

4 **ACCEPTABILITY OF YOGHURT PREPARED FROM MILK SUBSTITUTED WITH**
5 **BENTH SEED (*Adenopus breviflorus*) PROTEIN ISOLATE**

6 ¹Ishaya F. A., ¹Onipede A. E. and ²Omowaye-Taiwo O. A.

7 ¹Science Technology Department, Federal Polytechnic, Ado Ekiti.

8 ²Food Technology Department, Federal Polytechnic, Ado ekiti.

9 Corresponding author e-mail: funmineforsuccess@yahoo.com

10 **Abstract**

11 Protein has been isolated from *Adenopus breviflorus* seed flour and used as a supplement to the
12 production of yoghurt. The chemical composition of the seed flour and protein isolate was
13 determined using standard methods. The yoghurt so produced was analysed for its pH, total
14 solids, titratable acidity and the sensory qualities evaluated using the nine point Hedonic scale
15 from dislike extremely (1) and like extremely (9). The results obtained from the chemical
16 composition of the seed flour and protein isolates showed that they are good sources of protein
17 (30.44% and 94.14% respectively). There was a significant reduction in the mineral content and
18 antinutrients of the seed flour after protein isolation. The result obtained from the sensory
19 evaluation showed that milk can be substituted for *Adenopus breviflorus* protein isolate from the
20 production of yoghurt up to 25% level of substitution without affecting the sensory qualities.

21 Keywords: *Adenopus breviflorus*, protein isolate, yoghurt

22 **1.0 Introduction**

23 *Adenopus breviflorus* Benth is a tree plant, commonly known as “*Lagenaria breviflora*
24 Roberty,” belongs to the family Cucurbitaceae (Yasuyuki, 2005). Different parts of the plant
25 (stem, seeds and fruits and leaves) have been used in folklore medicine in West Africa as herbal
26 treatment for various diseases in man and livestock (Ajayi *et al.*, 2002; Sonaiya, 1999; Tomori *et*
27 *al.*, 2007). In addition to its medicinal application, so much has been reported on the taxonomy
28 (Morimoto *et al.*, 2005) and chemical constituents of the plant. *Adenopus breviflorus* seed is
29 an oil seed commonly found in the savannah and semi - savannah forest region in southern
30 Nigeria. The seeds are used as soup ingredient. They can be ground with hulls or without hulls
31 for preparation of soup. It is also known that within some ethnic groups, the seeds are roasted
32 and eaten whole. Oshodi (1992) reported the proximate chemical composition, nutritionally
33 valuable minerals and functional properties of *Adenopus breviflorus* seed flour which could
34 be used to assess its value in the food industries other than direct consumption by farmers.

35 Protein–Energy–Malnutrition (PEM) is a serious problem facing most developing nations as a
36 result of inadequate intake of good quality protein from sources such as meat, fish and poultry
37 products, which are out of reach to **much** populace due to poor economy, increase **from**
38 population pressure and other natural calamities such as drought and flood (Nordeide *et al.*,
39 1996). In order to arrest this situation, much attention has been focused on the exploitation and
40 utilization of plants. Ordinarily, plants provided nearly two thirds of the world supply of food
41 protein for human and animals in which 10 – 15% comes from legumes (Pirman *et al.*, 2001;
42 Abegunde, 2018). Protein isolates are the most refined form of protein products containing the
43 greatest concentration of protein but unlike flour and concentrates contains no dietary fibre (Jay
44 and Michael, 2004). They are very digestible and easily incorporated into different food
45 products. Protein isolates are nowadays believed to have played a major role in the development
46 of new class of formulated foods. It is high concentration of protein with the advantage of colour,
47 flavour and functional properties making it an ideal raw ingredient for use in beverages, infant
48 foods and children milk food, **texture** protein products and certain types of specialty foods
49 (Olaofe, 1998).

