

# **Effects of Highly Active Antiretroviral Treatment on Complete Blood Count parameters**

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## **ABSTRACT**

**Aim:** This study assesses the effects of HAART on complete blood count parameters among HIV infected participants.

**Study design:** Case control study

**Place and Methods:** This study was conducted in Tamale, Ghana from August, 2015 to November 2017.

**Methodology:** A total of 300 HIV infected participants with ages ranging from 19–79 years, administered with HAART for at least 6 months were recruited. Pre-HAART administration (baseline) demographic and clinical information, with initial full blood count results were retrieved from the medical records of the participants. Post HAART administration blood sample (5mLs) was taken from each participant into an EDTA vacutainer tube and complete blood count (CBC) performed using URIT 5250 haematology analyser. Participants transfused with blood for the last 4 months were excluded from the study.

**Results:** The study recorded significant decreases in WBC and Neutrophil % post HAART administration. Lymphocyte (%), Haemoglobin, Haematocrit, MCV, MCHC, RDW-SD were all significantly higher post HAART administration. Total Platelets count, MPV, PDW-SD, PCT and P-LCR were significantly lower post-HAART administration. A comparison of the effects of EFV and NVP administered with AZT/3TC backbone yielded the following results. The NVP group recorded a significantly higher HCT compared with the EFV group (**p-0.0073**). A significantly higher mean PCT, MPV, P-LCR, PLCC, PDW-SD were recorded in the EFV group compared to the NVP group respectively.

**Conclusion:** The administration of HAART is associated with significant improvements in erythroid and lymphoid lineages, reduce anaemia, improves immunity and general patient well-being. NVP improve erythroid cell indices while EFV ameliorate platelet indices. HAART regimen should be chosen based on the pre-HAART laboratory tests conducted on the individual.

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*Keywords: Highly Active Antiretroviral Therapy, HIV infection, haematological abnormalities*

## **1. INTRODUCTION**

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HIV infection results in severe systemic disorder, with characteristic impairment and progressive damage to both humoral and cellular immune responses (Brenchley et al., 2006). Haematological abnormalities may be the first laboratory findings in HIV infection and may involve all cell lines and noted to be strong independent predictors of morbidity and mortality in HIV infected persons (Calis et al., 2008, Kibaru et al., 2015). Calis and colleagues reported a prevalence of 3-38% moderate anaemia (Hb 8.0-9.9g/dL) and 50-91% mild anaemia (Hb10.0-12.0g/dL) among people HIV infected individuals in tropical areas, which increases as the disease progresses and varies with age, gender and the definition used to establish anaemia (Calis et al., 2008). The causes of anaemia in HIV infected individuals included decreased erythropoietin production, ineffective erythropoiesis,

26 opportunistic infections, neoplasia and micronutrient deficiency (Thulasi et al., 2016). Iron  
27 deficiency has also been reported as the commonest cause of nutritional anaemia among  
28 HIV infected individuals (Kibaru et al., 2015). The type of anaemia serves as a guide to the  
29 choice of HAART and treatment option for opportunistic infections. Neutropenia was  
30 reported in 10% more of HIV infected individual with advanced immunosuppression than in  
31 non-advanced HIV infected individuals (Consolini et al., 2007, Adetifa et al., 2006).  
32 Asymptomatic thrombocytopenia was reported in 20%-33% of HIV infected individuals but  
33 increased with progression of the disease (Karpatkin et al., 2002, Adetifa et al., 2006).  
34 Kibaru et al. (2015) reported that the use of zidovudine, lamivudine and stavudine was  
35 associated with significant amelioration in hemoglobin concentration and after 12 months of  
36 HAART use, the prevalence of anaemia reduced from 65.5 % to 46% (Kibaru et al., 2015). In  
37 another study, after 3 months of HAART administration, Huang et al. (2000) documented  
38 significant increases in mean Hb from 13.9 to 14.1 g/dl (Huang et al., 2000). Huang et al.  
39 (2000) reported improvements in mean cell volume (MCV) from 55 to 98.9fl to 105.5fl and  
40 106fl at 3, 6, 9, 12 months respectively after HAART use (Huang et al., 2000).  
41 The current regimens used in Ghana comprise two nucleotide reverse transcriptase  
42 inhibitors (NRTI) plus one nonnucleotide reverse transcriptase inhibitor (NNRTI) or two NRTI  
43 and one protease inhibitor (PI) (Daugas et al., 2005). The WHO recommends HAART for all  
44 HIV infected individuals since it improves morbidity and mortality associated with HIV  
45 infection, and offers better life expectancy (Aboulafia et al., 2000, Ajayi et al., 2009). Despite  
46 the beneficial effects associated with HAART use, studies have shown that steps must be  
47 taken to prevent life threatening side effects and HAART related haematotoxicity (Nubila et  
48 al., 2012). In resource constrained developing countries, based on availability, safety and  
49 efficacy, the WHO has made efavirenz, tenofovir in addition to lamivudine or emtricitabine  
50 plus nevirapine as the preferred first line antiretroviral medications, though so much  
51 information is not available on HAART use in these areas (WHO, 2016). Nevirapine remains  
52 the NNRTI of choice when efavirenz cannot be used. Better rates of HIV replication  
53 suppression is achieved in patients administered with efavirenz-based HAART who received  
54 tenofovir-emtricitabine compared with patients who received zidovudine-lamivudine (Darin  
55 et al., 2010). In order to avoid the toxicities associated with Zidovudine use, especially in  
56 individuals with anaemia, global access to tenofovir has been increased (Nii-Trebi et al.,  
57 2013). Not many studies on the haematological parameters abnormalities in HIV infected  
58 persons have been conducted in sub Saharan Africa. This study examines the effect of  
59 HAART on complete blood count parameters.

