

## Original Research Article

# Antifungal activity of copper, zinc and potassium compounds on mycelial growth and conidial germination of *Fusarium solani* f. sp. *piperis*

### ABSTRACT (ARIAL, BOLD, 11 FONT, LEFT ALIGNED, CAPS)

Fusariosis is a disease that causes economic damage to black pepper (*Piper nigrum* L.) producers. Despite being a major disease, there is no record of efficient chemical control. Thus, the objective was to evaluate the antifungal activity of copper, zinc and potassium compounds in mycelial growth and conidial germination of *Fusarium solani* f. sp. *piperis* in vitro. For inoculation in PDA (Potato Dextrose Agar) medium, 7 mm discs from the pure culture were transferred to Petri dishes. The plates were incubated at 25°C in a biochemical oxygen demand (BOD) chamber, with photoperiod of 12 h, for 15 days. Micronutrients were supplied as sulfates, CuSO<sub>4</sub> (copper sulfate) and ZnSO<sub>4</sub> (zinc sulfate), at concentrations of 1, 5, 10, 15 and 20 mmol/L. Potassium macronutrient (K) was supplied as KCl (potassium chloride) at concentrations of 30, 60, 90, 120 and 150 mmol/L. The experiment was performed using a completely randomized design with 6 treatments and ten replications. CuSO<sub>4</sub> showed fungicidal effect at concentrations of 10, 15 and 20 mmol/L. For ZnSO<sub>4</sub> mycelial growth was completely inhibited at concentrations of 15 and 20 mmol/L. There was no inhibition or reduction of fungal growth in the presence of K. Copper and zinc at minimal concentrations were efficient in controlling mycelial growth and inhibition of spore germination of *F. solani* f. sp. *piperis*. In contrast, potassium did not exert fungicidal or fungistatic effect on the fungus.

**Keywords:** *Fusarium solani*, antifungal activity, fungal growth inhibition, nutrients

### 1. INTRODUCTION (ARIAL, BOLD, 11 FONT, LEFT ALIGNED, CAPS)

Pathogenic fungi include a large and heterogeneous group of organisms that occupy an important position in both agriculture and natural populations [1]. The genus *Fusarium* spp. is classified as ascomycet, characterized by a hyaline mycelium, branched and septate, with phyllide-shaped sporophores and conidia of varying size and shape. It has spindle-shaped macroconidia with several septa [2]. *Fusarium* species are known as phytopathogens, saprophytes with worldwide distribution [3].

*Fusarium solani* (Mart.) Appel & Wr. emend. Snyder & Hans. f. sp. *piperis*, Albuquerque (Teleomorph *Nectria haematococca* Berk. & Br. f. sp. *piperis* Albuquerque) is the causal agent of fusariosis and has brought great economic damage to black pepper (*Piper nigrum* L.)

30 producers, decimating whole crops. The disease may start in the root system or in the shoot.  
31 When initiated by the roots, the root system is reduced and becomes necrotic, causing  
32 yellowish and flaccid leaves and premature fall. On the other hand, once started in the aerial  
33 part, it is characterized by the presence of yellowish branches in a very vigorous plant. With  
34 the evolution of the disease is observed drying in the upper and lower part of the plant [4-6].  
35 According to Pandey et al. [7], production losses due to plant diseases are a considerable  
36 challenge for the current agricultural production system worldwide, representing at least 25%  
37 of the total. Although fusariosis is a major disease, there is no record of efficient chemical  
38 control.

39 The methods used in the control of pathogenic isolates of *Fusarium* include the use of  
40 resistant varieties and soil disinfection with the chemical fungicide and crop rotation using  
41 non-host plants [8]. The use of resistant cultivars would be an alternative, but difficulties  
42 such as the identification of resistance genes or the pathogen's ability to adapt to new  
43 genotypes may make resistance a temporary solution [9]. In addition, the chemical control of  
44 *Fusarium* spp. is not fully efficient since the pathogen penetrates the vascular tissue of the  
45 plant [10]. Crop rotation would be of little efficiency since this pathogen is a soil fungus,  
46 capable of surviving for long periods in crop debris and presents several plant species as  
47 hosts [9]. Conventional synthetic fungicides are largely considered the most effective and  
48 economical means for treating the disease. However, the intensity of use and specific mode  
49 of action of most synthetic fungicides ultimately lead to resistance problems and an  
50 increased environmental cost [7, 11, 12]. Thus, alternative forms of control are of increasing  
51 interest, leading to the investigation and development of effective and sustainable products  
52 for the control of plant pathogens [13]. Zambolim et al. [14] reported that some micro and  
53 macronutrients have been identified as one of the main mineral elements associated with the  
54 induction of disease resistance in plants.

55 Due to their low cost, protective activity and reduced risk of resistance development  
56 controlled by the broad mode of action against pathogens, Cu compounds have been  
57 exploited to protect crops from many pests, including those that cause numerous bacterial  
58 and fungal infections [15]. According to Zambolim et al. [16], Cu ions in contact with spores  
59 or the pathogen's germ tube may accumulate in the membrane or penetrate and concentrate  
60 inside of the spores or mycelium, where they act by inhibiting enzymes essential to the  
61 metabolic process of microorganisms. Once accumulated in the cells, their effects become  
62 irreversible.