50 Yoghurt is a fermented milk product obtained from milk or milk products by lactic acid
51 fermentation through the action of *Streptococcus thermophilus* and *Lactobacillus bulgaricus*
52 (FAO/WHO, 1977). When a sufficient quantity of lactic acid is produced then the milk
53 coagulates and this coagulated milk is called yoghurt. Lactic acid fermentation of legume based
54 milks has been used as one of the approaches to prolong the shelf life of the products, create
55 variety, improve the nutritional value and as well enhance the acceptability of the product. The
56 probiotic yoghurt, having probiotic effect is a fermented milk product **of** adjuvant
57 microorganisms. Yoghurts **varies from** appearance, flavor and ingredients. There is a symbiotic
58 relationship between the two species of bacteria, *Lactobacillus bulgaricus* and *Streptococcus*
59 *thermophilus*; that is why there is more rapid acid development than in the single strain culture
60 (**Tamime and Deeth, 1980**). Various combinations of starter cultures are selected during
61 manufacturing of yoghurt to achieve desirable characteristics of product and also to provide the
62 consumers with a wide choice of therapeutic benefits. Depending on its activity, manufacturer
63 usually adds 2-4 % yoghurt starter culture. Now a days, there has been increasing trends to
64 fortify the dairy product with fruits (natural fruit juice, pulp, dry fruits) (Ghadge *et al.*, 2008).
65 Yogurt-like products have been prepared by some workers from soybean, cowpeas, coconut and
66 mug beans (Terna and Musa, 1998).

67 This study seeks the possibility of using *Adenopus breviflorus* seed protein **isolates from**
68 substituting or fortifying **with** milk in yoghurt production.

69 **2.0 Materials and Methods**

70 **Collection and Preparation of Sample**

71 *Adenopus breviflorus* seeds used were obtained in the dried form, from a local market in Edo
72 State, Nigeria. The seeds were screened to remove stones, bad ones and dirt after which they

73 were de-hulled, dried and milled into powder. The powdered samples were stored in screw-
74 capped air-tight container.

75

76 **Preparation of Defatted Flour Sample**

77 Defatted sample of the seed flour was prepared by continuous extraction method using n-hexane
78 for nine hours in a soxhlet apparatus. The defatted flour was then recovered and residual solvent
79 was removed by air-drying.

80 **Preparation of Protein Isolate**

81 In preparation of the protein isolate, defatted seed flour, was dispersed in distilled water at a meal
82 ratio of 1:20 w/v (flour /water). The mixture was stirred with a stirrer for 30 minutes after which
83 the pH of the slurry was adjusted to the pH at which the protein in the flour is most soluble (pre-
84 determined) using 0.1M HCl drop wisely. The solution was further stirred for 2 hours at $30 \pm 2^{\circ}\text{C}$
85 using a stirrer to enhance high degree of protein solubility. The slurry was centrifuged at 4,000
86 rpm for 30 minutes at 4°C . The residue obtained after decanting the supernatant was re-extracted
87 with half the volume of the same solvent under similar conditions. The pH of the combined
88 supernatants was adjusted to the pH at which the protein in the flour is least soluble (the
89 isoelectric point which has been predetermined) with 0.1M HCl to precipitate the protein. The
90 isolate was recovered by centrifugation for 30 minutes at 4°C after which it was dispersed in
91 distilled water and dialyzed against distilled water for 18 hours. The dialysate was freeze dried
92 and then stored in air-tight container in the deep freezer for further analysis.

93 Proximate composition of the raw seed and protein isolate was determined according to the
94 standard method of AOAC (2005). Ca, Mg, Mn, Ni, Cd, Cu, Fe, Pb, Co, Zn and Cr were
95 determined using Atomic Absorption spectrophotometric method (Buck 210VGP model). Na and
96 K were determined by flame photometric method (Jenway PFP7 model) while phosphorus was
97 determined spectrophotometrically by the use of phosphovanadomolybdate solution (AOAC,
98 1990).

99 Various standard methods were used in determination of antinutritional composition of the raw
100 seed and protein isolate. The method of Young and Greaves (1940) as described by Reddy and
101 Kove (1999) was used to determine phytin. Tannin content was determined by modifying the
102 procedure of Makkar (1994). Oxalate was determined by the method described by Day and
103 Underwood (1986). Standard method of AOAC (2005) was used for the determination of
104 cyanide. Saponin was determined using the method of Obadoni and Ochuko (2001). Boham and
105 Kocipal-Abyazan (1994) method was used for the determination of flavonoid. Alkaloid was
106 determined according the method of Harborne (1993).

