

Effect of ethephon and storage temperature on physico-chemical changes during ripening of mango (*Mangifera indica* L.) Cv. Neelum

Abstract

Mango fruits Cv. Neelum treated with ethephon of different concentrations i.e. 250, 500, 750 and 1000 ppm for five minutes at different temperature conditions viz., 16, 20, 24 and 28 °C with 80% RH ripened in ripening chamber and untreated fruits kept at ambient temperature (30-34 °C). Then the mango fruits were analyzed for physico-chemical changes and sensory qualities. It was found that mango Cv. Neelum ethephon dip treatment placed in the ripening chamber triggered the ripening process and showed that the significant increasing trends in L*, a*, b* values of colour, TSS (°Brix), PLW (%), reducing sugars (%), pH and decreasing trends in firmness (N), acidity during ripening in all the treatment combinations during advancement of storage period in ripening chamber. It was observed that mango fruits Cv. Neelum ripened by ethephon dip treatment of 750 ppm for 5 minutes at 20 °C, 80% RH showed better results in respect of a high overall acceptability score of 8.50.

Keywords: Mango, Ethephon, Ripening, Physico-chemical changes

Introduction

Mango (*Mangifera indica* L.) belongs to the family *Anacardiaceae*, is considered as one of the choicest fruits of the world because of its attractive colour, delicious taste, and excellent nutritional value. Mango is the most important tropical fruit in India accounting for 2.29 million ha of an area with a production of 15.19 million tons and a productivity of 6.63 t ha⁻¹. India is estimated to account for about 40 per cent of the world's mango production. (PIB, 2017). One-fifth of the total fruit produced in the country is mango next to Banana. Mango is one of the most extensively exploited fruit because of its flavour, fragrance and juice content. It is a good source of fiber, sugars, vitamins, and minerals along with anti-oxidants.

Fruit ripening is a genetically programmed stage of development overlapping with senescence (Watada *et al.*, 1984). Mango fruits ripen unevenly on the tree and the natural ripening process can be very slow and unpredictable. To overcome this problem, fruits can be ripened artificially by exposing the fruits to certain chemicals, which initiate the ripening process. (Campbell and Malo, 1969) found that the ripening of mature-green mangos was accelerated in response to ethylene released from 2- chloroethylphosphonic acid (Ethephon). Ethephon/ethrel is an ethylene-releasing chemical, which can be used to improve fruit colour development and stimulates ripening process of the fruit.

41 However, ripening of fruit generally is not completed on the tree because natural
42 ripening is a long process. Ethylene is one of the natural ripening hormones which is
43 responsible for accelerating the normal process of maturation, senescence and ripening. The
44 use of ethylene gas helps in achieving faster and more uniform ripening of fruits (Kader,
45 2002). Ethylene gas is commercially applied in the form of liquid i.e. Ethephon. In the case of
46 ethephon, the ripening is slightly cumbersome, the fruit sellers have to either dip the fruits in
47 a solution or pass through fumes of this chemical (Siddiqui and Dhua, 2009; Kulkarni *et al.*,
48 2004). In alkaline medium, ethylene is evolved from ethephon (Thompson and Seymour,
49 1982). The ripening of green mangoes was accelerated by 2-chloroethyl phosphonic acid
50 (ethephon). Early and uniform ripening and colour development can be achieved by dipping
51 of fruits in diluted ethephon (2-Chloroethyl phosphonic acid) solution which is recommended
52 for a number of climacteric fruits including mango (Venkatesan and Tamilmani, 2013, Gupta
53 *et al.*, 2015),
54

55 The ripening with ethephon treatment seems to hold promise to get good and uniform
56 quality. In this technique, the fruits are dipped in different concentrations and placed in a
57 ripening chamber for 24 h to induce the fruits to ripen. The most important thing in this
58 technique is temperature and relative humidity control inside the ripening chamber which
59 should range between 16-25 °C and 90-95% RH, depending upon the fruit type (Mahajan and
60 Ghuman, 2010). And also, the ripening technology can be used at wholesale markets before
61 distribution to retailers. Therefore, the present investigation was carried out to study the
62 effect of different concentrations of ethephon at different temperatures on physico-chemical
63 changes during ripening behaviour of mango Cv. Neelum.
64

65 **Materials and Methods**

66 The present research entitled “Effect of ethylene on physico-chemical changes during
67 ripening of mango (*Mangifera indica* L.) Cv. Neelum.” was carried out in the Process
68 Engineering Laboratory, Department of Processing and Food Engineering, Dr. NTR College
69 of Agricultural Engineering, Bapatla during the year 2018 - 2019. Physiologically mature,
70 healthy green fruits at an optimum maturity of mango Cv. Neelum was collected from
71 Agricultural Research Farm, Bapatla. Manually harvested fruits were washed with
72 chlorinated water for further investigation. The ripening chamber made up of polyurethane
73 foam panels having a thickness of 60 mm with a dimension of 3.6 x 3.0 x 3.6 m having the
74 capacity to hold 3500-4000 kg of mango fruits.

75 The selected mango fruits Cv. Neelum was kept in the ripening chamber at Dr. NTR
76 College of Agricultural Engineering, Bapatla. For the preparation of 250, 500, 750 and 1000
77 ppm of ethephon solutions, 0.64, 1.28, 1.92 and 2.56 mL of ethrel (2-Chloroethylphosphonic
78 acid 39% aqueous solution under the brand name ethefol) respectively were dissolved in
79 1litre of distilled water. Uniform sized fruits were dipped in ethephon solution for 5 min and
80 air-dried to remove excess moisture. Fruits treated with different concentrations such as 250,
81 500, 750 and 1000 ppm at different temperatures of 16, 20, 24 and 28 °C with relative
82 humidity 90% in ripening chamber and control sample under ambient conditions (35 °C, 55-
83 65% RH). Further, physico-chemical analysis of mango fruits was recorded at every two days
84 interval.