63 Interest in research on zinc (Zn) derivatives is increasing [17, 18], due to its strong  
64 antimicrobial activity at low concentrations and its non-toxic characteristics in adequate  
65 quantities. Zn as an essential micronutrient plays an important role in many integral  
66 metabolic processes [19]. It can also help increase chlorophyll and carotenoid biosynthesis  
67 and improve plant photosynthetic apparatus [20]. Significant optoelectric, physical and  
68 antimicrobial properties of Zn offer great potential for increasing agricultural productivity [21].  
69 Its mode of action is not completely understood, but it is known to act directly on the  
70 pathogen [16].

71 Among macronutrients of great importance to the plant, potassium (K) is one of the elements  
72 that has very positive results in reducing the incidence of pests and diseases [22, 23], being  
73 able to reduce the severity of more of 100 fungi [24]. Taiz and Zeiger [25] reported that K is  
74 an essential plant nutrient required as a cofactor for over 40 enzymes, many of which are  
75 involved in respiration and photosynthesis. As such, it is an important nutrient in plant  
76 disease prevention as it is involved in many cellular processes that influence disease  
77 severity. Its effect on the prevention of diseases caused by bacteria, fungi and nematodes  
78 has been reported [16, 26]. Increased resistance to disease from K fertilization has been

79 attributed to several mechanisms, such as cell permeability and decreased susceptibility of  
80 tissues to pathogen maceration and penetration [16, 27]. K influences the reduction of plant  
81 diseases due to the activation of enzymes involved in respiration and photosynthesis, carbon  
82 chain supply processes for defense substance synthesis, as well as stomatal regulation  
83 influencing mass flow solute transport [28].

84 The use of nutrients with antifungal action may be a strategy for controlling pathogens that  
85 cause invaluable economic losses. Nutrients such as Cu, Zn and K are easily accessible,  
86 inexpensive and still contribute to plant nutrition. Thus, the objective of this study was to  
87 evaluate the antifungal activity of Cu, Zn and K compounds against *Fusarium solani* f. sp.  
88 *piperis* growth.  
89

## 90 2. MATERIAL AND METHODS

### 91 2.1. Microorganism and cultivation

92 The isolate of *Fusarium solani* f. sp. *piperis* CML 2466, from the Coleção Micológica de  
93 Lavras, Federal University of Lavras - MG was used. The fungus was maintained on Petri  
94 dishes containing PDA (Potato Dextrose Agar) at 4 ° C. For inoculation, 7 mm pure culture  
95 discs were transferred to Petri dishes containing the same medium. The plates were  
96 incubated at 25° C in BOD (Biochemical Oxygen Demand), photoperiod of 12h for 15 days.  
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### 100 2.2. Copper, zinc and potassium concentrations

101 Micronutrients were supplied in PDA medium as copper sulfate (CuSO<sub>4</sub>) and zinc sulfate  
102 (ZnSO<sub>4</sub>) at concentrations of 1, 5, 10, 15 and 20 mmol/L. Potassium macronutrient (K) was  
103 supplied as potassium chloride (KCl) at concentrations of 30, 60, 90,120 and 150 mmol/L.  
104 The nutrients were diluted in sterile distilled water and at the time of plating were added to  
105 the PDA culture medium in laminar flow hood. After solidification, a 7 mm diameter fungal  
106 mycelium disc with 15-day-old was transferred to the center of each petri dish (68 mm  
107 diameter). The PDA medium with the fungus disc was used as control. The plates were  
108 sealed with Parafilm and maintained in BOD at 25°C with 12 h photoperiod.  
109

### 110 2.3. Mycelial growth

111 The evaluation of *F. solani* mycelial growth in the control plates and treatments was  
112 determined every 2 days by measuring the diameter of the colonies in orthogonal directions  
113 with the digital pachymeter until the control treatment colony reached edge of the plate, ie 12  
114 days after inoculation (DAI). The growth inhibition percentage was calculated according to  
115 Guo et al. [29], where the antifungal index (%) =  $(1 - D_a / D_b) \times 100$ , where:  $D_a$  gives the  
116 diameter of the growth zone in the test plate and  $D_b$  the diameter of the growth zone in the  
117 control plate.  
118  
119

### 120 2.4. Spore count

121 The spore suspension was prepared by adding 20 mL of sterile distilled water to each plate  
122 containing the fungus, which was scraped with a Drigalsky handle for efficient spore  
123 extraction. The spore count was performed in a Neubauer Chamber and the suspension was  
124 adjusted to a concentration of 10<sup>6</sup> spores/mL<sup>-1</sup>.  
125

