 3 g/per pot. Parameters recorded were: plant height, number of leaves, fresh and dry shoot weights, fresh root weight, and number and weight of pods per plant as plant growth parameters. Number of galls, number of nematode eggs in roots, and number of larva juveniles in soil as nematode parameters. Results obtained indicated significant differences ( $P \leq 0.05$ ) among the treatments in most parameters recorded and compared to controls. Generally, results from air dried leaf powder of *Baphia nitida* and burnt leaf ash of *Delonix regia* were better and compared favorably with the nematicide treated plants.

**Key words:** Air dried leaf, Burnt leaf ash, Carbofuran, *Meloidogyne* spp., *Abelmoschus esculentus*

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#### INTRODUCTION

Okra as a vegetable crop belongs to the genus *Abelmoschus*, Family *Malvaceae* and has two main species: *Abelmoschus esculentus* (L) Moench and *Abelmoschus caillei* (A. Chev) (Siemonsma, 1982). It is an annual crop mainly grown as a fruit and leafy vegetable in the tropics (Tamiyu *et al.*, 2012). It is widely distributed in the Tropical and Subtropical Regions of Asia, Southern Europe, the Mediterranean countries and the Americas (Andres *et al.*, 2005). Nigeria is the second largest producer of Okra in the world after India and largest producer in Africa (5.8 million tonnes), followed by Cote d'Ivoire, Ghana and others (FAOSTAT, 2011). In Nigeria, it is widely distributed and consumed in either fresh or dried forms (Farinde *et al.*, 2007).

Okra is important constituent of most local dishes in West Africa. It is used as a soup thickener and may also be served with rice and other food types (Tamiyu *et al.*, 2012). In Nigeria, okra is grown across different ecological zones because it serves as a source of income to framers as well as a cheap source of protein, vitamins A, B complex and C and mineral (Ca, P, Fe and K) to many households (Adebisi *et al.*, 2007). It is a nutritious vegetable that contains 86.1% of water, 2.2% of protein, 0.2% of fat, 9.7% of carbohydrate, 1.0% of fiber and 0.8% of ash (Saifullah and

41 Rabbani, 2009), hence it plays vital role in human diet. Okra seeds contain greenish yellow  
42 edible oil which is suitable for use as a bio-fuel (Farroq, *et al.*, 2010).

43 Okra is notorious for its susceptibility to root knot nematodes (Noling, 2012). *Meloidogyne* spp.  
44 are responsible for about 70-90% yield losses in okra (Saufiuddin *et al.*, 2011). Almost all the  
45 vegetables in tropical and warm temperate regions are severely attacked by plant-parasitic  
46 nematodes, particularly root-knot nematodes (*Meloidogyne* spp.) (Sikora and Fernandez, 2005).  
47 *Meloidogyne* spp. cause wilting, chlorosis, stunted growth, formation of galls in roots often  
48 leading to destruction of roots, poor growth, yield and crop failure when nematode population  
49 exceeds economic threshold level (Fourie *et al.*, 2001; Sikora and Fernandez, 2005). The  
50 symptoms for identifying disease caused by *Meloidogyne* spp. is the presence of galls, wilting,  
51 loss of vigor, yellowing of leaves (Kayani *et al.*, 2012). They have caused up to 80% yield losses  
52 in heavily infested soils (Kaskavalci, 2007). However the extent of damage is influenced by the  
53 cultivar, nematode species, level of soil infestation and environment (Ononuju, 1999). It is  
54 therefore necessary to control root-knot nematodes in order to avoid or minimize yield losses in  
55 okra.

56 Management of root-knot nematodes with synthetic nematodes can be very effective (Sikora and  
57 Fernandez, 2005; Adegbite and Agbaje, 2007 not found in the list of references). However, there  
58 is public outcry against synthetic nematicides due to their toxicity, persistence and hazards posed  
59 to non-target species and agriculturists (Oka *et al.*, 2014). As a result, there is a growing  
60 preference for plant products which are less harmful, effective, easily degraded, pollution free,  
61 leave no harmful residues, cheaper and not toxic to host plants and humans (Amadioha, 2003).  
62 These plants if explored for nematicidal activity might be alternatives to synthetic nematicides  
63 (Siji *et al.*, 2010).