85 **Independent Variables**

86 **Treatments for mango fruits**

87 ➤ Temperatures – 16, 20, 24, 28 °C

88 **Ripening agent**

89 ➤ Ethephon - 250, 500, 750 & 1000 ppm

90 **Dependent Variables**

91 **Physico-Chemical Parameters** - Physiological loss in weight, firmness, total soluble
92 solids, pH, titratable acidity, reducing sugars,
93 vitamin C and colour.

94 **Sensory Analysis** - 9 point hedonic scale

95 The physiological loss in weight (PLW) after each interval of storage was calculated
96 by subtracting the final weight from the initial weight of the fruits and expressed in per cent.
97 The fruit firmness was measured with the help of a fruit penetrometer (Model FT- 327, USA)
98 using an 8 mm stainless steel probe. Total soluble solids (TSS) was determined by Hand
99 refractometer and expressed in °brix. Reducing sugars and titratable acidity of fruits were
100 recorded by a method as suggested by Ranganna (2010). Ascorbic acid of mango fruits were
101 estimated by a method as suggested by using 2,6-dichlorophenol indophenol dye titration
102 method of Sadasivam and Manickam, 2009. The pH measurement was determined by using a
103 Digital pH meter (Model HI9810, Hanna Ins.). The colour of mango juice samples was
104 measured using Hunter lab colour flex meter (M/s. Hunter lab, Reston, VA, USA; model

105 CFLX-45). The measurement was done with an illuminate observer combination of D65/10°.
106 The surface colour was quantified in terms of L*, a* and b* values of CIELAB colour space.
107 Data were recorded after 2, 4, 6 and 8 days of storage. Sensory evaluation was carried out by
108 ten untrained panelists based on acceptability, using the 9-point hedonic scale with a panel of
109 ten judges.

110 **Treatments**

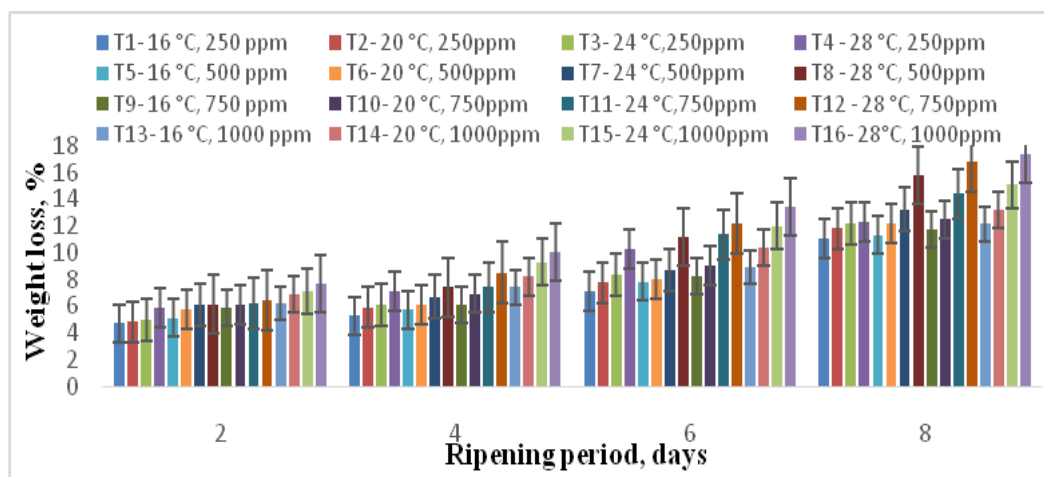
T1	Temp. - 16 °C, 250 ppm
T2	Temp. - 20 °C, 250 ppm
T3	Temp. - 24 °C, 250 ppm
T4	Temp. - 28 °C, 250 ppm
T5	Temp. - 16 °C, 500 ppm
T6	Temp. - 20 °C, 500 ppm
T7	Temp. - 24 °C, 500 ppm
T8	Temp. - 28 °C, 500 ppm
T9	Temp. - 16 °C, 750 ppm
T10	Temp. - 20 °C, 750 ppm
T11	Temp. - 24 °C, 750 ppm
T12	Temp. - 28 °C, 750 ppm
T13	Temp. - 16 °C, 1000 ppm
T14	Temp. - 20 °C, 1000 ppm
T15	Temp. - 24 °C, 1000 ppm
T16	Temp. - 28 °C, 1000 ppm

111

112 **Results and Discussion**

113 **Weight loss**

114 It was observed from Fig. 1 that the physiological loss in weight was significantly
115 increased with the increase of ethephon concentrations. The maximum weight loss (19.0%)
116 was observed in T15 on 8th day of storage whereas 10.5% in the control sample on the 11th
117 day of storage. Similarly, the loss in weight of fruit during storage both at ambient and in the
118 ripening chamber increased with the enhancement of storage days and was observed in pear
119 (Dhillon and Mahajan, 2011) and papaya (Singh *et al.*, 2012). Continuous processes of
120 respiration and transpiration have resulted in weight loss. Mangoes kept at higher
121 temperatures showed greater weight loss than those at lower temperatures. Results indicated
122 that the difference in total weight loss between ethephon treated and control sample increased
123 with increase in temperature. The higher weight loss at higher temperatures could be related
124 to the higher evapotranspiration rate and respiration rate at the higher temperatures as
125 previously reported by Lebibet *et al.* (1995).