### 126 2.5. Statistical analysis

127

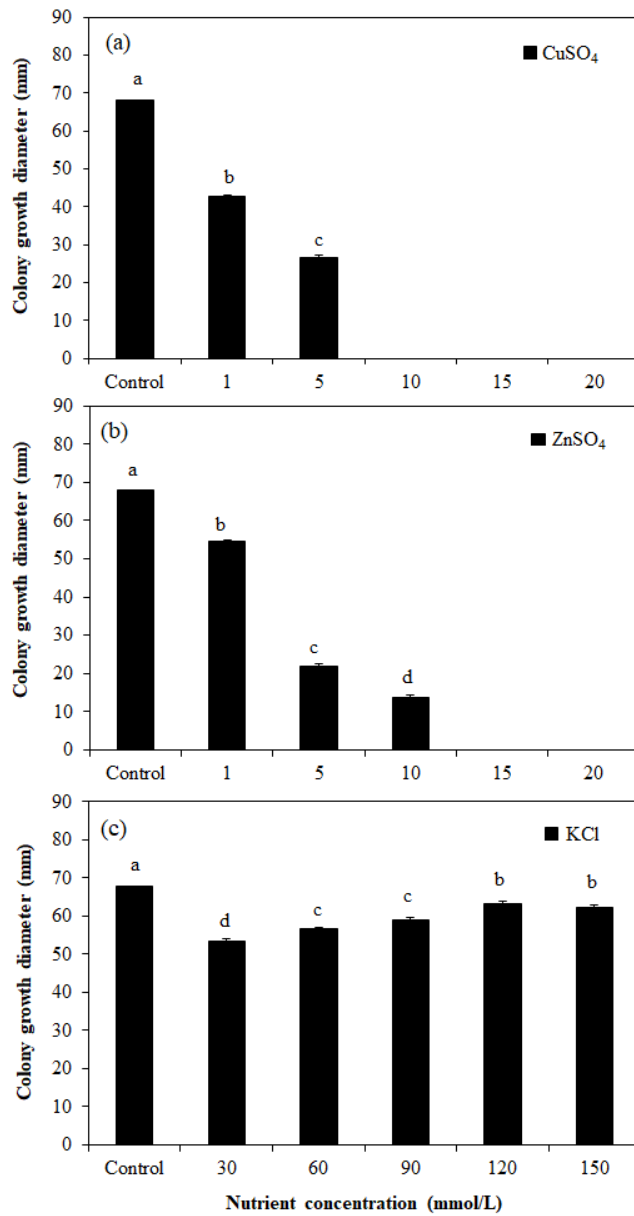
**Comment [LL1]:** Actually you have 3 experiments (Cu, Zn and K) with 5 doses for each and analyzed in 6 periods (2, 4, 6, 8, 10 and 12 days). You should improve your statistics. Quantitative data is performed regression and not average test.

128 The experiment was performed using a completely randomized design with 6 treatments and  
129 10 replications for each treatment (Cu, Zn and K). Each repetition consisted of a petri dish.  
130 All data were submitted to analysis of variance (ANOVA) and Tukey test using Genes  
131 software [30].  
132

### 133 3. RESULTS

#### 134 3.1. Mycelial growth

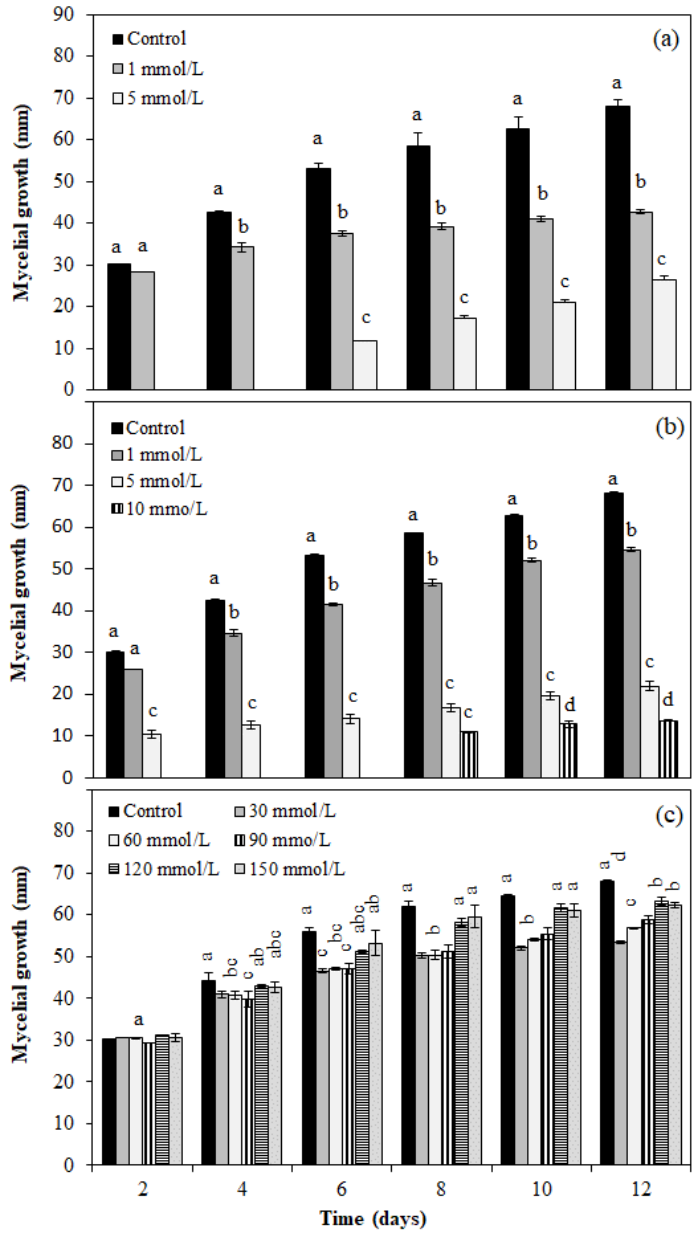
135  
136 The mycelial growth of *F. solani* was dependent on the nutrient and the dose used. Fungal  
137 growth was completely inhibited in some treatments. The  $\text{CuSO}_4$  showed fungicidal effect at  
138 concentrations of 10, 15 and 20 mmol/L, completely inhibiting growth of the colonies (Fig.  
139 1a). However, at a concentration of 5 mmol/L, a fungistatic effect was observed since  
140 mycelial growth was initiated 6 DAI (Fig. 2a). For  $\text{ZnSO}_4$  mycelial growth was completely  
141 inhibited at concentrations of 15 and 20 mmol/L, showing fungicidal effect and significantly  
142 reduced ( $P \leq 0.05$ ) at concentrations of 5 and 10 mmol/L, exerting fungistatic effect (Fig. 1b).  
143 This result is ratified after eight days of incubation by observing mycelial growth at a  
144 concentration of 10 mmol/L (Fig. 2b). There was a significant difference for KCl treatment  
145 ( $P \leq 0.05$ ) between the tested concentrations. However, there was no inhibition or reduction of  
146 fungal growth in the presence of this nutrient (Fig. 1c and 2c).  
147  
148