64 The objective of this study is to determine the effect of air dried leaf powder and burnt leaf ash  
65 of *Baphia nitida*, *Pentaclethra macrophylla* and *Delonix regia* on the population of root-knot  
66 nematode (*Meloidogyne* spp.) on the growth and yield of okra, gall formation and also to  
67 compare the effect of these plant materials with the synthetic nematicide (Carbofuran 3G).

## 68 MATERIALS AND METHODS

### 69 LOCATION OF EXPERIMENTAL SITE

70 The experiment was carried out in the Department of Plant Health Management, Michael Okpara  
71 University of Agriculture, Umudike, Abia State, Nigeria located on latitude 5<sup>0</sup>2<sup>1</sup>N and longitude  
72 7<sup>0</sup>33<sup>1</sup>E. It lies in the humid tropical rain forest zone with annual rainfall of 1916mm per annum,  
73 altitude of 112m above sea level and relative humidity of 76% with temperature range of 19-  
74 35<sup>0</sup>C (N.R.C.R.I, 2010).

### 75 EXPERIMENTAL DESIGN

76 The experiment was laid out in a Completely Randomized Design (CRD) on a platform in an  
77 open field using plastic pots. Okra variety *Pusa suwaan* was planted. There were nine treatments  
78 replicated four times including the control, giving a total of 36 pots.

### 79 EXPERIMENTAL MATERIALS

80 **Source of seed:** Seeds of okra variety *Pusa suwaan* were obtained from National Root Crop  
81 Research Institute Umudike, Abia State, Nigeria.

82 **Treatments:** The materials used were: burnt leaf ash and air dried leaf powder of *Baphia nitida*,  
83 *Pentaclethra macrophylla* and *Delonix regia* and a synthetic nematicide (Carbofuran 3G).

84 **Treatments' sources and preparation:** The plant materials were obtained within the school  
85 premises, while the synthetic nematicide (Carbofuran 3G) was obtained from a chemical store.  
86 The plant materials were collected and washed under running tap to remove soil particles and  
87 reduce contamination by microorganisms. They were then spread on a clean platform for air  
88 drying. This was done to reduce the moisture content after which, each material was divided into  
89 two parts: one part was ground into powder using an electric grinder, while the other parts were  
90 burnt to ash in a controlled environment and each part were put in a clean plastic container ready  
91 for application.

92 **Soil preparation and sterilization:** Top soil was collected and sifted to remove large soil  
93 particles, stones and plants debris. The soil was poured into a cut metal drum. It was moistened  
94 and as a source of heat applied underneath. It was sterilized at 80°C for 20 minutes after which, it  
95 was left to cool (Ononuju *et al.*, 2014).

96 **Extraction of nematode eggs from inoculum:** Eggs of root-knot nematodes were extracted  
97 from the heavily galled roots of *Basella alba* (Ceylon spinach). The galled roots were washed in  
98 tap water and cut into pieces of 2-cm long, and placed in a beaker containing 200mls of 0.5%  
99 NaOCl (Sodium hypochloride) solution and was shaken vigorously for about 5 minutes (Hussey  
100 and Barker, 1973). This was done to prevent egg damage, while the gelatinous matrix is being  
101 removed. The solution was poured over two nested sieves of 75µm mesh 25µm mesh to collect  
102 the eggs. Eggs in the 25µm mesh sieve were rinsed with cold water and washed into beaker. The  
103 cut roots in the original beaker were washed twice with water to obtain additional eggs. The  
104 number of eggs in 1ml of water was estimated by counting four samples of a milliliter each using  
105 Doncaster's counting dish under a stereomicroscope and the average was taken (Hussey and  
106 Barker, 1973).