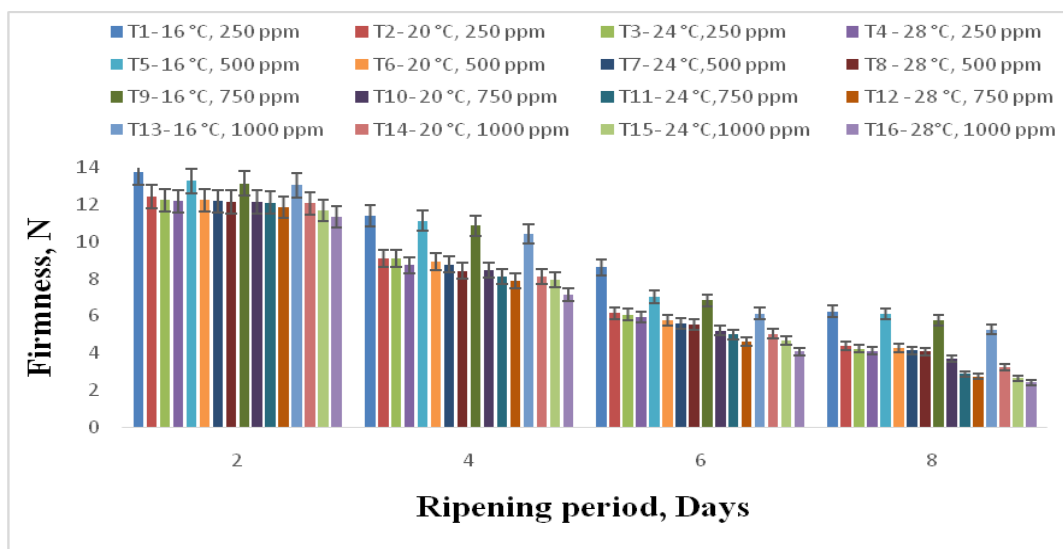


126
127 **Figure 1. Effect of different concentrations of ethephon at different temperatures on**
128 **weight loss during ripening of mango Cv. Neelum**

129 **Firmness**

130
131 It was observed from Fig.2 fruit firmness decreases with an increase in temperature
132 and concentration. At the initial stage, the firmness of mango fruit was found to be 15.83 N.
133 Fruit firmness decreased with an increase in ethephon concentration and also decreased with
134 days of storage. The maximum fruit firmness was observed in the control sample and
135 minimum in treatment, T15 ethephon.

136
137 The decrease in firmness, during ripening, may be due to the breakdown of insoluble
138 protopectin into soluble pectin or by cellular disintegration leading to membrane permeability
139 (Brinston *et al.*, 1988). The interaction between temperature and ethephon in firmness
140 showed that differences in firmness between ethephon treated and control mango fruits were
141 less at higher temperatures than at lower temperatures. The reason could be starch hydrolysis
142 or conversion of sugars.

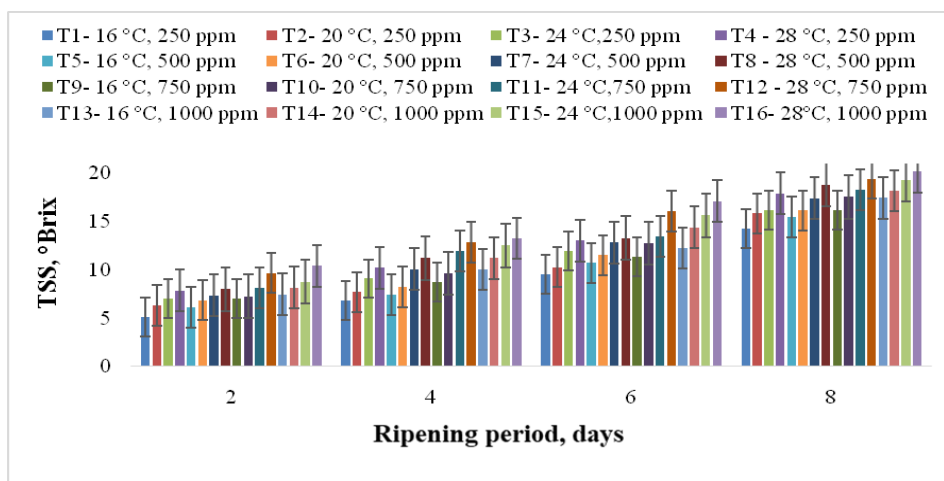


143
 144 **Figure 2. Effect of different concentrations of ethephon at different temperatures on**
 145 **firmness during ripening of mango Cv. Neelum**

146
 147 **Total Soluble Solids**

148
 149 It was observed from Fig. 3 the total soluble solids increased with an increase in
 150 ethephon concentrations and increased with days of storage. The highest value of TSS was
 151 observed in 1000 ppm (20.14 °brix) during 8 days of storage period. Ethephon treated
 152 mangoes possessed greater TSS than the control sample. The TSS might be increased due to
 153 hydrolysis of starch, cellulose and pectin substances into simpler substances or might be due
 154 to a decrease in moisture content. Similar results were also reported by Sakhale *et al.*, (2006).

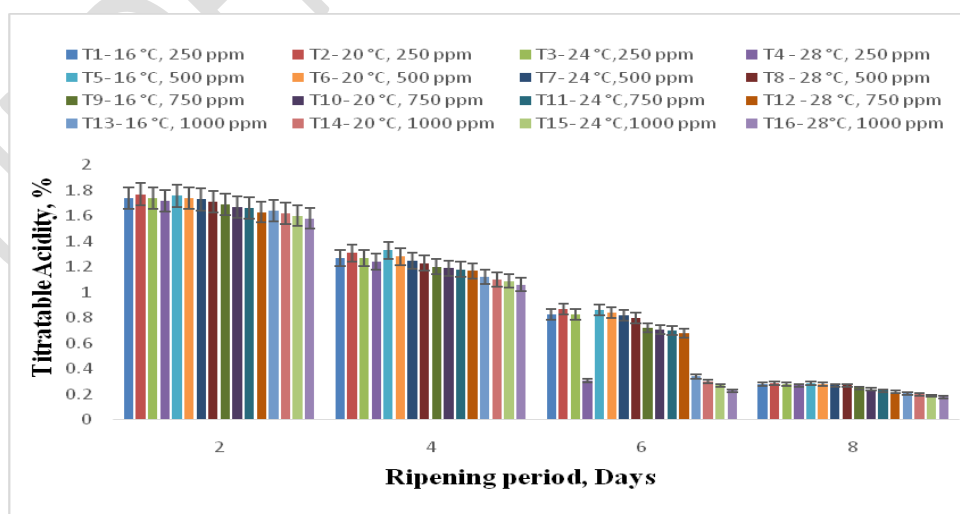
155
 156 Mangoes kept at higher temperatures showed greater TSS than those at lower
 157 temperatures. The increase in TSS was the outcome of the conversion of carbohydrates into
 158 simple sugars through a complex mechanism during the storage and the conversion rate was
 159 increased with the increase in temperature. This conversion is also considered to be one of the
 160 important indexes of the ripening process in mango and other climacteric fruit (Doreyappy-
 161 Gowda and Huddar, 2001; Kays, 1991; Kittur *et al.*, 2001).