**Fig. 1. Antifungal activities of CuSO<sub>4</sub> (a), ZnSO<sub>4</sub> (b) and KCl (c) against *F. solani* f. sp. *piperis* on PDA at different concentrations.**

Data are shown as average values. Columns followed by the identical letter are not statistically different according to by Tukey test, at 5% probability ( $P \leq 0.05$ ). Bars represent the standard error of the mean.

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Fig. 2. Effect of CuSO<sub>4</sub> (a), ZnSO<sub>4</sub> (b) and KCl (c) on the mycelial growth of *F. solani* f. sp. *piperis*, 12 days after inoculation. Control: only PDA medium.

159 Data are shown as average values. Columns followed by the identical letter are not statistically  
 160 different according to by Tukey test, at 5% probability ( $P \leq 0.05$ ). Bars represent the standard error  
 161 of the mean.  
 162

### 163 3.2. Percent growth inhibition (P.I.).

164 The mycelial growth inhibition index confirmed the efficiency of the antifungal activity of  
 165  $\text{CuSO}_4$  and  $\text{ZnSO}_4$  (Table 1). For  $\text{CuSO}_4$ , at a concentration of 5 mmol/L, there was  
 166 inhibition greater than 50% 12 DAI. The other concentrations inhibited 100% fungal growth.  
 167 Similar results were observed for  $\text{ZnSO}_4$  (Table 1). However, for KCl, in none of the  
 168 evaluated concentrations was observed P.I. below 50%. At 2 DAI there was growth induction  
 169 (Table 1) with no fungistatic or fungicidal effect for this nutrient.  
 170  
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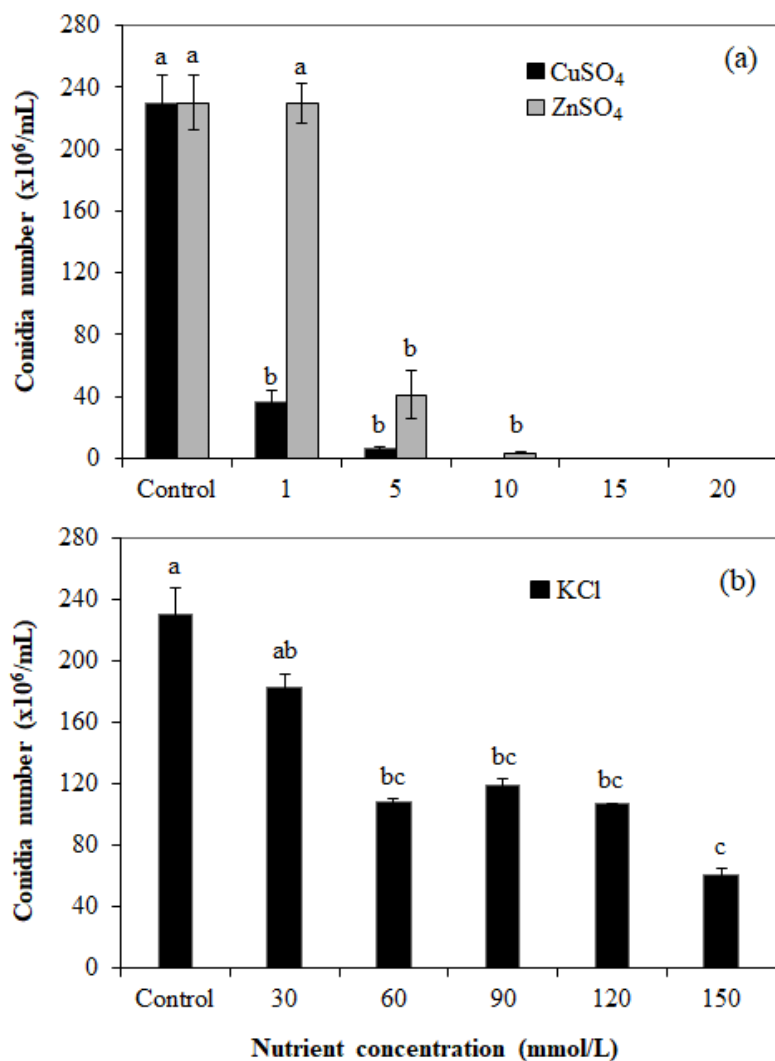
172 **Table 1. Percent growth inhibition (P.I.) of *F. solani* f. sp. *piperis* in  $\text{CuSO}_4$ ,  $\text{ZnSO}_4$   
 173 and KCl**  
 174  
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Time (days)	P.I. (%)					
	2	4	6	8	10	12
<b><math>\text{CuSO}_4</math> mmol/L</b>						
Control	0	0	0	0	0	0
1	12.0	19.4	29.4	33.0	34.5	37.2
5	100	100	77.8	70.7	66.4	61.0
10	100	100	100	100	100	100
15	100	100	100	100	100	100
20	100	100	100	100	100	100
<b><math>\text{ZnSO}_4</math> mmol/L</b>						
Control	0	0	0	0	0	0
1	14.4	18.8	22.0	20.3	17.0	19.8
5	65.7	70.4	73.6	71.5	68.9	67.7
10	100	100	89.9	81.5	79.5	79.9
15	100	100	100	100	100	100
20	100	100	100	100	100	100
<b>KCl mmol/L</b>						
Control	0	0	0	0	0	0
30	-0.9	7.1	16.6	18.7	19.2	21.4
60	-3.8	8.0	15.6	18.8	16.2	16.5
90	2.8	9.8	15.8	17.4	14.0	13.3
120	-3.0	2.7	8.2	6.1	4.4	7.0
150	-1.4	3.7	4.8	3.8	5.0	8.3