107 **Sowing of seeds:** Okra seeds were sown at two seeds per hole in plastic pots with diameter 22.5-  
108 cm containing 6- kg sandy loam soil mixture that has been sterilized. Two weeks after, the  
109 seedlings were thinned down to a healthy plant per pot.

110 **Inoculation of plants with nematode eggs:** Three weeks after emergence, the plants were  
111 inoculated with a calculated volume of 1000 eggs of *Meloidogyne* spp. extracted by Hussey and  
112 Barker (1973) method. This was applied near the base of plant by making holes around the Okra  
113 plants.

114 **Application of treatments:** Thirty (30g) each of the air dried leaf powder and burnt leaf ash of  
115 *B. nitida*, *P. macrophylla* and *D. regia* were applied evenly on the surface of the soil in each  
116 bucket according to the treatments and their replications, three days after inoculation. A synthetic  
117 nematicide (Carbofuran 3G) at 3.0g ai per plant was applied. The controls include plants whose  
118 soils were inoculated but untreated (control 1) and un-inoculated and untreated (control 2)  
119 respectively.

120 The treatment combinations were as follows:

- 121 1. T1 Nematode + Burnt leaf ash of *Baphia nitida*  
 122 2. T2 Nematode + Air dried leaf ash of *Baphia nitida*  
 123 3. T3 Nematode + Burnt leaf ash of *Pentaclethra macrophylla*  
 124 4. T4 Nematode + Air dried leaf ash of *Pentaclethra macrophylla*  
 125 5. T5 Nematode + Burnt leaf ash of *Delonix regia*  
 126 6. T6 Nematode + Air dried leaf ash of *Delonix regia*  
 127 7. T7 Nematode + synthetic nematicide (Carbofuran 3G).  
 128 8. T8 Inoculated but untreated (Control 1)  
 129 9. T9 Un-inoculated and untreated (Control 2)

130 (NPK 15:15:15) was applied at the rate of 100kg/ha to give a rate of 0.4g per bucket. Water was  
 131 applied to the plant as at when required.

132

### 133 DATA COLLECTION

134 Data collected at the end of the experiment after twelve weeks included:

- 135 • Plant height- measured using a meter rule in centimeter.  
 136 • Number of leaves- counted ~~with hand~~ **visually**(cm).  
 137 • Fresh and dry shoot and fresh root weights- measured using a digital laboratory weighing  
 138 balance in grams (g).  
 139 • Number of eggs in root and ~~larva~~ **juveniles** in soil- extracted using the modified  
 140 Baermann technique (Hooper, 1969), and counted using Domncaster's counting dish  
 141 under an electronic stereomicroscope.  
 142 • Number of galls in roots- counted ~~with hand~~ **visually**.  
 143 • Number of pods- counted ~~with hand~~ **visually**.  
 144 •  
 145 • Weight of pods- measured using a digital laboratory weighing balance in grams (g).

### 146 STATISTICAL ANALYSIS

147 The data collected were subjected to Analysis of Variance (ANOVA) and means were separated  
 148 using Least Significant Difference (LSD) at 5% probability level ( $P < 0.05$ ) by using computer  
 149 software "Genstat Discovery Edition 4".

## 150 RESULTS AND DISCUSSION

### 151 RESULTS

152 The effect of treatments on **plant height** and **number of leaves** are shown in Table 1. **As for** plant  
 153 height, significant differences ( $P < 0.05$ ) ~~was~~ **were** observed between control 2 (un-inoculated and  
 154 untreated) (27.17cm) and all the treatments **being less than control 2**. There was also a significant  
 155 difference between the plants treated with air dried leaf powder of *B. nitida* (18.77cm) and  
 156 control 1 (6.65cm). **As for** number of leaves, there was no significant difference recorded among  
 157 the treatments including the controls. ~~Although~~ **It is worthy noticed** that the highest number of  
 158 leaves (15.00) was recorded for the plants treated with burnt leaf ash of *B. nitida*, ~~while~~  
 159 **compared to** the lowest (6.00cm) was recorded for control 1(inoculated but untreated).