162
163 **Figure 3. Effect of different concentrations of ethephon at different temperatures on**
164 **total soluble solids during ripening of mango Cv. Neelum**

165 **Titrateable acidity**

166 It is observed from the Fig.4 that acidity of the mango fruit was decreased by post
167 harvest application of ethephon and the response varied within the concentrations. A
168 maximum decrease (0.18%) in total acidity was found in T16 on 8th day of the storage
169 treatment. The result indicates that the acidity content declined significantly with the increase
170 in days of storage. Riberau-Gayon (1968) suggested that the transformation of organic acids
171 into sugars may be the reason for decreasing organic acids during fruit ripening. The
172 titrateable acidity was also decreased with the increase in storage temperature. The decrease in
173 acidity was attributed to the conversion of citric acid into sugars and their further utilization
174 in the metabolic process of the fruit (Doreyappy-Gowda and Huddar, 2001; Mizrach *et*
175 *al.*, 1997; Rathore *et al.*, 2007; Srinivasa *et al.*, 2002).



176
177 **Figure 4. Effect of different concentrations of ethephon at different temperatures on**
178 **titrateable acidity during ripening of mango Cv. Neelum**

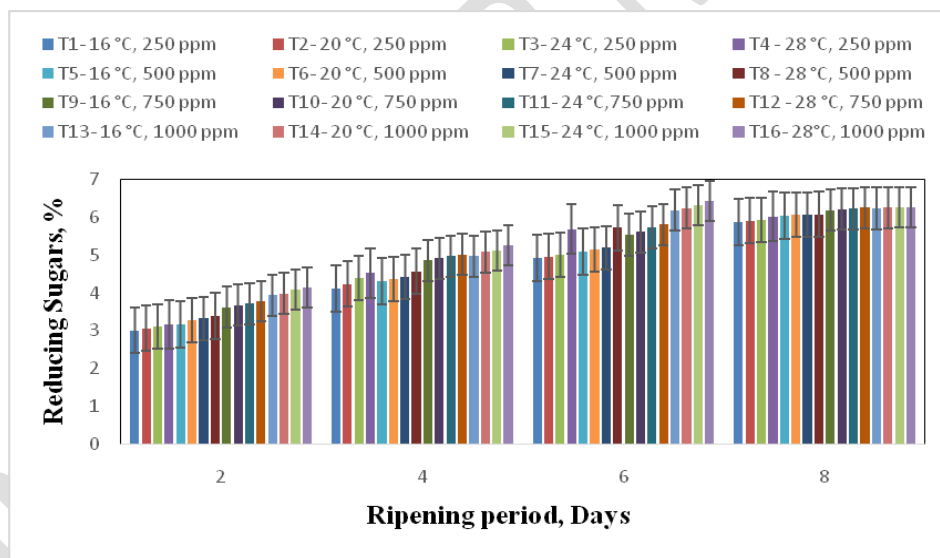
179 **Reducing sugars**

180

181 The results revealed that the sugar content increased with increasing ethephon
182 concentration and storage temperatures as shown in Fig. 5. The maximum reducing sugar
183 content (6.26%) was observed in T16 on 8th day of storage. The values of sugar content
184 increased up to 8 days. In the present experiment, ethephon enhanced the rate of
185 accumulation of reducing sugars in mango fruits. Similarly, a high percentage of sugar with
186 ethephon application was observed by Singh *et al.*, 2012 in mango. It was due to the faster
187 rate of respiration and the formation of sugar content with the oxidation of carbohydrates.

188

189 From the Fig. 5, it was also observed that the reducing sugar content was lowest for
190 samples stored at 16 °C and highest for samples stored at 28 °C. The increasing trend may be
191 the polysaccharides were converted into soluble sugar through a hydrolytic conversion
192 process, which was sensitive to temperature and/or to sunlight exposure for climacteric fruits
193 during the ripening process (Campestre *et al.*, 2002; Kays, 1991; Martinez *et al.*, 1997).



194

195

196 **Figure 5. Effect of different concentrations of ethephon at different temperatures on**
197 **reducing sugars during ripening of mango Cv. Neelum**

198 **Ascorbic acid**

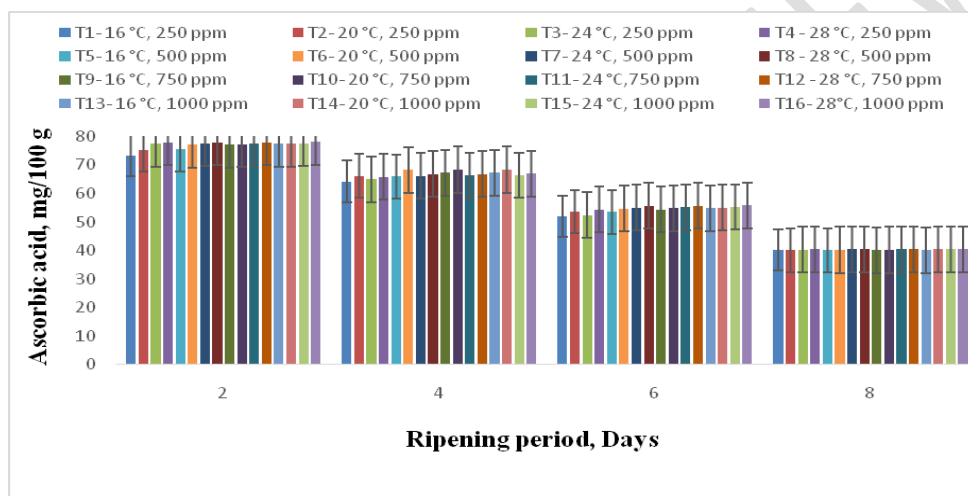
199 It was observed from Fig. 6 that the ascorbic acid increased with an increase in
200 ethephon concentrations and decreased with days of storage. The minimum value of ascorbic
201 acid content (40.08 mg/100 g) was observed in T1 on 8th day of storage. The values of
202 ascorbic acid content decreased up to 8 days. A decline in the ascorbic acid content of the