107

108 **Production of Yoghurt**

109 The milk and the protein isolates were weighed and mixed at different concentrations (25%,
110 50%, and 75%), while 100% milk was used as control sample. The mixtures were heated at 82 to
111 92⁰C for 30 minutes to pasteurize it and then cool about 45⁰C. The mixed starter culture was
112 added and stirred very well for complete mixing. The whole mixtures were allowed to stand in
113 an incubator for about 5 hours of 45⁰C. The mixtures were transferred into the refrigerator and
114 allowed to cool about thereby stopping further fermentation.

115 **Determination of pH of Yoghurt**

116 pH was measured using a pH meter (WTW-pH 330, Weilheim Germany).

117 **Determination of Titratable Acidity**

118 Acidity was determined by titrating the samples against 0.1M NaOH using phenolphthalein as
119 indicator.

120 **Determination of Total Solid of Yoghurt**

121 The weight of the residue obtained from moisture content analysis was expressed as percentage
122 total solid using the formula:

$$Total\ solid\ (\%) = \frac{weight\ of\ dry\ yoghurt}{weight\ of\ sample} \times 100$$

123

124 **Sensory Evaluation of Yoghurt**

125 Sensory evaluation was carried out on the samples of yoghurt by panel of 10 judges selected
126 from their consistency in scoring and the samples were evaluated for colour, taste, flavor
127 consistency and overall acceptability using the nine point Hedonic scale from dislike extremely
128 (1) and like extremely (9).

129 **Statistical Analysis of Data**

130 The data generated from all the results obtained were subjected to statistical analysis of variance
131 (ANOVA) using SPSS17 computer package and the mean values separated by Duncan's multiple
132 range test. Values reported are mean of triplicate determinations.

133

134

135

136 **3.0 Results**

137 **Table 1:** Proximate composition of raw seed flour and protein isolate (%)

Parameters	Moisture content	Fat	Crude protein	Ash	Crude fibre	Carbohydrate
Raw seed	4.05 ± 0.09	52.63 ± 0.33	30.44 ± 0.19	3.33 ± 0.11	3.15 ± 0.03	6.40 ± 0.31
Protein isolate	2.58 ± 0.04	1.21 ± 0.01	94.10 ± 0.02	1.08 ± 0.11	1.03 ± 0.06	ND

138 ND = not detected

139 **Table 2:** Mineral composition of raw seed flour and protein isolate (mg/100g)

Minerals	Raw seed	Protein isolate
Iron	12.32 ± 0.81	1.46 ± 0.04
Chromium	BDL	BDL
Nickel	BDL	BDL
Copper	0.39 ± 0.54	0.28 ± 0.02
Cobalt	BDL	BDL
Manganese	0.40 ± 0.30	0.07 ± 0.02
Zinc	21.32 ± 0.30	3.63 ± 0.01
Lead	BDL	BDL
Calcium	120.53 ± 0.62	25.16 ± 0.08
Magnesium	125.06 ± 0.11	18.51 ± 0.02
Potassium	129.62 ± 0.77	116.60 ± 0.09
Sodium	151.26 ± 0.31	86.43 ± 0.04
Phosphorus	164.94 ± 0.05	95.13 ± 0.03
Cadmium	BDL	BDL

140 BDL = below detection limit

141 **Table 3:** Antinutritional composition of raw seed and protein isolate

Parameters	Raw seed	Protein isolate
Phytate (mg/g)	11.74 ± 0.02	4.67 ± 0.02
Tannin (mg/100g)	0.14 ± 0.02	0.13 ± 0.01
Saponin (%)	1.32 ± 0.02	0.37 ± 0.03
Oxalate (mg/g)	1.33 ± 0.01	1.24 ± 0.02
Cyanide (mg/kg)	2.34 ± 0.02	0.66 ± 0.02
Alkaloid (%)	0.46 ± 0.02	0.14 ± 0.02
Flavonoid (%)	1.31 ± 0.01	0.26 ± 0.01