160 **Table 1: Effect of treatments on the plant height and number of leaves of okra infested**  
 161 **infected with root-knot nematode (*Meloidogyne* spp.).**

Treatments	Plant height(cm)	No. of leaves per plant
Burnt leaf ash of <i>Baphia nitida</i>	11.20	15.00
Air dried leaf powder of <i>Baphia nitida</i>	18.77	13.00
Burnt leaf ash of <i>Pentaclethra macrophylla</i>	12.62	8.00
Air dried leaf powder of <i>Pentaclethra macrophylla</i>	12.00	10.00
Burnt leaf ash of <i>Delonix regia</i>	11.07	13.00
Air dried leaf powder of <i>Delonix regia</i>	7.25	10.00
Synthetic nematicide (Carbofuran 3G)	10.25	12.00
Inoculated but untreated (control 1)	6.65	6.00
Un-inoculated and untreated (control 2)	27.17	11.00
LSD <sub>(0.05)</sub>	6.25	NS

162 Where, NS = No Significant Difference

163

164 Table 2 shows the effect of treatments on number and weight of pods. There was no significant  
 165 difference ( $P < 0.05$ ) between the treatments and the controls on the number of pods. **Although It**  
 166 **is worthy noticed that** the highest number of pods was recorded by control 2 (11) and **equals to**  
 167 **that recorded to** plants treated with synthetic nematicide (11) **respectively**, with the least number  
 168 of pods (5.0) recorded for ~~plants that were inoculated but untreated~~ control 1. Significant  
 169 differences ( $P < 0.05$ ) were recorded on the weight of pods. All the treatments ~~differed~~ **were**  
 170 significantly **less** from control 2 (34.9g), except the plants treated with synthetic nematicide  
 171 (28.3g). **Also, there were significant differences between all treatments and control 1.**  
 172 **Nevertheless plants treated with air dried leaf powder of *B. nitida* (23.4g) were compared**  
 173 **favorably with the synthetic nematicide.(?)**

174 **Table 2: Effect of treatments on the number and weight of Okra pods.**

Treatments	No. of pods per plant	Weight of pods (g)
Burnt leaf ash of <i>Baphia nitida</i>	7.0	21.3
Air dried leaf powder of <i>Baphia nitida</i>	10.0	23.4
Burnt leaf ash of <i>Pentaclethra macrophylla</i>	7.0	15.6
Air dried leaf powder of <i>Pentaclethra macrophylla</i>	8.0	21.4
Burnt leaf ash of <i>Delonix regia</i>	10.0	22.0
Air dried leaf powder of <i>Delonix regia</i>	6.0	16.1
Synthetic nematicide (Carbofuran 3G)	11.0	28.3
Inoculated but untreated (control 1)	5.0	6.1
Un-inoculated and untreated (control 2)	11.0	34.9
LSD <sub>(0.05)</sub>	NS	9.75

175 Where, NS = No Significant Difference

176 The effect of treatments on fresh shoot, **and** dry shoot and fresh root weights **was illustrated in**  
 177 **Table 3. On As for** fresh shoot weight, the effect of air dried leaf powder of *B. nitida* recorded

178 the highest weight (480g) which differed significantly compared favorably with control 1  
 179 (205g) and burnt leaf ash of *Baphia nitida* (133g), but did not with control 2 and other  
 180 treatments. On Regarding dry shoot weight, there was no significant difference recorded among  
 181 the treatments including the controls. Although The highest dry shoot weight (85.2g) was  
 182 recorded for the plants treated with burnt leaf ash of *P. macrophylla*, while compared to the  
 183 lowest (30g) was recorded for the inoculated but untreated plants control 1. On Regarding fresh  
 184 root weight, all the treatments significantly differed from control 2 (371g). However, the other  
 185 same treatments did not differ significantly from control 1 and from each other.