203 mango fruits might be due to the utilization of ascorbic acid in the respiration process during
 204 ripening at ambient conditions. A similar trend was also observed by Sakhale *et al.*, (2006),
 205 William *et al.*, (2009), Pandarinathan and Sivakumar (2010), in mango fruits.

206

207 Further, it is also observed that the ascorbic acid was decreased with the ripening of
 208 the fruit or with the increase in storage temperature. This trend was due to conversion of acid
 209 into sugars and their further utilization in metabolic process of the fruit and that the chemical
 210 and biological process was increased with the increase in storage temperature (Doreyappu-
 211 Gowda and Huddar, 2001; Mizrach *et al.*, 1997; Rathore *et al.*, 2007; Srinivasa *et al.*, 2002).

212



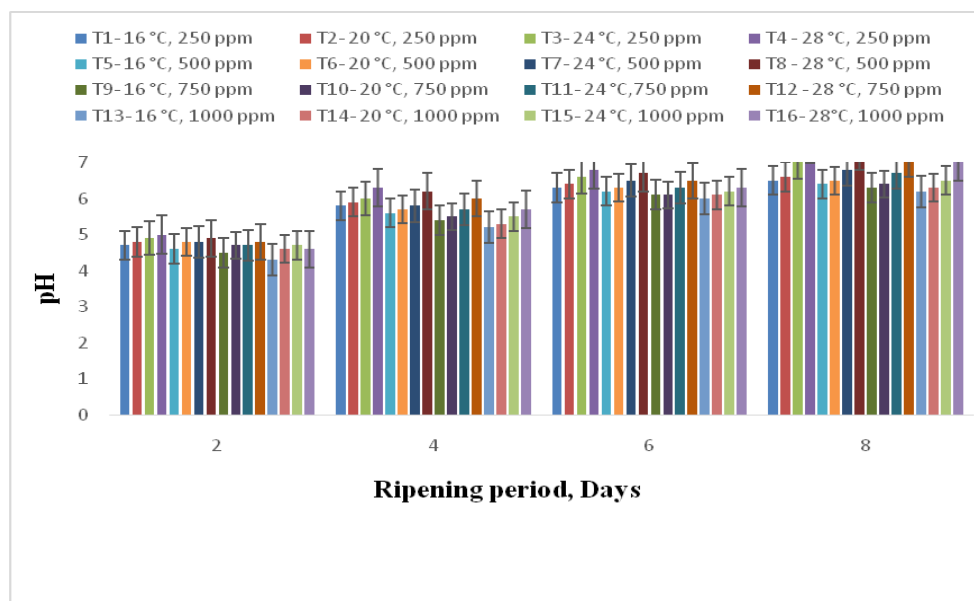
213

214 **Figure 6. Effect of different concentrations of ethephon at different temperatures on**
 215 **ascorbic acid during ripening of mango Cv. Neelum**

216

217 **pH**

218 The results revealed that the pH increased during ripening and decreased with an
 219 increase in ethephon concentrations. as shown in Fig.7. The pH value of the control sample is
 220 4.1. The pH value of mango Cv. Neelum fruits are in the range of 4.3-7.3. Also, results
 221 indicated that the pH increased with an increase in storage temperatures during the ripening
 222 of mango fruits. These observations were attributed to the conversion of citric acid and
 223 ascorbic acid into sugar and other products with the ripening process and whose rate of
 224 conversion was increased with the temperature (Absar *et al.*, 1993; Kumar and Singh, 1993;
 225 Rathore *et al.*, 2007; Yuniarti, 1980).



226

227 **Figure 7. Effect of different concentrations of ethephon at different temperatures on pH**
 228 **during ripening of mango Cv. Neelum**

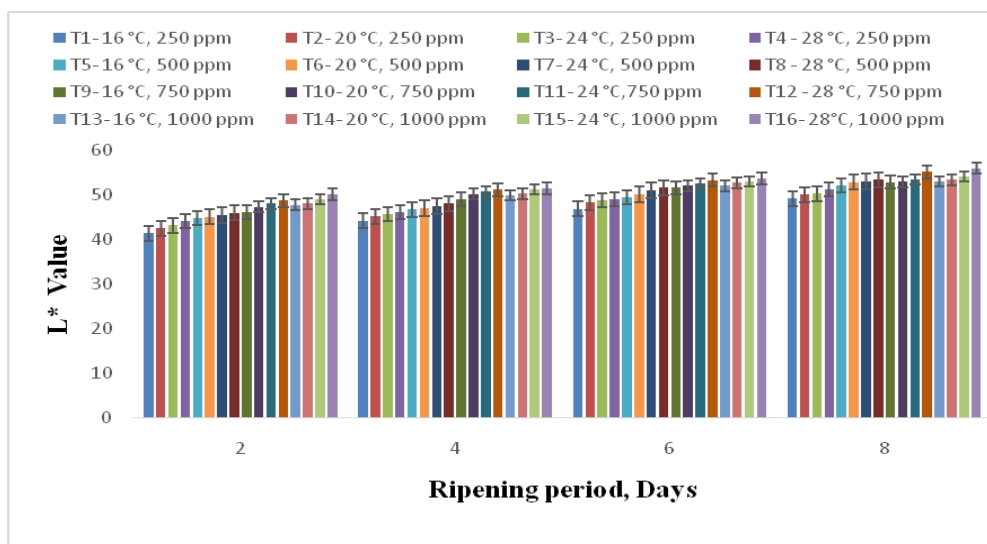
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230 **L* value for colour (Lightness)**