## 61 **2. MATERIAL AND METHODS / EXPERIMENTAL DETAILS / METHODOLOGY**

### 62 **Study Design**

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64 This was a cross sectional study carried out from 12<sup>th</sup> August 2016 to 21<sup>st</sup> December 2017.

### 65 66 **Study population**

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68 A total of 300 HIV infected participants with ages ranging from 19 to 79years and have been  
69 on HAART for at least 6 months were recruited. Participants who have been transfused with  
70 blood for less than 4 months were excluded from the study. **The sample size was arrived at  
71 using the cochran's formula with a national HIV prevalence of 1.6% (Raima Carol, 2018).**

### 72 73 74 **Data Collection**

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76 Pre-HAART administration (baseline) information such as weight, systolic and diastolic blood  
77 pressure, age, date of HAART initiation and initial complete blood count results were  
78 retrieved from the medical records of the participants. Post HAART information such as age,

79 weight, blood pressure measurement, HAART type and duration of HAART use were  
80 recorded using questionnaire which had been pre-tested among 10 HIV infected individuals  
81 administered with HAART to clear possible ambiguity and difficulty in answering the  
82 questions. Data from the pre-tested questionnaires were however not included in the results  
83 analysis.

84 Blood samples (5mLs) was taken into an EDTA vacutainer for Complete Blood Count (CBC)  
85 analysis. The CBC was performed using URIT 5250, a 5-part differential, 28 parameters  
86 haematology analyser, from URIT Medical Electronic (group) Co., LTD, China,  
87 <http://www.urit.com/index.aspx>. The HAART comprised two Nucleotide Reverse  
88 Transcriptase Inhibitor (NRTI) plus one Nonnucleotide Reverse Transcriptase Inhibitor  
89 (NNRTI) or two NRTIs and a Protease Inhibitor (PI). The participants were further stratified  
90 into short term (<52 months), medium term (≥52 but ≤104months), long term (>104months)  
91 based on the duration of HAART.

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### 93 **Statistical analysis**

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95 Data was entered into Microsoft excel 2016 and exported to GraphPad prism version 6.0  
96 ([www.graphpad.com](http://www.graphpad.com)) for analysis. Data was presented as number, percentages, means and  
97 standard deviation. Means were compared between groups using Student's paired t-test and  
98 ANOVA and  $p < 0.05$  was considered statistically significant.

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## 101 **3. RESULTS AND DISCUSSION**

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103 Table 1 shows the demographic data of the study population. Out the 300 participants,  
104 majority (80.7%) were females, 176 (58.7%) were short-term HAART users, 96 (32%) were  
105 medium-term HAART users while 28 (9.3%) were long-term HAART users.

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**Table 1: Demographic characteristics of the study population**

<b>Variables</b>	<b>Pre-HAART (n=300)</b>	<b>Post-HAART (n=300)</b>
Age (years)	35.4 ± 9.4	<b>39.7 ± 10.0</b>
Weight (Kg)*	58.8 ± 12.8	<b>64.3 ± 25.6</b>
<b>Age Group (years)</b>		
10-19	10 (3.3%)	3 (1%)
20-29	76 (25.3%)	36 (12%)
30-39	125 (41.7%)	120 (40%)
40-49	64 (21.3%)	96 (32%)
50-59	20 (6.7)	34 (11.3%)
60-69	5 (1.7%)	13 (4.3%)
70-79	0	1 (0.3%)
<b>Gender</b>		
Male	58 (19.3%)	58 (19.3%)
Female	242 (80.7%)	242 (80.7%)
<b>HAART Duration (Months)</b>		
Short term	0	176 (58.7%)
Medium term	0	96 (32%)
Long term	0	28 (9.3%)

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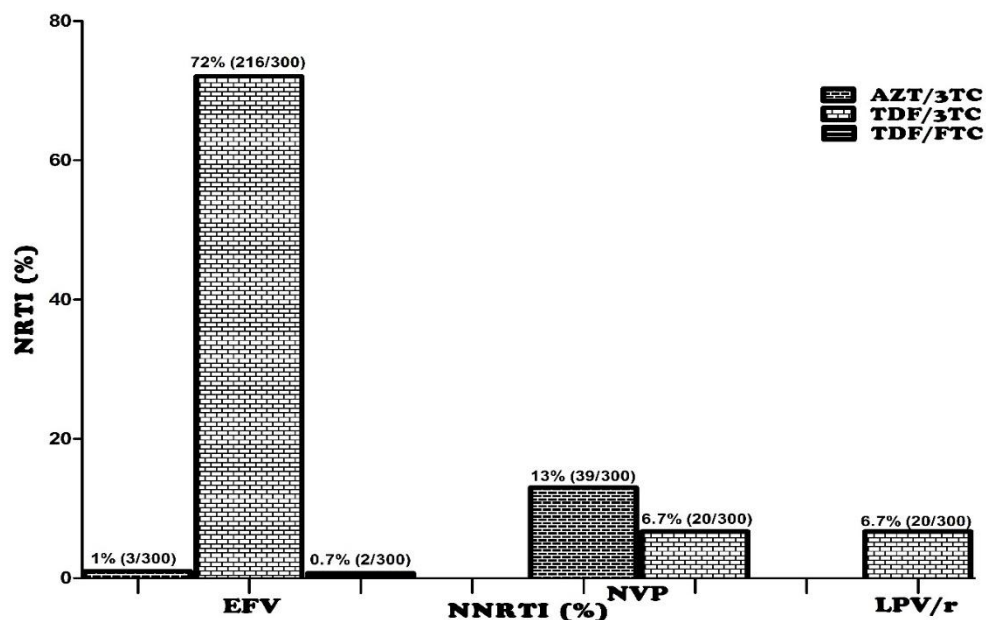
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*Data presented as mean ± SD, frequency (percent), \* - comparison between pre and post HAART administration using paired t-test. P-values < 0.05 considered significant.*