142

143

144

145 **Table 4:** General Notation of Yoghurt Samples Formulation

Samples	Dano milk (0% fat)	Protein isolate
Sample 1	25%	75%
Sample 2	50%	50%
Sample 3	75%	25%
Sample 4 (control)	100%	---

146

147

148 **Table 5:** The Result of Chemical Properties of Yoghurt Samples

Samples	pH	Titrateable Acidity (%)	Total Solid (TS) (%)
Sample 1	5.49	0.91	13.30
Sample 2	5.36	0.97	16.60
Sample 3	5.29	1.00	16.60
Sample 4	6.19	0.89	7.60

149

150 **Table 6:** Sensory Evaluation of Yoghurt

	Sample 1	Sample 2	Sample 3	Sample 4
Colour	3.80 ^b ± 2.20	2.80 ^b ± 1.14	7.00 ^a ± 1.15	7.70 ^a ± 1.06
Taste	3.80 ^b ± 2.39	2.70 ^b ± 1.83	6.50 ^a ± 1.43	7.40 ^a ± 1.35
Flavour	3.80 ^b ± 2.80	2.50 ^b ± 1.84	6.70 ^a ± 0.95	6.60 ^a ± 1.96
Consistency	2.60 ^b ± 1.78	2.40 ^b ± 1.71	6.80 ^a ± 1.23	7.80 ^a ± 1.03
Acceptability	2.50 ^b ± 1.78	2.80 ^b ± 1.75	7.30 ^a ± 0.82	8.20 ^a ± 0.92

151 Values with different superscript on the same row are significantly different ($p \leq 0.05$)

152 4.0 Discussion

153 The moisture content of the seed flour and protein isolates ranged from 2.58% to 4.05%. The
 154 moisture content of the seed flour is lower than that for similar legumes like *Cassia floribida*,
 155 6.0% (Vandavel and Janardhanam, 2001), *Lathyrus martimus*, 9.7% (Chavan *et al.*, 1999), *Lupin*

156 Species, 6.6% (Ruin-lopez *et al*, 2009). The moisture content obtained for the sample and isolate
157 are lower than the 10% recommended for storage stability of flours and this might be
158 advantageous in terms of prolonging the shelf-life and retaining their qualities. The moisture
159 content decreased significantly on isolation.

160 The high crude fat obtained (52.63%) suggests that it is an oil seed. The value, however,
161 compared very well with 47.9- 51.1% in *Citrullus vulgaris*, 47.02% in *flute* pumpkin and with
162 the range of 42.9- 57.3g/100g reported for some species of cucurbitaceae (Oshodi 1992; Ige *et*
163 *al*; 1984; Fagbemi and Oshodi 1991; and Fokou *et al*; 2004 respectively). There was reduction in
164 the value obtained for the protein isolate.

165 30.44% was recorded for the crude protein in the seed flour. This value is lesser than 49.8%
166 protein in soya *beans* reported by Osundahunsi and Aworh (2003). Higher protein content was
167 recorded in the protein isolate, 94.10 %. The value was higher than those reported for similar
168 legumes; mung beans isolate, 87.9% (Rahman *et al*, 2000), Chickpea isolate 88.1% (Sanchez *et*
169 *al*, 1999), *Canarvalia einsformis*, 73.3%, (Chel-guerrero *et al*, 2002). The variations in protein
170 contents of the different legume protein *isolate* are attributed to genetic make-up of legumes
171 along with some environmental factors (Kaur, 2007). The high protein content obtained in the
172 isolate suggests that it may be a better protein supplement than the seed flour and also contribute
173 significantly to alleviating the problem of protein malnutrition in the third world and developing
174 countries.

175 The total ash content *of* the seed flour, 3.33%, is comparable with some reported works (Oshodi
176 1992; Fokou *et al*, 2004; Aremu *et al*, 2006). The value is in excellent agreement with an
177 acceptable total range values for legumes which are between 2.4 -5.0% (FAO, 1989). There was
178 significant reduction in the ash content of the *isolates*.