231

232

233 It was observed from Fig. 8 that the L* value for colour (Lightness) increases with
 234 increase in ethephon concentration as well as storage temperature during the enhancement of
 235 storage period in mango fruits. At the initial stage, the L* value of mango fruit was found to
 236 be 36.43. At the end of the 2nd, 4th, 6th and 8th day of storage, treatment T15 recorded the
 237 highest L* value (51.47, 52.72, 56.04). This is because of the fact that ethylene effects on the
 238 tissue and degreening of fruits. As the ethylene or ethephon triggered the ripening process,
 239 there is a rapid change in the colour from dark (green) to lightness (yellow) and it increased
 240 during the period of storage. Similar findings were reported by Daware (2012), Deepa and
 241 Preetha (2014) and Gill *et al.* (2015) in Dusehari mango fruits. Mangoes ripened at higher
 242 temperatures were significantly less green than those at lower temperatures.



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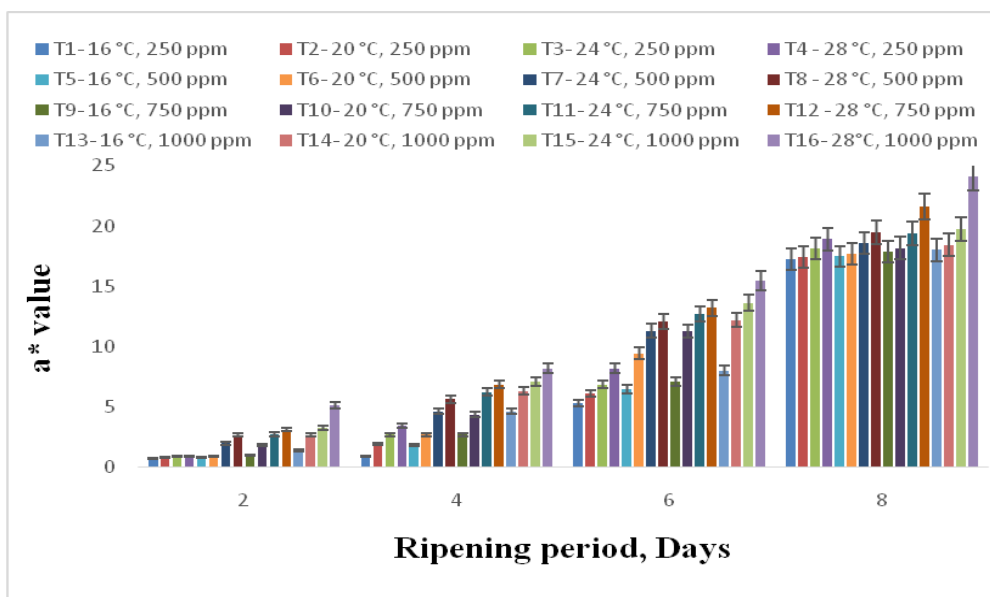
244 **Figure 8. Effect of different concentrations of ethephon at different temperatures on L***
 245 **during ripening of mango Cv. Neelum**

246 **a* and b* value (Redness/Greenness and Yellowness)**

247

248

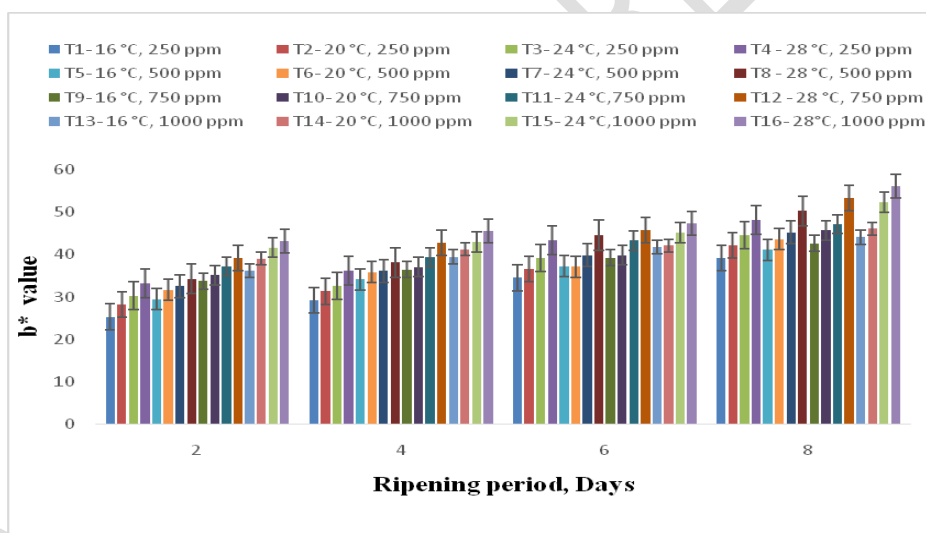
249 The results revealed that the a* and b* value increased during ripening with the
 250 increase in ethephon concentrations and storage temperatures as shown in Fig. 9 & 10. At the
 251 initial stage, a* and b* value of mango fruit was found to be -11.12 and 25.51. It was also
 252 observed that a* values for colour were less in untreated fruits in all days. This might be due
 253 to the breakdown of chlorophyll leading to the disappearance of green colour. During
 254 ripening, the peel colour changed from dark green to bright yellow and this is due to the
 255 change in chlorophyll which gradually unmasked the carotenoid pigments present in unripe
 256 mango fruits. As the ethylene triggered the ripening process, there was a rapid change in the
 257 colour from dark (green) to redness and it increased during the period of storage. Similarly,
 258 the b* values for colour were lower in untreated fruits i.e. control sample at all days. This
 259 might be due to changes during ripening period (loss of greenness, increase in redness and
 260 yellowness) occurred as a result of the breakdown of the chlorophyll in the peel. Similar
 261 findings were also reported by Daware (2012), Deepa and Preetha (2014), Gill *et al.* (2015) in
 mango fruit.