### **Distribution of HAART in the Study Population**

113 HAART regimen was distributed as follows, TDF+3TC+EFV (72%), AZT+3TC+NVP (13%),  
 114 TDF+3TC+NVP (6.7%), TDF+3TC+LPV/r (6.7%), AZT+3TC+EFV (1%) and TDF+FTC+EFV  
 115 (0.7%) regimen.



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 117 **Fig. 1. Percentage distribution of the various types of HAART regimen administered at**  
 118 **the ART centers**  
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120 Table 2 compared complete blood count (CBC) parameters in the study population pre and  
 121 post-HAART administration. White blood cells (WBC) (%) 7.446±5.066 vrs 4.733±2.008,  
 122 neutrophil (%) 51.52±18.37 vrs 44.26±14.46, basophils (%) 4.0±0.2 vrs 3.9±0.06 were  
 123 significantly higher in pre-HAART administration while lymphocyte (%) 33.11±14.05 vrs  
 124 43.94±12.87 increased significantly post-HAART administration.

125 Haemoglobin (g/dL) 10.88±2.255 vrs 11.63±2.401, haematocrit (%) 32.75±6.239 vrs  
 126 34.85±6.932, mean cell volume (MCV) 81.96±10.77 vrs 88.0±12.96, mean cell haemoglobin  
 127 concentration (MCHC) 33.52±1.651 vrs 34.50±5.368, red cell distribution width-standard  
 128 deviation (RDW-SD) 44.26±11.78 vrs 68.69±42.64 were significantly higher post-HAART  
 129 administration (Table 2).

130 Again, thrombocytopenia was significantly pronounced post-HAART administration, with  
 131 platelet (PLT) 284.7±148.1 vrs 254.4±145.7, mean platelet volume (MPV) 9.028±1.773 vrs  
 132 6.840±1.601, platelet distribution width (PDW) 14.55±6.180 vrs 8.881±4.37, plateletcrit  
 133 (PCT) 0.260±0.140 vrs 0.169±0.085 and platelet-large cell ratio (P-LCR) 28.87±9.586 vrs  
 134 20.60±15.12 significantly decreased post-HAART administration (Table 2).

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136 **Table 2: Complete blood count tests parameters pre and post HAART administration**

Parameters	Pre-HAART (n-300)	Post-HAART (n-300)	P – value
White blood cell count (10 <sup>9</sup> /L)	7.5 ± 5.1	4.7 ± 2.0	< .00
Lymphocytes %	33.1 ± 14.1	43.9 ± 12.9	< .00
Monocytes %	10.0 ± 4.9	9.8 ± 5.7	.31
Neutrophils %	51.5 ± 18.4	44.3 ± 14.5	.00
Eosinophils %	4.2 ± 3.7	3.7 ± 3.4	.63
Basophils %	0.4 ± 0.2	0.08 ± 0.06	.00
Red Blood Cell Count (10 <sup>12</sup> /L)	4.0 ± 0.9	3.9 ± 0.7	.79
Haemoglobin (g/dL)	11.0 ± 2.6	11.6 ± 2.4	.04

Haematocrit (HCT, %)	32.8 ± 6.3	<b>34.9 ± 6.9</b>	<b>.01</b>
Mean Cell Volume (MCV, fL)	82.0 ± 10.8	<b>88.0 ± 13.0</b>	<b>.00</b>
Mean Cell Haemoglobin (MCH, pg)	29.4 ± 18.1	30.5 ± 5.3	.78
MCHC (g/dL)	33.5 ± 1.7	<b>34.5 ± 5.4</b>	<b>.03</b>
RDW-SD (fL)	44.3 ± 11.8	<b>68.7 ± 42.6</b>	<b>.00</b>
RDW-CV (%)	13.9 ± 4.9	15.7 ± 10.7	.13
Platelets (10 <sup>9</sup> /L)	<b>284.7 ± 148.1</b>	254.4 ± 145.7	<b>.00</b>
MPV (fL)	<b>9.0 ± 1.8</b>	6.8 ± 1.6	<b>&lt; .00</b>
PDW (fL)	<b>14.6 ± 6.2</b>	8.9 ± 4.4	<b>&lt; .00</b>
Platelet crit (PCT, %)	<b>0.3 ± 0.1</b>	0.2 ± 0.1	<b>&lt; .00</b>
P-LCR (%)	<b>28.9 ± 9.6</b>	20.6 ± 15.1	<b>&lt; .00</b>
P-LCC (10 <sup>9</sup> /L)	72.8 ± 45.7	48.4 ± 27.7	0.32