### 176 3.3. Conidia Number.

177 Twelve days after inoculation (12 DAI), the conidia number of *F. solani* was inhibited in the  
 178 presence of  $\text{CuSO}_4$ ,  $\text{ZnSO}_4$  and KCl. The Cu micronutrient reduced by 84% the conidial  
 179 germination at 1mmol/L concentration in relation to the control. The same was not observed  
 180 for Zn at the same concentration (Figure 3a, Table 2). In the other Cu and Zn  
 181 concentrations, conidial germination was significantly inhibited ( $P \leq 0.05$ ), with values greater  
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186 than 80%. For K treatment, there was a 20.6% reduction in the number of conidia at 30  
 187 mmol/L. The other concentrations presented a reduction greater than 50% when compared  
 188 to the control (Figure 3b, Table 2).  
 189



190  
 191  
 192 **Fig. 3. Conidia production of *F. solani* f. sp. *piperis* on CuSO<sub>4</sub>, ZnSO<sub>4</sub> (a) and KCl (b) in**  
 193 **different concentrations.**

194 *Data are shown as average values. Columns followed by the identical letter are not statistically*  
 195 *different according to by Tukey test, at 5% probability (P≤0.05). Bars represent the standard error*  
 196 *of the mean.*  
 197



198 **Table 2. Percent growth inhibition (P.I.) of *F. solani* f. sp. *piperis* in CuSO<sub>4</sub>, ZnSO<sub>4</sub>**  
 199 **and KCl**  
 200

Reduction in Conidia number (%)	
mmol/L	CuSO <sub>4</sub>
1	84.2
5	97.2
10	100.0
15	100.0
20	100.0
ZnSO <sub>4</sub>	
1	0.0
5	82.2
10	98.4
15	100.0
20	100.0
KCl	
30	20.6
60	52.9
90	48.3
120	53.6
150	73.5

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 202

#### 4. DISCUSSION

203  
 204  
 205 The widespread and persistent nature of *Fusarium* spp. may be due to its ability to maintain  
 206 and multiply in a wide variety of complex carbohydrates and proteins, thus resisting adverse  
 207 climates and high levels of toxic substances such as many antibiotics and fungicides. *F.*  
 208 *solani* seems to incorporate some of the most difficult members of the genus [31].

209 Inhibition of mycelial growth of *F. solani* f. sp. *piperis* in vitro revealed significant variations in  
 210 fungus sensitivity to different nutrients tested. In the present study, CuSO<sub>4</sub>, ZnSO<sub>4</sub> were  
 211 more effective in inhibiting fungal growth, while KCl showed relatively weak effect comparing  
 212 it to other nutrients. However, it was observed that Cu was the most effective compound  
 213 against mycelial growth and *F. solani* conidia germination, showing significant inhibition at  
 214 relatively low concentrations of the compound.

215 According to Resende et al. and Melo et al. [32, 33], Cu, Zn and K compounds are used for  
 216 the control of microorganisms. They can have a direct effect on the pathogen (fungicidal or  
 217 fungistatic effect) or activate the natural defense of plants, resulting in induced resistance  
 218 [34, 35].

219 The Cu is currently used due to its antifungal properties. In particular, Cu is responsible for  
 220 interference with homeostatic processes and cell membrane functions, protein synthesis  
 221 damage, reactive oxygen species production, and DNA disruption [36, 37]. Civardi et al. [38]  
 222 observed that Cu exerted toxic effect on *Rhodonia placenta* fungal cell by breakdown of  
 223 different basic metabolic processes. Significant antifungal activity of Cu has been revealed in

224 a number of pathogenic species including *Fusarium* sp., *Aspergillus niger*, *Rhizoctonia*  
225 *solani*, *Alternaria solani*, *Alternaria alternate* and *Phoma destructiva* [7, 39, 40].

226 Regarding Zn, several studies have shown its antibacterial activity [41-44]. However, there  
227 are few studies reporting the suggested mechanism for antifungal activity of Zn compounds  
228 [45, 46]. Some authors suggest that such a mechanism may be based on the formation of  
229 reactive oxygen species that disrupt the integrity of the cell membrane, preventing pathogen  
230 growth [47, 48, 44, 46]. According to He et al., Król et al. and Ashajyothi et al. [46, 49, 50],  
231 Zn compounds showed fungistatic potential against *Fusarium* sp., *Botrytis cinerea*,  
232 *Penicillium expansum*, *Aspergillus niger* and *Rhizopus stolonifer*. Chand et al. [51] observed  
233 that among the micronutrients tested Zn presented the greatest inhibition of mycelial growth  
234 of *Fusarium oxysporum* f. sp. *cuban*.