262

263 **Figure 9. Effect of different concentrations of ethephon at different temperatures on a***
 264 **value during ripening of mango Cv. Neelum**

265



266

267 **Figure 10. Effect of different concentrations of ethephon at different temperatures on**
 268 **b* value during ripening of mango Cv. Neelum**

269

270 **Organoleptic evaluation at ripening**

271

272 The data (scores) on the organoleptic evaluation of Neelum mango fruits are
 273 presented in Table 1. It could be revealed from the data that fruits ripened at ethephon, 750
 274 ppm, 20 °C temperature, 80% RH (T10) recorded maximum (8.50) sensory score in overall
 275 acceptability. The mango fruits ripened at ambient temperature (control) recorded the lowest
 276 6.25 as overall acceptability.

277

278 **Table 1 Effect of various treatments on organoleptic evaluation during ripening of**
 279 **mango Cv. Neelum fruits**

Treatments	Colour	Flavour	Taste	Texture	Overall acceptability
Control	6.00	7.00	6.00	6.00	6.25
T1	7.50	7.50	7.25	8.00	7.56
T2	7.50	7.50	8.00	8.00	7.75
T3	8.00	7.50	7.50	8.00	7.75
T4	8.00	8.00	7.50	8.00	7.88
T5	7.50	8.00	7.50	8.00	7.75
T6	7.50	7.50	7.50	8.00	7.63
T7	8.25	8.00	7.50	7.65	7.85
T8	7.65	8.25	8.00	8.00	7.98
T9	8.50	8.25	8.00	8.25	8.25
T10	8.50	8.75	8.50	8.25	8.50
T11	8.50	8.00	8.50	8.25	8.31
T12	8.50	8.75	8.50	8.25	8.50
T13	8.45	8.00	8.00	8.00	8.11
T14	7.65	8.25	8.00	8.00	7.98
T15	8.50	8.00	8.50	8.25	8.31
T16	8.50	8.25	8.00	8.25	8.25

280

281 **Conclusion**

282

283 It was found that mango Cv. Neelum ethephon dip treatment triggered the ripening
 284 process. It was also noticed that more the concentration of ethephon, faster was the ripening
 285 process and showed the significant increasing trends in L*, a*, b* values of colour and PLW
 286 (%) and decreasing trends in firmness (N) in all the treatments during advancement of the
 287 storage period in ripening chamber. It was observed that mango fruits Cv. Neelum ripened in
 288 ripening chamber in 8 days whereas fruits ripened at ambient conditions in 12 days.
 289 Similarly, mango fruits Cv. Neelum ripened by ethephon dip treatment of 750 ppm at 20 °C,
 290 80% RH for 5 minutes showed better results in respect of high overall acceptability score of
 291 8.50.

292

293 **REFERENCES**

- 294 Absar, N., Karim, M.R., Amin, M.A.L., 1993. A comparative study on the changes in the
 295 physic-chemical composition of ten varieties of mango in Bangladesh at different
 296 stages of maturity. *Bangladesh Journal of Agriculture Research*. 18, 201–208.
- 297 AOAC (1985) Official Methods of Analysis, Association of Official Analytical. Chemicals,
 298 AOAC, Benjamin Franklin, Station, Washington D.C.USA.
- 299 Brinston, K., Dey, P.M., John, M.A., Pridhan, J.B .(1988). Post-harvest changes in *Mangifera*
 300 *indica* mesocarp walls and cytoplasmic polysaccharides. *Phytochemistry*. 27:719–723.

- 301 Campbell, C.W. and Malo, S. E. (1969). The effect of 2-chloroethylphosphonic acid on
302 ripening of mango fruits. *Carib. Reg. Proc. Amer. Soc. Hort. Sci.*, 13 : 221-226.
- 303 Campestre, C., Marsilio, V., Lanza, B., Iezzi, C., Bianchi, G., 2002. Phenolic compounds and
304 organic acids change in black oxidized table olives. *ISHS Acta Hort.* 586: IV
305 *International Symposium on Olive Growing*. ISBN: 978-90-66057-56-2.
- 306 Dhillon, W.S. and Mahajan. B.V.C. (2011). Ethylene and ethephon induced fruit ripening in
307 pear. *Journal of Stored Products and Postharvest Research*, 2(3): 45-51.
- 308 Doreyappy-Gowda, I.N.D., Huddar, A.G., 2001. Studies on ripening changes in mango
309 (*Mangifera indica* L.) fruits. *Journal of Food Science and Technology Mysore* 38,
310 135–137.
- 311 Daware, P. M. 2012. Studies on period of exposure to ethylene gas in KKV fruit ripening
312 chamber on storage behavior of mango (*Mangifera indica* L.) Cv. Alphonso, Thesis
313 M Sc (Ag). Dr. Balasaheb Sawant Konkan Krishi Vidyapeeth, Dapoli, Maharashtra,
314 India.
- 315 Deepa, J. and P. Preetha. 2014. Influence of Exposure Time, Temperature and Ethylene
316 Concentration on The Ripening of Mango Fruits. *Trends in Biosciences* 7(20): 3260-
317 3267.
- 318 Gill, P.S., Jawandha, S.K., Kaur, N. and Verma, A. (2015). Changes in fruit colour of
319 Dushehari mangoes during ethephon induced ripening. *Int. J. of Agri. Envr. and*
320 *Biotechnol.*, 8 (1):97-101.
- 321 Kumar, P., Singh, S., 1993. Effect of GA₃ and ethephon on ripening and quality of mango CV.
322 Amarpali. *Horticulture Journal* 6, 19–23.
- 323 Kays, S.J., 1991. Post harvest Phys. of Perishable Plant Products. Van Nostr and Rein Hold
324 Book. AVI Publishing Co., pp. 149–316.
- 325 Kittur, F.S., Saroja, N., Habibunnisa, N., Tharanathan, R.N., 2001. Polysaccharide-based
326 composite coating formulations for shelf-life extension of fresh banana and mango.
327 *European Food Research and Technology* 213, 306–311.
- 328 Kulkarni, S.G., Kudachikar, V.B., Vasanta, M.S., Prakash, M.N.K., Prasad, B.A. and
329 Ramana, K.V.R . (2004). Studies on effect of ethephon dip treatment on ripening
330 behaviour of mango variety Neelum. *Mysore Journal of Food Science and*
331 *Technology*, 41:216-220.
- 332 Kader, A. A. (2002) *Post harvest technology of Horticultural Crops* 3rd edition Cooperative
333 Extension, Division of Agriculture and Natural Resources, University of California,
334 Oakland, California. Pub. 3311. P 535.
- 335 Lebibet, D., I. Metzidakis, D. Gerasopoulos, C.H. Olympios and H. Passam, 1995. Effect of
336 storage temperatures on the ripening response of banana *Musa*. sp. fruit grown in the
337 mild winter climate of Crete. *Acta Hort.*, 379: 521–6.
- 338 Mizrach, A., Flitsanov, U., Fuchs, Y., 1997. An ultrasonic non destructive method for
339 measuring maturity of mango fruit. *Transactions of ASAE* 40, 1107–1111.
- 340 Martinez, B.E., Guevara, C.G., Contreras, J.M., Rodriguez, J.R., Lavi, U., 1997. Preservation
341 of mango Azucar variety (*Mangifera indica* L.) at different storage stages:
342 *Proceedings of the fifth international mango symposium (Tel Aviv, Israel, 1996)*, 2,
343 pp. 747–754.