137 *Student's paired t-test comparison of complete blood count parameters before and after*  
 138 *HAART administration. Data presented as mean ± SD and P-values <0.05 considered*  
 139 *significant.*

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141 Table 3 shows the influence of duration of HAART usage on CBC parameters. Neutrophil  
 142 (%) and basophils (%) decreased significantly (p<0.05) from short-term HAART users,  
 143 medium-term through to long-term HAART users. The significant reduction in neutrophil (%)  
 144 was from 46.36±14.26 in short-term HAART users to 39.53±15.13 in long-term HAART  
 145 users while basophils (%) were from 0.065±0.049 to 0.053±0.032 to 0.002±0.001 in short-  
 146 term, medium-term to long-term HAART users respectively.

147 However, lymphocytes (%), haemoglobin concentration, haematocrit, mean cell volume  
 148 (MCV) and mean cell haemoglobin (MCH) increased significantly (p<0.05) with duration on  
 149 HAART uses. Haemoglobin concentration increased significantly from 11.31±2.394,  
 150 12.25±2.172 and 12.93±7.352 in short-term, medium-term and in long-term HAART users  
 151 respectively. While for lymphocytes, the significant increased were between 42.13±13.18 in  
 152 short-term to 48.74±11.50 in long-term HAART users and from 46.03±12.04 in medium-term  
 153 to 48.74±11.50 in long-term HAART users. For haematocrit, there was a significant  
 154 increased between 33.93±5.945 in short-term to 36.49±8.026 in medium-term HAART users  
 155 only but for MCV and MCH, significant increased were observed between short-term  
 156 (86.56±13.02fL and 29.60±5.132ng) and median-term (91.20±12.43fL and 31.64±5.612ng)  
 157 HAART users as well as short-term (86.56±13.02fL and 29.60±5.132ng) and long-term  
 158 (92.25±14.67fL 33.04±5.172ng) and HAART users respectively.

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161 **Table 3 Complete Blood Count based on the duration of HAART usage**

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PARAMETER	DURATION OF HAART			F test	P - value
	Short-term n-176	Medium-term n-96	Long-term n-28		
White blood cell count (10 <sup>9</sup> /L)	4.7 ± 1.6	4.5 ± 1.7	4.9 ± 2.0	0.8	.46
Lymphocytes %	42.1 ± 13.2	46.0 ± 12.0 <sup>+</sup>	48.7 ± 11.5 <sup>*</sup>	5.1	<b>.01</b>
Monocytes %	9.4 ± 5.5	10.7 ± 6.4	10.7 ± 8.3	1.7	.18
Neutrophils %	46.4 ± 14.3	41.7 ± 14.0	39.5 ± 15.1 <sup>*</sup>	5.0	<b>.01</b>
Eosinophils %	3.4±2.8	3.4 ± 2.6	3.0±2.8	0.1	.91
Basophils %	0.07 ± 0.05 <sup>@</sup>	0.05 ± 0.03 <sup>+</sup>	0.002 ± 0.001 <sup>*</sup>	11.4	<b>&lt; .00</b>
Red Blood Cell Count (10 <sup>12</sup> /L)	4.0 ± 0.7	4.0 ± 0.6	3.748 ± 0.7	1.2	.29
Haemoglobin (g/dL)	11.3 ± 2.4 <sup>@</sup>	12.3 ± 2.2 <sup>+</sup>	12.9 ± 7.4 <sup>*</sup>	4.9	<b>.01</b>
Haematocrit (HCT, %)	33.9 ± 6.0 <sup>@</sup>	36.5 ± 8.0	35.6 ± 8.1	4.5	<b>.01</b>

Mean Cell Volume (MCV, fL)	86.6 ± 13.0 <sup>@</sup>	91.2 ± 12.4	92.3 ± 14.7 <sup>*</sup>	5.2	.01
Mean Cell Haemoglobin (MCH, pg)	29.6 ± 5.1 <sup>@</sup>	31.6 ± 5.6	33.0 ± 5.2 <sup>*</sup>	7.9	.00
MCHC (g/dL)	34.4 ± 6.0	34.8 ± 4.8	35.0 ± 3.7	0.3	.76
RDW-SD (fL)	65.7 ± 13.4	66.9 ± 12.6	68.9 ± 11.8	0.8	.46
RDW-CV (%)	14.7 ± 5.1	14.2 ± 2.8	15.4 ± 7.1	0.5	.78
Platelets (10 <sup>9</sup> /L)	251.1 ± 102.9	254.8 ± 213.4	227.8 ± 82.2	0.5	.61
MPV (fL)	6.9 ± 1.6	6.8 ± 1.7	6.8 ± 1.7	0.3	.72
PDW (fL)	9.0 ± 4.5	8.8 ± 4.5	8.0 ± 2.7	0.7	.52
Platelet crit (PCT, %)	0.2 ± 0.1	0.19 ± 0.13	0.16 ± 0.07	0.5	.61
P –LCR (%)	20.3 ± 9.5	19.2 ± 8.2	21.2 ± 9.4	0.7	.49
P –LCC (10 <sup>9</sup> /L)	50.6 ± 30.1	44.4 ± 22.2	47.8 ± 26.3	1.6	.20