179 The crude fibre content of the raw seed and protein isolate are generally low. The values
180 obtained are much higher than that of green *peas* (0.5- 0.93%). The results are in agreement with
181 those reported for mung *peas* and field pea (Naezk *et al*, 1986; Summer *et al*, 1981).

182 The mineral composition of the raw seed *flours* and protein isolate are presented in table 2. The
183 levels of phosphorus, sodium, potassium, magnesium and calcium were relatively high in the raw
184 seed flour than the isolate. The values of iron, zinc and copper were low while chromium, nickel,
185 lead and cadmium were not detected.

186 Though isolation decreased the values of these mineral elements, the potassium and phosphorus
187 *is* still high. The high calcium content has been reported to reduce blood pressure (Ranhotra *et*
188 *al*; 1998). Calcium is important *to* bone and teeth formation, blood clotting and in muscle
189 contraction. Magnesium in the blood as an activator of many enzyme systems and maintains the
190 electrical potential *for* nerves. Iron is important *to* blood formation and it is especially needed by
191 pregnant and lactating mothers. Manganese is an antioxidant nutrient that is important in the
192 breakdown of amino acids and the production of energy. The more important minerals involved

193 in the building of rigid structures to support the body i.e. Ca, P and Mg were more abundant in
194 the raw seed flour than the isolates. There may be need for **fortifications** of the isolates before
195 use.

196 The antinutritional composition of the raw seed and protein **isolates** are shown in tables 3. The
197 antinutrients were higher in the raw seed flour but on isolation they were significantly reduced.
198 This may be due to the procedures involved **in isolation**. Phytic acid might have been lost during
199 the process of isolation **from** the protein since it is soluble in water. It has been reported that
200 processing methods reduces the phytic acid of seeds **to the minimum levels** (Enujiugha and
201 Agbede, 2000). The concentration of tannin in this work is within the range obtained for
202 chickpea seeds, 0.07 % – 0.22 % (**Sharma et al., 2013**). Polyphenols and tannin are known to
203 inhibit digestive enzymes, and also reduce absorption of vitamins like B12 (**Liener, 1989**). They
204 also form complexes **about** Ca, Zn, Mg and Fe thereby reducing protein and mineral
205 bioavailability. Though tannin-protein complexes are insoluble in water and thus decreases
206 protein digestibility (Carnovale *et al*, 1987), the low concentration of tannin observed in this
207 work will have no nutritional significance.

208 The values of alkaloid, flavonoid, saponin and oxalate obtained are very low, therefore no
209 nutritional discomfort **are** expected. The low levels of the antinutrients reported **on** the isolates
210 are desirable from the functional and nutritional viewpoint and in the preparation of high quality
211 food products.

212 The percentage protein content of the protein **isolates** was found to be 94.10%. This showed that
213 the protein extracted is an isolate and not a concentrate. The results of the chemical properties of
214 four yoghurt samples produced are presented in table 5. The pH of the samples range from 5.29
215 to 6.19, addition of isolate reduced the pH of the yoghurt. This result is relatively high compared
216 to the result of Rodrigues *et al* (2010) who concluded that pH of yoghurt is 4.30 to 5.08. The
217 increase in pH of yoghurt may lead to short time of preservative of the yoghurt. The titratable
218 acidity of the four yoghurt samples ranges from 0.89 to 1.0. The addition of the isolate increased
219 the titratable acidity of the yoghurt. This result is in agreement with Younus *et al*, (2002) who
220 analysed the Quality evaluation of market yoghurt/ dahi and recorded 0.89 and 1.13 titratable
221 acidity. The percentage of the total solid ranges from 7.6 to 16.6 and the addition of the isolate
222 **increases** the total solid of the yoghurt. This content agrees with the findings of Muhammed *et al*,
223 (2005) who reported a higher total solid of 17.11%. However, Weaver (1993) reported that low
224 percentage of total solids in yoghurt can lead to malfunction of the starter culture.