- 344 Mahajan, B.V.C., Kaur, T., Gill, M.I.S., Dhaliwal, H.S., Ghuman, B.S. and Chahil, B.S.
 345 (2010). Studies on rip-ening behaviour and quality of winter guava with ethylene gas
 346 ethephon treatments. *J. Food Sci. Technol.*, 45: 81–84.
- 347 Pandarinathan, S. and S.Sivakumar. 2010. Studies on biochemical changes in mangoes due to
 348 artificial ripening. *Int. J. Agri. Sci.* 1: 347-355.
- 349 PIB. 2017. [http://pibphoto.nic.in/documents/rlink/2017/aug/ p201783101.pdf](http://pibphoto.nic.in/documents/rlink/2017/aug/p201783101.pdf) dated 14.1.18
- 350 Ranganna S (1997) *Handbook of Analysis and Quality Control for fruit and vegetable*
 351 *Products*. 2nd edn. Pp 1112. Tata Mc.Graw Hill Pub.Co.Ltd., New Delhi, India.
- 352 Ribereau Gayon, G. (1968). Etudedes mechanisms synthese at de transformation delacide
 353 mailique,de l'acide tartique at de l'acide mailique,chaz *Vitis vinifera* L.
 354 *Phytochemistry*, 7:1471-1482.
- 355 Rathore, H.A., Masud, T., Sammi, S., Soomro, A.H., 2007. Effect of storage on physico-
 356 chemical composition and sensory properties of Mango (*Mangifera indica* L.) variety
 357 Dosehri. *Pakistan Journal of Nutrition* 6, 143–148.
- 358 Siddiqui, M.W. and Dhua R.S. (2009). Standardization of ethrel treatment for inducing
 359 ripening of mango var. Himsagar. International Congerence on Horticulture-2009,
 360 *Horticulture for Livelihood Security and Economic Growth*, November 09-12, 2009,
 361 Bangalore, India, 325pp.
- 362 Singh, J.P. and Mandal, B.K.(2000). Role of wrapper and post–harvest application of CaNO₃
 363 on the storage be-haviour of sub-tropical litchi cv. Manaraji. *Journal of Applied*
 364 *Biology*, 10(1): 37- 42.
- 365 Sakhale, B. K., V. N. Pawar, and B. M. Kapse. 2006. Effect of Ethrel on Hastening of Onset
 366 of Ripening in Kesar Mango (*Mangifera indica* L.). In Proceedings of 8th
 367 International Mango Symposium, February 2006, 820: 635-642.
- 368 Srinivasa, P., Baskaran, C.R., Ramesh, M.N., Prashantand, K.V.H., Tharanthan, R.N., 2002.
 369 Storage studies of mango Packed using biodegradable Chitosan film. *European Food*
 370 *Research and Technology* 215, 504–508.
- 371 Singh, P., M.K. Singh, M. Kumar, and S. Malik. 2012. Effect of physic-chemical treatment
 372 on ripening behaviour and post-harvest quality of Amrapali Mango (*Mangifera indica*
 373 L.) during storage. *J. Environ. Bio.*, 33: 227-232.
- 374 Thompson, A.K. and G.B. Seymour, 1982. Comparative effect of acetylene and ethylene gas
 375 on the initiation of banana ripening. *Ann. Appl.Biol.*, 101: 410.
- 376 William, O. A., O. Ibok, and O. E. William. 2009. Effect of Ethy-Gen II® ripening
 377 concentrate on ripening and sensory properties of mangoes (*Mangifera indica* L.).
 378 *Pakistan Journal Nutrition*, 8 (10):1641-1644.
- 379 Watada, L.E., Herner, R.C., Kader, A.A., Romani, R. I. and Staby, G.L. (1984). Terminology
 380 for the description of developmental stages of horticultural crops. *J. Amer. Soc. Hort.*
 381 *Sci.*, 19 : 20.
- 382 Yuniarti, 1980. Pysico-chemical changes of Arumanis mangoes during storage at ambient
 383 temperature. Bulletin Penelition Horticulture Indonesia 8, 11–17.
- 384