235 The marked toxic effect of Cu and Zn against fungal spores compared to mycelial growth  
236 can be attributed to the structural differences between the spore wall and the fungal  
237 vegetative phase. Bartnicki-Garcia [52] observed that the chitin content of many fungal  
238 species is significantly higher in the hyphae wall compared to the spore wall, making the  
239 latter more susceptible to some compounds. In addition, during the spore germination  
240 process, the presence of enzymes such as disulfide reductase and glucanases result in  
241 weakening of the cell wall, facilitating germ tube lengthening and thus creating sites of  
242 greater sensitivity to toxic substances in contact with the cell fungal. In general, conidial  
243 germination may reflect reproductive capacity and fungal development. Savi et al. [18]  
244 suggest that the effect of Zn compounds on fungal growth may be related to their property,  
245 altering reproductive capacity in terms of conidia viability. Malandrakis et al. [53], studying  
246 the effect of copper and zinc on various microorganisms, found that Cu was effective against  
247 *Alternaria alternata*, *Botrytis cinerea*, *Monilia fructicola*, *Verticillium dahliae*, *Colletotrichum*  
248 *gloeosporioides*, *Fusarium oxysporum* f. sp. *radicis lycopersici* and *Fusarium solani* while Zn  
249 exerted a fungicidal effect against *M. fructicola*, *F. solani* and *V. dahliae*.

250 Although K has not exerted inhibition or reduction of *F. solani* mycelial growth and spore  
251 production, there are reports in the literature that K acts as an inducer of resistance to plant  
252 diseases [54-56]. The use of K as a plant fertilizer may decrease the incidence of fungal and  
253 bacterial as well as insect diseases [57], mainly due to changes in primary metabolism and  
254 plant hormonal responses [27, 55]. Dordas [58] observed that the application of KCl on  
255 foliage can prevent the attack of mildew on wheat.

## 256 257 **5. CONCLUSION** 258

259 The results obtained in this work provide evidence that copper and zinc exhibited beneficial  
260 antifungal activity against *F. solani* f. sp. *piperis* under laboratory conditions. In contrast, no  
261 potassium concentration was active against the fungus. High antifungal activity was  
262 observed at low concentrations of copper and zinc, favoring the use of these compounds.  
263 Scientific evaluations are being carried out in the field to verify the performance of these  
264 nutrients as growth inhibitors of *Fusarium*.

## 265 266 267 **COMPETING INTERESTS** 268

269 The authors declare that they have no conflicts of interest.  
270

271 **REFERENCES**

272

273 1. Burdon JJ, Silk J. Sources and patterns of diversity in plant-pathogenic fungi.  
274 *Phytopathology*. 1997;87:664-669.

275 2. Leal PC, Cantanhede KL, Silva LM, Bezerra GFB. Micotoxinas do *Fusarium* e seu  
276 potencial carcinogênico. 70ª ed. São Paulo: NewsLab Press;2005. Brazil

277 3. Hennequin C, Abachin E, Symoens F, Lavarde V. Identification of *Fusarium* species  
278 involved in human infections by 28S rRNA gene sequencing. *J Clin Microbiol*. 1999;37:3586-  
279 3589.

280 4. Ventura JA, Costa H. Manejo da fusariose da pimenta-do-reino no estado do Espírito  
281 Santo. Vitória: Instituto Capixaba de Pesquisa, Assistência Técnica e Extensão Rural;2004.  
282 Brazil.

283 5. Tremacoldi CR. Principais doenças fúngicas da pimenteira-do-reino no estado do Pará e  
284 recomendações de controle. Belém: Embrapa Amazônia Oriental;2010. Brazil.

285 6. Silva SO, Neto APD, Silva MB. Pimenta-do-reino: importância da defesa fitossanitária  
286 para a sustentabilidade da atividade na região norte do Espírito Santo. *Rev Bras Agropecu*.  
287 *Sustent*. 2011;1:88-92. Brazil.

288 7. Pandey S, Giri K, Kumar R, Mishra G. Nanopesticides: opportunities in crop protection  
289 and associated environmental risks. *Proc Natl Acad Sci Sect B Biol Sci*. 2018;88:1287-  
290 1308.

291 8. Agrios GN. *Plant Pathology*. 5th ed. New York: Academic Press; 2005.

292 9. Sutton JC. Strategies for biological control of necrotrophic pathogens in perennial crops.  
293 *Fitopatol Bras*. 2000;25:235-238.

294 10. Tokeshi H. Doenças e pragas agrícolas geradas e multiplicadas pelos agrotóxicos.  
295 *Fitopatol Bras*. 2000;25:264-271. Brazil.

296 11. Kah M, Kookana RS, Gogos A, Bucheli TD. A critical evaluation of nanopesticides and  
297 nanofertilizers against their conventional analogues. *Nat Nanotechnol*. 2018;13:677-684.

298 12. Sun Q, Li J, Le, T. Zinc oxide nanoparticle as a novel class of antifungal agents: current  
299 advances and future perspectives. *J Agric Food Chem*. 2018;66:11209-11220.