225 The panelist evaluation indicated that there was no significant difference between the yoghurt
226 sample of up to 25% level of substitution and the yoghurt produced for 100% milk (0% fat) in all
227 parameters evaluated. This shows that at higher level of substitution (above 25%) the yoghurt
228 produced gets more undesirable to the panelist.

229

230

231 5.0 CONCLUSION

232 The result of chemical composition of the seed flours and protein isolates revealed that they are
233 good sources of proteins and carbohydrate. It also revealed the possibility of substituting the
234 conventional milk with protein isolates from *A. breviflorus* in the production of yoghurt up to
235 20% level of substitution without affecting the sensory qualities.

236 References

- 237 Abegunde S.M. (2018). Proximate Composition, Phytochemical Analysis and
238 Elemental Characterization of *Raphia taedigera* Seed, *Asian Journal of Chemical*
239 *Sciences* 5(2): 1-8. DOI: 10.9734/AJOCS/2018/45819
- 240 Ajayi, G. O., Awajo, N. C. and Abulu, L. E. (2002): Themiracidic and cercaricidal activity of
241 the methanolic extract of *Lagenaria breviflora* Roberty family Cucurbitaceae fruit in
242 *Schistosomamansoni*. *Niger Q J Hosp Med.*; 12: 57–59.
- 243 AOAC (1990): Official Methods of Analysis, 15th Edition. Washinton DC Association of
244 Official Analytical Chemists, pp 1250-1255.
- 245 AOAC (2005): International Official Methods of Analysis (18th edition). Washinton DC
246 Association of Official Analytical Chemist.
- 247 Aremu, M. O., Olaofe, O. and Akintayo, E. T. (2006): A comparative study of some Nigerian
248 underutilized legume flours. *Pakistan j. Nutri*; 5(1) 34-38.
- 249 Boham, B. A. and kocipal-Abyazan, R. (1994): Flavonoids and condensed tannins from leaves of
250 Hawairan.
- 251 Carnovale, E., Lintas, C. and Lombardi-Boccia, G. (1983). Effect of some anti-nutritional factors
252 on the *in vitro* protein digestibility of faba bean and pea. *Proc. Int. Congr. Food Sci.*
253 *Technol.*, 6th, Dublin, p12-23.
- 254 Chavan, U. D., Shadidi, F., Bal, A. K. and Mckenzie, D. B. (1999): Physico-chemical properties
255 and nutrient composition of beach pea (*Lathyrus matitimus* L.) *Food Chemistry*, 66, 43-
256 50.
- 257 Chel-Guerrero, L., Perez-Flores V., Bentacur-Ancona D. and Davila-Ortiz G. (2002): Functional
258 properties of flours and protein from *Phaseolus lunatus* and *Canavaliaen formis* seeds.
259 *Journal of agricultural and food Chemistry*, 50, 584-591.