163 *One-way ANOVA comparison of Complete Blood Count parameters in short, medium and*  
164 *long-term HAART users. Data presented as mean ± SD. (<sup>@</sup>) - comparison between short*  
165 *and medium term, (<sup>\*</sup>) - comparison between short and long term, (<sup>†</sup>) - comparison between*  
166 *medium and long term HAART users. P-values < 0.05 considered significant.*  
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168 Table 4 considered the effects of EFV and NVP administered with a common backbone  
169 AZT/3TC. Haematocrit 32.50±5.328 vrs 33.87±5.559 was significantly higher in the NVP  
170 group compared with EFV group. However, PCT 0.263±0.124 vrs 0.158±0.062, MPV  
171 8.667±1.290 vrs 6.479±1.502, P-LCR 34.70±9.930 vrs 17.32±8.894, P-LCC 111.0±78.89 vrs  
172 41.64±21.54 and PDW-S 12.97±3.083 vrs 7.567±2.255 were significantly higher in the EFV  
173 group compared with the NVP group. The EFV group had a more favourable effect on  
174 platelet indices compared with the NVP group. Comparatively, there were no significant  
175 variations in the effects of EFV, LPV/r and NVP administered with a TDF/3TC back bone on  
176 CBC parameters  
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188 **Table 4 Effects of EFV, NVP and LPV/r with common backbones (AZT/3TC and TDF/3TC)**

PARAMETER	AZT/3TC BACKBONE			TDF/3TC BACKBONE			
	EFV GOURP	NVP GROUP	P - value	EFV GROUP	LPV/r GROUP	NVP GROUP	P - value
White blood cell count (10 <sup>9</sup> /L)	3.9±1.6	4.6±1.4	.38	4.7±1.7	5.2±2.5	5.1±4.3	.39
Lymphocytes %	35.5±27.0	45.8±13.5	.24	43.8±12.9	44.1±11.3	43.7±10.4	.99
Monocytes %	2.7±1.9	3.6±2.7	.59	3.8±3.7	1.7±0.9	3.7±1.9	.28
Neutrophils %	0.07±0.02	0.06±0.03	.66	0.06±0.04	0.05±0.02	0.08±0.06	.43
Eosinophils %	8.9±2.0	10.1±6.2	.73	9.5±5.2	10.9±7.7	10.8±7.3	.39
Basophils %	55.3±25.7	41.8±17.1	.21	44.6±14.0	43.4±13.1	44.1±13.0	.93
Red Blood Cell Count (10 <sup>12</sup> /L)	3.5±0.3	3.5±0.7	.98	4.0±0.7	4.0±0.7	4.01±0.6	.97
Haemoglobin (g/dL)	10.5±2.4	11.8±2.08	.33	11.6±2.4	11.86±2.5	11.4±3.5	.86
Haematocrit (HCT, %)	32.5±5.3	<b>33.9±5.6</b>	<b>.01</b>	34.6±5.7	34.9±7.5	39.8±15.45	.01
Mean Cell Volume (MCV, fL)	92.6±13.5	97.4±13.6	.55	87.0±12.1	87.0±13.9	92.0±13.3	.22
Mean Cell Haemoglobin (MCH, pg)	30.1±6.6	34.3±5.4	.21	29.9±4.8	29.4±4.3	31.4±8.1	.39
MCHC (g/dL)	32.1±2.7	35.2±3.7	.17	34.5±5.5	33.6±4.4	33.8±7.8	.68
RDW-SD (fL)	71.9±9.8	72.9±13.9	.90	68.7±49.7	64.7±11.9	65.2±12.2	.89
Platelets (10 <sup>9</sup> /L)	321.3±182.0	242.9±71.6	.11	254.4±161.9	267.3±112.5	247.1±112.5	.91
Platelet crit (PCT, %)	<b>0.3±0.1</b>	0.2±0.1	<b>.01</b>	0.17±0.09	0.2±0.1	0.2±0.1	.51
MPV (fL)	<b>8.7±1.3</b>	6.5±1.5	<b>.02</b>	6.8±1.6	7.1±1.7	6.7±2.0	.75
P –LCR (%)	<b>34.7±9.9</b>	17.3±8.9	<b>.00</b>	21.0±17.0	21.7±8.3	18.8±6.9	.81
P –LCC (10 <sup>9</sup> /L)	<b>111.0±78.9</b>	41.6±21.5	<b>.00</b>	46.9±23.0	55.3±30.2	46.7±28.1	.33
PDW-S (fL)	<b>13.0±3.1</b>	7.5±2.3	<b>.00</b>	9.0±4.5	10.2±6.9	8.6±2.0	.46

189 *Comparison of the individual influences of EFV and NVP (with AZT/3TC backbone) was done using Student unpaired t-test whilst ANOVA*  
 190 *was used to compare between groups of TDC/3TC backbone on Complete Blood Count parameters (CBC). Data presented as mean ± SD*  
 191 *and P-values < 0.05 were considered significant.*

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195 Results in Table 6 shows the effects of AZT and TDF administered with 3TC/EFV or  
 196 3TC/NVP. In comparing AZT with TDF (administered with 3TC/EFV), P-LCC 111.0±78.89  
 197 vrs 46.88±22.97 was significantly higher in the AZT group compared with the TDF group. In  
 198 comparing the AZT group with TDF group (administered with 3TC/NVP), RBC 3.525±0.651  
 199 vrs 4.008±0.614 and HCT 33.87±5.559 vrs 39.80±15.48 were significantly higher in the TDF  
 200 group compared with the AZT group. However, RDW-SD 33.87±5.559 vrs 39.80±15.48 was  
 201 significantly higher in the AZT group compared with the TDF group.