300 13. Deliopoulos T, Kettlewell PS, Hare MC. Fungal disease suppression by inorganic salts: a  
301 review. *Crop Protec*. 2010;29:1059-1075.

302 14. Zambolim L, Rodrigues FA, Capucho AS. Resistência a doenças de plantas induzida  
303 pela nutrição mineral. In: Controle alternativo de pragas e doenças. Venzon M, Paula Junior  
304 TJ, Pallini A, Editors. Viçosa: Epamig/CTZM; 2005. p. 275-318.

305 15. Keller AA, Adeleye AS, Conway JR. Comparative environmental fate and toxicity of  
306 copper nanomaterials. *NanoImpact*. 2017;7:28-40.

- 307 16. Zambolim L, Ventura AJ, Zanão LA. Efeito da nutrição mineral no controle de doenças  
308 de plantas. Viçosa: Editora Independente,2012. Brazil.
- 309 [17. Savi GD, Vitorino V, Bortoluzzi AJ, Scussel VM. Effect of zinc compounds on *Fusarium*  
310 *verticillioides* growth, hyphae alterations, conidia, and fumonisin production. J Sci Food  
311 Agric. 2013;93:3395–3402.
- 312 18. Savi GD, Vitorino V, Bortoluzzi AJ, Scussel VM. Antifungal properties of zinc-compounds  
313 against toxigenic fungi and mycotoxin. Int J Food Sci Technol. 2013;48:1834–1840.
- 314 19. Rout GR, Das P. Effect of metal toxicity on plant growth and metabolism. Agronomie  
315 2003;23:3–11.
- 316 20. Aravind P, Prasad MNV. Zinc protects chloroplasts and associated photochemical  
317 functions in cadmium exposed *Ceratophyllum demersum* L., a fresh water macrophyte. Plant  
318 Sci. 2004;166:1321–1327.
- 319 21. Hussain I, Singh NB, Singh A, Singh H. Green synthesis of nanoparticles and its  
320 potential application. Biotechnol Lett. 2016;38:545–560.
- 321 22. Graham DR. Effects of nutrients stress on susceptibility of plants to disease with  
322 particular reference to the trace elements. Adv Bot Res. 1983;10:221–276.
- 323 23. Sharma S, Duveiller E, Basnet R, Karki CB. Effect of potash fertilization on  
324 *Helminthosporium* leaf blight severity in wheat, and associated increases in grain yield and  
325 kernel weight. Field Crop Res. 2005;93:142–150.
- 326 24. Raij, van B. Potássio: necessidade e uso na agricultura moderna. Piracicaba:  
327 Potafós,1990. Brazil.
- 328 25. Taiz L, Zeiger E. Fisiologia e Desenvolvimento Vegetal. 6ª ed. Porto Alegre:  
329 Artmed,2017. Brazil.
- 330 26. Rawat N, Pumphrey MO, Li S, Zhang X. Wheat Fhb1 encodes a chimeric lectin with  
331 agglutinin domains and a pore-forming toxin-like domain conferring resistance to *Fusarium*  
332 head blight. Nat Genet. 2016;48:1576–1580.
- 333 27. Wang M, Zheng Q, Shen Q, Guo S. The critical role of potassium in plant stress  
334 response. Int J Mol Sci. 2013;14:7370–7390.
- 335 28. Bloom AJ. Nutrição mineral. In: Fisiologia Vegetal. Taiz L, Zeiger E, Editors. Porto  
336 Alegre: Artmed,2004. p. 96-103.
- 337 29. Gu Z, Chen R, Xing R, Liu S. Novel derivatives of chitosan and their antifungal activities  
338 in vitro. Carbohydr Res. 2006;341:351–354.
- 339 30. Cruz CD. Genes Software – extended and integrated with the R, Matlab and Selegen.  
340 Acta Sci. 2016;38:547-552.
- 341 31. Smith SN. An overview of ecological and habitat aspects in the genus *Fusarium* with  
342 special emphasis on the soil-borne pathogenic forms. Plant Pathol Bull. 2007;16:97–120.