- 260 Day, R. A. (Jnr) and Underwood, A. L. (1986): Quantitative analysis 5th edition, Prentice – Hall
261 publication, 704.
- 262 Enujiugha, V. A. and Agbede, J. O. (2000): Nutritional and Antinutritional characteristics of
263 African oil bean (*Pentaclethra macrophylla* benth) seed. *Applied Tropical Agriculture*,
264 5(1), 11-14.
- 265 Fagbemi, T. N. and Oshodi, A. A. (1991): Chemical composition and functional properties of
266 full fat fluted pumpkin seed flour (*Telfairia occidentalis*). *Nig Food J.* (9) 26-32.
- 267 FAO (1989): Utility of tropical foods. In Tropical beans. Food and Agricultural Organization
268 publication. 22-26.
- 269 FAO/WHO (1977): Report joint FAO/WHO expert committee on the code of principles
270 concerning milk and milk products. FAO/WHO, Rome
- 271 Fokou, E., Achu, M. B. and Tehounguel, F. M. (2004): Preliminary nutritional evaluation of five
272 species of egusi seeds in Cameroon, *African J. Food. Agric, Nut and Dev. Rural outr.*
273 Program 4(1) 1-7.
- 274 Ghadge, P. N., Prasad, K. and Kadam, P. S. (2008): Effect of fortification on the physic chemical
275 and sensory properties of buffalo milk yoghurt. *Electronic Journal of Environmental,*
276 *Agriculture and Food Chemistry* 7 2890-2899.
- 277 Harbone, J. B. (1993): Photochemical Methods, Chapman and Utal Ltd.
- 278 Ige M. M., Ogunsua A. O. and Oke O. I (1984): Functional properties of the proteins of some
279 Nigeria oil seeds, conophor seeds and three varieties of melon seeds *J. Agric. Food*
280 *Chem.* 32: 822-825.
- 281 Jay, R. H. and Michael, J. F. (2004): Macronutrient Utilization during Exercise: Implications for
282 performance and supplementation. *Journal of Sports Science and Medicine* 3: 118-130.
- 283 Kaur, S. (2007): Integrated Food Science and Technology for the Tropics. Macmillan Publishers
284 London .
- 285 Liener, I. E. (1989): Anti-nutritional factors in legume seeds: state of the art. In J. Huisman, T.
286 F., B. van der Poel, and I. E. Liener, (2nd edition). Recent Advances of Research in Anti-
287 nutritional factors in legume seeds. PUDOC, Wageningen, the Netherlands, pp 6-13.

- 288 Makkar, H. P. S. (1994): Anti-nutritional factors in foods for livestock, In: Gill M, Owen E,
289 Pollot GE, Lawrence TLJ. (Eds). Animal production in developing countries. *Occasional*
290 *publication, British Society of Animal Production*; 16: 69-85.
- 291 Morimoto, Y., Maundu, P., Fujimaki, H. and Morishima, H. (2005): Diversity of landraces of
292 the white flowered gourd (*Lagenariasiceraria*) and its wild relatives in Kenya: Fruit and
293 seed morphology. *Genet. Res. Crop Evol*; 52: 737-747.
- 294 Muhammed, B.F., Abubakar, M.M., Adegbola, T. A. and Oyawoye, E. O. (2005): Effects of
295 culture concentration and inoculation temperature on physicochemical, microbial and
296 organoleptic properties of yogurt. *Nig. Food J.*, 23: 156-165.
297
- 298 Naezk, M., Rubin, L. J. and Shaidi, L. J. (1986): Functional properties and phytate content of pea
299 protein preparations. *J. food sci.*, 51: 1245-1247.
- 300 Nordeide, M. B., Hatloy, A., Folling, M., Lied, E. and Oshaug, A. (1996): Nutrient composition
301 and Nutritional importance of green leaves and wild food resources in an Agricultural
302 district, koutiala, in southern Mali. *International Journal of Food Sciences Nutrition*.
303 47: 455-468.
- 304 Obadoni, B. O. and Ochuko, P. O. (2001): Phytochemical Studies and Comparative efficacy of
305 the crude extracts of some homeostic plants in Edo and Delta States of Nigeria. *global J.*
306 *Pure Applied science* 86: 203-208.
- 307 Olaofe, O., Arogundade, L.A., Adeyeye, E.I. and Falusi, O.M. (1998): Composition and food of
308 the variegated grasshopper. *Tropical Science* 38: 233-237.
- 309 Oshodi, A. A (1992): Proximate composition, nutritionally valuable minerals and functional
310 properties of *Adenopus Breviflorus* benth seed flour and protein concentrate. *Food*
311 *Chemistry* 45: 79-83.
- 312 Osundahunsi, O.F. and O.C. Aworh, (2003). Nutritional evaluation, with emphasis on the protein
313 quality of maize-based complementary foods enriched with soya bean and cowpea
314 Tempe. *International Journal of Food Science and Technology*, 38: 809-813.