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203 **Table 5 Comparison of the individual effects of AZT and TDF (administered with**  
 204 **3TC/EFV and 3TC/NVP) on Complete Blood Count parameters**  
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PARAMETER	3TC/EFV			3TC/NVP		
	AZT	TDF	P = value	AZT	TDF	P = value
WBC	3.9±1.6	4.7±1.7	.42	4.6±1.4	5.1±4.3	.52
Lymphocyte (%)	35.5±27.0	43.8±12.9	.28	45.8±13.5	43.7±10.4	.55
Eosinophil (%)	2.7±1.9	3.8±3.7	.64	3.6±2.7	3.7±2.0	.94
Basophil (%)	0.07±0.03	0.06±0.04	.81	0.06±0.03	0.08±0.05	.22
Monocyte (%)	8.9±2.0	9.6±5.2	.83	10.1±6.2	10.8±7.3	.71
Neutrophil %	55.3±25.7	44.6±14.0	.20	41.8±17.1	44.1±13.0	.61
RBC	3.5±0.3	4.0±0.7	.20	3.5±0.7	<b>4.0±0.6</b>	<b>.01</b>
HBG	10.5±2.4	11.6±2.4	.42	11.8±2.1	11.4±3.5	.65
HCT	32.5±5.3	34.6±5.7	.53	33.9±5.6	<b>39.8±15.5</b>	<b>.04</b>
MCV	92.6±13.5	87.0±12.1	.43	97.4±13.6	92.0±13.3	.16
MCH	30.1±6.6	29.9±4.8	.93	34.3±5.4	31.4±8.1	.11
MCHC	32.1±2.7	34.5±5.5	.44	35.2±3.7	33.8±7.8	.36
RDW-CV	16.2±1.6	14.0±7.5	.61	13.9±3.5	14.4±3.0	.60
RDW-SD	71.9±9.8	68.7±49.7	.91	<b>72.9±13.9</b>	65.2±12.2	<b>.04</b>
PLT	321.3±182.0	254.4±161.9	.48	242.9±71.6	247.1±112.5	.85
PCT	0.3±0.1	0.2±0.1	.05	0.2±0.1	0.2±0.1	.33
MPV	8.7±1.3	6.8±1.6	.05	6.5±1.5	6.7±2.0	.57
P - LCR	34.7±9.9	21.0±17.0	.17	17.3±8.9	18.8±6.9	.52
P - LCC	<b>111.0±78.9</b>	46.9±23.0	<b>&lt; .00</b>	41.6±21.5	46.7±28.1	.45
PDW-S	13.0±3.1	9.0±4.5	.12	7.6±2.3	8.6±2.0	.10

206 **Comparison of the effects of AZT and TDF (administered with 3TC/EFV and 3TC/NVP) on**  
 207 **Complete Blood Count were done using Student's unpaired t-test. Data presented as mean**  
 208 **± SD and P - values < 0.05 considered significant.**  
 209

210

## 211 Discussion

212

212 The findings in this study show that, the percentage of short, medium and long-term HAART  
 213 users increased gradually from 9.3% in long-term HAART users through 32.0% in medium-  
 214 term HAART users to 58.7% in short-term users. This reflects a gradual increase in the  
 215 number of individuals enrolled on HAART. This shows that more HIV infected individuals are  
 216 being recruited unto HAART. This results are in agreement with the findings of Atuyambe et  
 217 al. (2008) and Nii-Trebi et al. (2013) who also recorded an increased number of HIV infected  
 218 individuals on HAART.

219 The mean duration of HAART is denoted by the mean difference between Post HAART age  
 220 and pre-HAART age. The participants have been on HAART for a long duration as the post-  
 221 HAART age is significantly higher than the pre-HAART age. This shows that the participants  
 222 have been on HAART long enough for HAART to elicit its effect on CBC parameters.



223 There was a significant increase in the body weight of the study population post HAART  
224 administration. This indicates an improvement in the general health status of the individuals  
225 or the absence of frequent illness in the study population as a result of immune restoration  
226 indicated by improvement in lymphocyte count. Guillén et al. (2007) and Newell (Newell et  
227 al., 2003) reported increases in the body weight of people living with HIV post HAART  
228 administration.