- 343 32. Resende MLV. Indução de resistência na cafeicultura: perspectivas de uso. In: Manejo  
344 fitossanitário da cultura do cafeeiro. Blun, LEB, Editor. Lavras: Universidade Federal de  
345 Lavras,2008. p. 25-35. Brazil
- 346 33. Melo LGDL, Silva EKC, Campos Neto JRM, Lins SRO. Indutores de resistência abióticos  
347 no controle da fusariose do abacaxi. Pesq Agropec Bras. 2016;51:1703-1709. Brazil.
- 348 34. Spolti P, Valdebenito-Sanhueza RM, Campos AD, Del Ponte EM. Modo de ação de  
349 fosfitos de potássio no controle da podridão olho de boi em maçã. Summa Phytopathol.  
350 2015;41:42-48. Brazil.
- 351 35. Costa BHG, Resende MLV, Ribeiro Júnior PM, Mathioni SM. Suppression of rust and  
352 brown eye spot diseases on coffee by phosphites and by-products of coffee and citrus  
353 industries. J Phytopathol. 2014;162:1-8.
- 354 36. Krumova EZ, Pashova SB, Dolashka-Angelova PA, Stefanova T. Biomarkers of oxidative  
355 stress in the fungal strain *Humicola lutea* under copper exposure. Process Biochem.  
356 2009;44:288-295.
- 357 37. Rai M, Ingle PA, Pandit R, Paralikar P. Copper and copper nanoparticles: role in  
358 management of insect-pests and pathogenic microbes. Nanotechnol Rev. 2018;1-14.
- 359 38. Civardi C, Schwarze FWMR, Wick P. Micronized copper wood preservatives: An  
360 efficiency and potential health risk assessment for copper-based nanoparticles. Env Poll.  
361 2015;200:126-132.
- 362 39. Nemati A, Shadpour S, Khalafbeygi H, Ashraf S. Efficiency of hydrothermal synthesis of  
363 nano/microsized copper and study on in vitro antifungal activity. Mater Manuf Process.  
364 2015;30:63-69.
- 365 40. Aleksandrowicz-Trzcinska M, Szaniawski A, Olchowik J, Drozdowski S. Effects of copper  
366 and silver nanoparticles on growth of selected species of pathogenic and wood-decay fungi  
367 in vitro. For Chron. 2018;94:109-116.
- 368 41. Yamamoto O. Influence of particle size on the antibacterial activity of zinc oxide. Int J  
369 Inorg Mater. 2001;3:643-646.
- 370 42. Stoimenov PK, Klinger RL, Marchin GL, Klabunde JS. Metal oxide nanoparticles as  
371 bactericidal agents. Langmuir. 2001;18:6679-6686.
- 372 43. Zhang L, Jiang Y, Ding Y, Povey M. Investigation into the antibacterial behaviour of  
373 suspensions of ZnO nanoparticles (ZnO nanofluids). J Nanopart Res. 2007;9:479-489.
- 374 44. Liu Y, He L, Mustapha A, Li H. Lin Antibacterial activities of zinc oxide nanoparticles  
375 against *Escherichia coli* O157:H7Y. J Appl Microbiol. 2009;107:1193-1201.
- 376 [45] Sawai J, Yoshikawa T. Quantitative evaluation of antifungal activity of metallic oxide  
377 powders (MgO, CaO and ZnO) by an indirect conductimetric assay. J Appl Microbiol  
378 2004;96:803-9.
- 379 [46] He L, Liu Y, Mustapha A, Lin M. Antifungal activity of zinc oxide nanoparticles against  
380 *Botrytis cinerea* and *Penicillium expansum*. Microbiol Res 2011;166:207-215.

- 381 [47] Feng QL, Wu J, Chen GQ, Cui FZ. A mechanistic study of the antibacterial effect of  
382 silver ions on *Escherichia coli* and *Staphylococcus aureus*. J Biomed Mat Res 2000;52:662–  
383 668.
- 384 [48] Applerot G, Lipovsky A, Dror R. Enhanced antibacterial activity of nanocrystalline ZnO  
385 due to increased ROS mediated cell injury. Adv Funct Mater 2009;19:842–852.
- 386 [49] Król A, Pomastowski P, Rafińska K, Railean-Plugaru V. Zinc oxide nanoparticles:  
387 synthesis, antiseptic activity and toxicity mechanism. Adv Colloid Interf Sci 2017;249:37–52.
- 388 [50] Ashajyothi C, Prabhurajeshwar C, Handral HK, Kelmani CR. Investigation of antifungal  
389 and anti-mycelium activities using biogenic nanoparticles: an eco-friendly approach. Environ  
390 Nanotechnol Monit Manag 2016;5:81–87.
- 391 [51] Chand G, Jaiswal US, Maru AK. Effect of micronutrients on Panama wilt of banana  
392 (*Fusarium oxysporum* f. sp. *cubense*) and its synergistic action with *Trichoderma viride*.  
393 Conference Proceedings of International Conference on Innovative Approaches in Applied  
394 Sciences and Technologies (iCiAsT-2016) February 01-05, 2016.
- 395 [52] Bartnicki-Garcia S. Cell wall chemistry, morphogenesis, and taxonomy of fungi. Annu  
396 Rev Microbiol 1968;22:87–108.
- 397 [53] Malandrakis AA, Kavroulakis N, Chrysikopoulos CV Use of copper, silver and zinc  
398 nanoparticles against foliar and soil-borne plant pathogens. Sci Total Environ 2019;670:292–  
399 299
- 400 [54] Holzmüller EJ, Jose S, Jenkins MA. Influence of calcium, potassium, and magnesium  
401 on *Cornus florida* L. density and resistance to dogwood anthracnose. Plant Soil  
402 2007;290:189–199.
- 403 [55] Zorb C, Senbayram M, Peiter E. Potassium in agriculture—status and perspectives. J  
404 Plant Physiol 2014;171:656–669.
- 405 [56] Gao X, Zhang S, Zhao X, Wu O. Potassium-induced plant resistance against soybean  
406 cyst nematode via root exudation of phenolic acids and plant pathogen-related genes. PLoS  
407 ONE 2018;13:1-13.
- 408 [57] Amtmann A, Troufflard S, Armengaud P. The effect of potassium nutrition on pest and  
409 disease resistance in plants. Physiol Plantarum 2008;133:682–691
- 410 [58] Dordas C. Role of nutrients in controlling plant diseases in sustainable agriculture. A  
411 review. Agro Sustain Dev 2008;28:33–46.

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#### 415 **DEFINITIONS, ACRONYMS, ABBREVIATIONS**

416 Here is the Definitions section. This is an optional section.

417 **Term:** Definition for the term

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