- 315 Pirman, T., Stibily, V., Stekar, J.M.A. and Combe, E. (2001): Amino Acid Composition of Beans
316 and Lentil. *Zb. Biotech. Fak. Univ. Kmet. Zootech*; 78 (1): 57-68.
- 317 Rahma, E.H., Dudek, R.M, Gornitz,E. and Schwenke, K.D. (2000): Physicochemical
318 characterization of mung bean (*Phaseolus aureus*) protein isolates. *J.Sci. Food*
319 *Agric.*,80:477-483.
- 320 Ranhotra, G.S.,Gelroth, J.A, Leinen,S.O., Viners, M.A and Lorenz, K.J. (1998). Nutritional
321 profile of some edible plants from Mexico. *J. Food comp. and analysis*. 11: 298-304.
- 322 Reddy, M. B. and Kove, M. (1999): The impact of Food Processing on the Nutritional Quality of
323 Vitamins and Minerals. *Adv. Exp. Med. Bio.* 459: 99-106.
- 324 Rodrigues, L. A., Ortolani, M. B. T. and Nero, L. A. (2010): Microbiological quality of yoghurt
325 commercialized in Viçosa, Minas Gerais, Brazil, *African journal of microbiology*
326 *research* 4(3):210-213.
- 327 Ruiz-Lopez, M.A., Garcia-Lopez, P.M., Castañeda-Vazquez, H., Zamora, N.J.F., Garzón, P.,
328 Bañue-Los Pineda, J., Burbano, C., Pedrosa, M.M., Cuadrado, C. and Muzquiz, M.,
329 (2000): Chemical composition and antinutrient content of three *Lupinus* species from
330 Jalisco, Mexico. *J Food Compos Anal* 13, 193-199.
- 331 Sanchez-Vioque, R, A. Clemente, J. Vioque, J. Bautista, and F. Millan (1999): Protein isolates
332 from chickpea (*Cicera rietinum* L.): chemical composition, functional properties and
333 protein characterization. *Food Chem.* 64: 237-243.
- 334 Sharma, S., Yadav, N., Singh, A. and Kumar, R. (2013): Nutritional and antinutritional profile of
335 newly developed chickpea (*Cicer arietinum* L) varieties, *International Food Research*
336 *Journal* 20(2) p805-810.
- 337 Sonaiya, E. B. (1999): Family Poultry and Food Security. Research requirement in Science,
338 *Technology and Socio-Economics*.
- 339 Summer, A. K., Nielsen, M. A., and Youngs, C. G. (1981): Production and Evaluation of Pea
340 Protein Isolates. Presented at the 40th Annual Meeting of the Institute of food
341 Technologists, New Orleans.

- 342 Tamime, A. Y. and Deeth, H. C. (1980): Yoghurt: Technology and Biochemistry. *Journal of*
343 *Food Protection* **43**(12) 939-977.
- 344 Terna, G. and Musa, A. (1998): Soybeans Yogurt Production Using Starter Culture from
345 “Nono”. *Nigerian Journal Sof Biotechnology*, **9**, 17-23.
- 346 Tomori, O. A., Saba, A. B. and Dada-Adegbola, H. O. (2007): Antibacterial activity of ethanolic
347 extract of whole fruit of *Lagenaria brevipflora* Robertys. *J. Anim. Vet. Adv.*2007; 6: 752-
348 757.
- 349 Vandavel, V. and Janardhanan, K. (2001): Nutritional and Antinutritional attributes of the
350 underutilized legume *Cassia floribunda* car. *Food Chemistry* 73: 209-215.
- 351 Weaver, J. J. (1993). *Health Benefits of yoghurt*. Retrieved on July 06, 2008, from
352 www.Leaf lady.org/yoghurt/htm
- 353 Yasuyuki, M., Patrick, M., Hiroshi, F. and Hiroko, M. (2005): Diversity of landraces of the
354 white-floweredGourd (*Lagenariasiceraria*) and its wild relatives in Kenya. *Genet. Res.*
355 *Crop Eval*; 52: 737–747.
- 356 Young, S. M. and Greaves J. S. (1940): Influence of variety and treatment on phytin content of
357 wheat. *Food Res.*, 5: 103-105.
- 358 Younus, S., Aziz, T. and Masud, T. (2002): Quality Evaluation of Market Yoghurt /Dahi,
359 *Pakistan Journal of Nutrition* 1(5)