229 The study found significant decline in total WBC post HAART administration and a  
230 consistent decline in neutrophil and basophil counts from short-term through to long-term  
231 among post HAART users post HAART administration. This indicates gradual development  
232 of severe leucopenia with time. Similarly, in a study by Nacoulma et al. (2007), leucocyte  
233 counts decreased from  $2.27 \times 10^3 \text{mm}^3$  at baseline to  $1.9 \times 10^3 \text{mm}^3$  after six months of AZT  
234 administration. Leucopenia may be largely due to neutropenia caused by HIV suppression of  
235 the bone marrow, leading to ineffective granulopoiesis (Kimura et al., 1990, Firnhaber et al.,  
236 2010). Kimura et al. (1990) observed neutropenia may also be caused by the presence of  
237 antigranulocyte antibodies which attack and destroy granulocytes. Many of the HAART  
238 drugs are considered as myeloid suppressive, especially Zidovudin, hence should be  
239 reviewed for patients who already have cytopenias (Akinbami et al., 2010). The leucopenia  
240 in the present study may possibly be due to the presence of inflammation in the tissues as a  
241 result of exposure to the drug. There is direct leucopenic inhibitory effect on matured  
242 granulocytes in the peripheral system or on the myeloid progenitor cells in the bone marrow.  
243 Consolini et al. (2007) reported leucopenia after six months of AZT based HAART  
244 (Gallicchio et al., 1994, Ukoha et al., 2015, Consolini et al., 2007). However, the findings of  
245 this study contradicted the findings of [3] Kibaru and his colleagues who did not record  
246 leucopenia post HAART administration (Kibaru et al., 2015). This was attributed to the  
247 presence of acute and recurrent bacterial infection which sustained the production  
248 granulocytes (Kibaru et al., 2015).

249 Lymphocyte count post HAART administration increased significantly in a time dependent  
250 manner. This is accounted for by the steady-state equilibrium in favour of endogenous cell  
251 provisions over virus mediated cell killing. In the tap and drain model proposed by Wei et al.  
252 (1995), decreased viral killing of lymphocytes quickly turn the balance in favour of  
253 lymphocyte production and survival which allows at least partial immune reconstitution to  
254 occur. This signifies a shrinkage in the myeloid lineage in favour of the lymphoid lineage  
255 (Amegor et al., 2009). Many other studies also found an increase in lymphoid tissue activity  
256 after HAART administration (Baker et al., 2007, Grossman and Herberman, 1997, Mohri et  
257 al., 1998). Studies suggest that, the observed increase in lymphoid activity was a response  
258 from the immune system following HAART administration (Baker et al., 2007, Grossman and  
259 Herberman, 1997, Mohri et al., 1998).

260 This study also recorded significant improvements in erythroid cell line and indices. HGB,  
261 HCT, MCV and RDW-SD were significantly higher post HAART administration in a time  
262 dependent manner. The present study findings are consistent with studies by (Thulasi et al.,  
263 2016) and (Sullivan et al., 1998) who also found increases in red cell indices post-HAART  
264 administration. These findings are also consistent with the findings of Ogunbusuyi (2015)  
265 who reported the prevalence of anaemia as higher in treatment naïve patients compared  
266 with those on HAART. Other studies reported a decline in the erythroid lineage due to the  
267 toxic effect of medications on the bone marrow, nutritional deficiency especially B12, iron  
268 deficiency, decreased erythropoietin release, gastrointestinal bleeding, malabsorption,  
269 autoimmune antibodies to haemopoietic precursors etc (Schmaier and Lazarus, 2011) while  
270 other studies attribute the increased incidence of anaemia to the direct effect of HIV on bone  
271 marrow stroma as a prelude to bone marrow failure (Akinbami et al., 2010).

272 Thrombocytopenia was recorded as an early haematological abnormality in HIV infected  
273 individuals in sub-Saharan Africa (Munyazesa et al., 2012). The incidence of  
274 thrombocytopenia post HAART administration in this study was significant. This finding is  
275 consistent with the findings of Akinbami et al. (2010) who also recorded higher incidence of

276 thrombocytopenia post HAART administration. The possible explanation may be due to  
277 increased platelet destruction by the deposition of immune complexes on platelets and  
278 decreased platelet production, which results in decreased MPV and plateletcrit (Amegor et  
279 al., 2009). Thrombocytopenia is also a consequence of HIV's direct infection and destruction  
280 of megakaryocytes (Kasturi et al., 2006). Amegor et al. (2009) reported that  
281 thrombocytopenia increased as immunological incompetence of the participants worsens.  
282 The presence of anti-platelet antibodies which leads to an increased destruction of platelets  
283 has also been hypothesized (Thulasi et al., 2016). However, the results of this study is not in  
284 line with the findings of a study conducted in Kenya which recorded a decrease in  
285 thrombocytopenia from 20% to 6.5% after 6 months of ART (Enawgaw et al., 2014, Kibaru et  
286 al., 2015).

287 The predominant HAART regimen in this study was the TDF+3TC+EFV which is the WHO  
288 recommended alternate first line HAART regimen in some resource limited areas (Wester et  
289 al., 2009, WHO, 2003). Majority (72%) of the study population were on TDF+3TC+EFV, 13%  
290 were on AZT+3TC+NVP (which is the standard WHO recommended first line HAART  
291 regimen in resource limited settings), while 6.7% were on TDF+3TC+NVP which is also an  
292 alternate first line regimen (Wester et al., 2009, WHO, 2003). TDF+3TC+LPV/r was the  
293 commonest second line HAART regimen recorded in this study. This implies that 6.7% of the  
294 HIV infected individuals experienced treatment failure and have switched to a second line  
295 therapy. Also, 0.7% of the study population were administered with TDF/FTC/EFV, which is  
296 an alternate first line HAART regimen in resource rich areas. According to the WHO  
297 guidelines on HAART issued in 2003, the formulation of first line HAART includes the  
298 combination of AZT/3TC plus stavudine (d4T) or NVP or EFV (WHO, 2003). In 2006, this  
299 recommendation was revised to include TDF or ABC as alternative first-line NRTIs while  
300 encouraging health care givers not to include (d4T) base combinations to minimize its  
301 possible accumulation in the mitochondria leading to d4T associated complications (Wester  
302 et al., 2009). Majority of the regimen in this study included AZT, 3TC and either EFV or NVP  
303 or LPV/r, depending on the availability of the regimen at the ART centers.