186 **Table 3: Effect of treatments on fresh shoot, dry shoot and fresh root weights of Okra**  
 187 **infested with root-knot nematode (*Meloidogyne* spp.)**

Treatments	Fresh shoot weight (g)	Dry shoot weight (g)	Fresh root weight (g)
Burnt leaf ash of <i>Baphia nitida</i>	133.0	32.8	66
Air dried leaf powder of <i>Baphia nitida</i>	480.0	76.8	187
Burnt leaf ash of <i>Pentaclethra macrophylla</i>	335.0	85.2	118
Air dried leaf powder of <i>Pentaclethra macrophylla</i>	326.0	46.2	57
Burnt leaf ash of <i>Delonix regia</i>	308.0	64.0	136
Air dried leaf powder of <i>Delonix regia</i>	389.0	63.5	165
Synthetic nematicide (Carbofuran 3G)	362.0	70.0	132
Inoculated but untreated (control 1)	205.0	30.0	122
Un-inoculated and untreated (control 2)	629.0	68.2	371
LSD <sub>(0.05)</sub>	267.1	NS	133.2

188 Where, NS = No Significant Difference

189 Effect of treatments on the number of nematode eggs and galls in roots, galls in roots and larva  
 190 juveniles in soil are shown in Table 4. On As for number of galls in roots, all the treatments  
 191 significantly reduced no. of galls compared favorably with control 1, but the treatments did not  
 192 differ significantly from each other. Similar observations were made on the number of eggs in  
 193 root. On As for number of larva juveniles in soil, the treatments and the controls did not differ  
 194 significantly from each other. However apart from control 2 (000), plants treated with nematicide  
 195 had the least number of larva juveniles (126), while the highest number (475) was recorded for  
 196 plants treated with burnt leaf ash of *P. macrophylla*.

197 **Table 4: Effect of treatments on the population of *Meloidogyne* spp. in roots and soil.**

Treatments	No. of galls in roots	No. of eggs in roots	No. of larva juveniles in soil
Burnt leaf ash of <i>Baphia nitida</i>	0.75	126.00	350
Air dried leaf ash of <i>Baphia nitida</i>	0.25	102.00	250
Burnt leaf ash of <i>Pentaclethra macrophylla</i>	0.50	102.00	475
Air dried leaf ash of <i>Pentaclethra macrophylla</i>	1.75	202.00	351
Burnt leaf ash of <i>Delonix regia</i>	1.00	158.00	425
Air dried leaf ash of <i>Delonix regia</i>	0.75	126.00	300
Synthetic nematicide (Carbofuran 3G)	0.00	78.00	126
Inoculated but untreated (control 1)	5.00	575.00	425

Un-inoculated and untreated (control 2)	0.00	000.00	000
LSD <sub>(0.05)</sub>	2.23	272.5	NS

198 Where, NS = No Significant Difference

## 199 DISCUSSION

200 The results on the effect of the treatments on plant height agreed with the findings of Bawa *et al.*,  
 201 (2014) in which the plant extracts used had a significant effect on the height of tomato. They are  
 202 still in agreement with the findings of Kayani *et al.*, (2012) who confirmed that the symptoms for  
 203 identifying diseases caused by *Meloidogyne* spp. is the presence of galls, wilting, loss of vigor,  
 204 yellowing of leaves. Also Perry *et al.*, (2009) confirmed that the symptoms also include,  
 205 yellowing of leaves and stunting, loss of vigor, wilting due to lack of moisture, decay of tissue  
 206 due to secondary infection and yield loss. The growth reduction in crops such as Okra due to  
 207 nematode varies depending on its population density of nematode, level of cultivar susceptibility  
 208 and environmental condition to which the host is subjected to (Ononuju and Fawole, 2000).