304 Comparison of lymphocyte values in NVP and EFV groups showed higher counts in the NVP  
305 group compared with the EFV group. These findings are consistent with findings of a Kenyan  
306 study which recorded higher lymphocyte values in NVP group compared to the EFV group  
307 (Naluande, 2017). The higher lymphocyte count signifies restoration of immunity following  
308 HAART administration. Comparison of AZT and TDF, both administered with a 3TC/NVP  
309 shows that RBC and HCT were both higher in the TDF group. AZT/3TC/NVP had better  
310 result on erythroid cell indices compared to AZT/3TC/EFV. The worst decrease in both  
311 erythroid and myeloid cell lines were recorded in the AZT/3TC/EFV. This demonstrated the  
312 compensatory effect of NVP over AZT when administered together, and showed AZT's  
313 toxicity on progenitor cells in the bone marrow stroma (Akinbami et al., 2010). This was  
314 consistent with a study by Hema, 2011 who also found NVP more favourable to erythroid cell  
315 indices compared with AZT or EFV.

316 The AZT/3TC/EFV group recorded the highest platelet large cell count (PLCC). This shows  
317 increased variation in the sizes of thrombocytes. This corroborates with the findings of a  
318 study conducted by Munyazesa et al. (2012) in sub-saharan Africa who found increased  
319 variation in platelet sizes post HAART administration. From the findings of this study, a  
320 comparison of EFV and NPV administered with AZT/3TC backbone showed that EFV  
321 improved platelet indices as opposed to NVP which did not improve platelet indices. This  
322 may probably be the reason why the 2013 WHO HIV treatment guidelines recommended  
323 EFV as the first line choice of NNRTIs (<http://www.who.int/hiv/pub/guidelines/arv2013>).  
324 Thrombocytic indices; PLT, PCT, MPV, PLCR, P-LCC and PDW-S were all higher in the  
325 EFV group compared with the NVP group. The results of this study show that EFV has an  
326 ameliorative influence on platelet indices compared to NVP.

327 There was no significant difference in the effect of AZT and TDF administered with 3TC/EFV  
328 on complete blood count parameters. A comparison of AZT and TDF administered with

329 3TC/NVP shows a more positive influence on erythroid cell lines in the TDF group compared  
330 with the AZT group. Again, in this study TDF/FTC/EFV which is the gold standard first line  
331 regimen in resource rich area had the best effect on platelet indices (Wester et al., 2009).

332

## 333 CONCLUSION AND RECOMMENDATION

334

335 In conclusion, the administration of HAART is associated with significant improvements in  
336 erythroid and lymphoid lines, which reduces anaemia, improves immunity and general  
337 patient well-being. Cumulatively, NVP has a much more significant improvement on erythroid  
338 cell indices while EFV significantly improves ameliorated platelet indices. As found in this  
339 study, NVP favoured lymphoid lineage growth compared with EFV. TDF favoured RBC  
340 indices compared with AZT and EFV improved platelet indices as opposed to NVP which did  
341 not improve platelet indices. AZT/3TC/NVP, TDF/3TC/LPV/r, TDF/3TC/NVP should be  
342 considered for improvement in erythroid cell indices. AZT/3TC/EFV and TDF/FTC/EFV  
343 should be considered in patients with thrombocytopenia. This makes HAART a suitable  
344 choice taking into account side effects in the management of people living with  
345 HIV to prevent the development of AIDS especially when started early. This study  
346 therefore provides added basic information to encourage health workers to  
347 intensify the effectiveness of HAART campaign to get HIV infected individuals to  
348 initiate HAART early. Also, ART should be chosen base on the results of the initial  
349 laboratory tests conducted on the patient.

350

351

## 352 COMPETING INTERESTS

353

354 Authors have declared that no competing interests exist.

355

356

## 357 CONSENT

358

359 A consent was sought from each participant before being included in the study. Consent  
360 form was given to each participant to sign or thumb-print and confidentiality was assured.  
361 Subjects who did not give their consent were excluded from the study. A copy of the written  
362 consent is available for review by the Editorial office of your journal.

363

## 364 ETHICAL CONSIDERATIONS

365

366 Ethical clearance was sought from by the Committee for Human Publication and Research  
367 Ethics of the Kwame Nkrumah University of Science and Technology, Kumasi Ghana.

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499

## 500 DEFINITIONS

501 Short term HAART users = Participants who have been on HAART for less than or equal to  
502 52 months.

503

504 Medium term HAART users = Participants who have been on HAART for more than 52  
505 months but less than or equal to 104 months.

506

507 Long term HAART users = Participants who have been on HAART for more than 104  
508 months.