209 The reduction in the fresh shoot and root weight could be due to slow rate of action of the active  
 210 ingredients in the plant materials (Barman and Das, 1996), or due to poor nutrients and water  
 211 flow as a result of formed giant galls on the plant roots (Ploeg, 2001) which inhibit the normal  
 212 growths and development of shoots and roots of plants affected.

213 The reduced number of nematode population and number of galls compared to control  $\geq 1$  agrees  
 214 with the observations of Adegbite and Adesiyani (2001 not found in the list of references) who  
 215 indicated that as the higher the nematode population, the higher the number of galls and the  
 216 higher the number of damage done— occurred to plant which hinders the transportation of water  
 217 and mineral nutrients from roots to aerial parts of the plant. The results obtained confirmed the  
 218 findings of Ozores-Hampton (2002) that the use of organic amendments suppressed soil phyto-  
 219 parasitic nematode populations. Abulusoro *et al.*, (2004) reported that the susceptibility of  
 220 tomato plants infected with root-knot nematodes (*Meloidogyne* spp.) show stunted growth, yield  
 221 loss and conspicuous root galls, but a number of plants are thought to contain biologically active  
 222 ingredients which when applied in the soil, reduced the incidence of plant-parasitic nematodes.  
 223 Sikora and Fernandez (2005) also found that application of sesame seed extract reduced the  
 224 incidence of root-knot nematodes and the severity of galling on okra roots. According to Akhtar  
 225 and Mahmood (1993), sesame seed extracts have a systemic activity against nematodes which  
 226 may have accounted for the lower number of galls and mean population in treated plants. Also  
 227 Vats *et al.*, (1996) reported reduction of galls and egg masses when some *M. javanica* infected  
 228 tomato plants were treated with leaf extracts of *Azadirachta indica* and *Eucalyptus tereticornis*.

229 The reduced number and weight of pods agrees with Bolles *et al.*,(2012) who reported that  
 230 nematode (*Meloidogyne* spp.) are found to be serious pest of Okra, damaging stands and greatly  
 231 decreasing and delaying the production of pods. Also, Adesiyani *et al.*, (1990) reported that some  
 232 crop varieties could be effective host of root-knot nematode that suffer no statistically significant  
 233 damage. However the extent of damage is influenced by the cultivar, nematode species, level of  
 234 soil infestation and environment (Ononuju, 1999). According to Gommers (1981) active  
 235 compounds with nematicidal activity which have been found to limit the activities of plant-  
 236 parasitic nematodes in plants includes alkaloids, diterpenes, fatty acids, glucosinolates,  
 237 isothiocyanates, phenols, polyacetylenes, sesquiterpenes and thienyls. These compounds were  
 238 extracted from different plants part such as roots, leaves and so on which could be responsible

239 for the positive effects of air dried leaf ash and burnt leaf ash of these plants on the population of  
240 nematode in the soil and root as well as in pod formation.

## 241 CONCLUSION AND RECOMMENDATION

242 The different treatments significantly decreased the population of root-knot nematode  
243 (*Meloidogyne* spp.). There were significant differences between the controls and the treatments  
244 in some of the parameters measured, while there were no significant differences in others.  
245 Synthetic nematicide (Carbofuran 3G) appeared to be more effective in the control of root-knot  
246 nematode. However, the performance of the synthetic nematicide was not significantly different  
247 from the results obtained from air dried powder of *Baphia nitida* and burnt leaf ash of *Delonix*  
248 *regia* at the rate applied. From the findings of this study, air dried powder of *B. nitida* and burnt  
249 leaf ash of *D. regia* appeared to have significantly performed better than other treatments and is  
250 being recommended as an alternative for the synthetic nematicide (Carbofuran 3G) in the control  
251 of root-knot nematode. Notwithstanding, further research work both in green house and field  
252 trials at different rates are needed in order to authenticate this findings.

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