

## **Optimization of Cellulose Production by *Curvularia pallescens* Isolated from Textile Effluent**

### **Abstract:**

**Introduction:** Celluloses are important industrial enzymes and find application in several industrial processes. Effects of pH, temperature, incubation time, source of carbon and nitrogen were tested in submerged fermentation process in the production of cellulose by *Curvularia pallescens* isolated from textile effluent.

**Aims:** The present study was attempted in a fungus; *Curvularia pallescens* isolated from textile effluent for maximizing its production under optimal conditions in submerged fermentation by using inexpensive substrate wheat bran.

**Study design:** The production medium was prepared in distilled water, supplemented with 4.5% wheat bran, 0.05% KCl, 0.2% KH<sub>2</sub>PO<sub>4</sub>, (carbon source), yeast extract (nitrogen source), maintained with pH of 5.5 and incubated at 28<sup>o</sup>C for 120h was found optimal for the production of cellulose.

**Results:** The test fungus achieved maximum FPA activity followed by cellobiohydrolase, endoglucanase and β-glucosidase activity at 46.76, 42.06, 26.94 and 3.56 U/ml respectively at pH 5.5 (figure-4). The temperature of 28<sup>o</sup>C produced maximum cellulase activity. Highest activity recorded was of FPA (38.94 U/ml), followed by cellobiohydrolase (30.29 U/ml), endoglucanase (22.41 U/ml), and β-glucosidase (3.98 U/ml). The effect of process parameters such as the effect of temperature, pH and inoculum size was investigated. Maximum cellulase and xylanase having an enzyme activity of 694.45 and 931.25 IU, respectively, were produced at 30<sup>o</sup>C incubation temperature.

**Conclusion:** The effect of process parameters such as effect of temperature, pH and inoculum size was also investigated. The production of primary metabolites by microorganisms is highly influenced by their growth, which is determined by the availability of the nutrients in the substrates.

**Keywords:** Cellulase, *Curvularia pallescens*, textile effluent, submerged fermentation, wheat bran

### **INTRODUCTION**

Cellulases are important industrial enzymes and find application in several industrial processes (Kang *et. al.*, 2004). Currently, the most important application is the bio-bleaching of pulp and, the production of dissolving pulp, the treatment of wastewater. The cost of production and low yields of these enzymes are the major problems for industrial application. Therefore, investigations on the ability of the cellulose hydrolyzing microbial strains to utilize inexpensive

39 substrate have been done (Kang *et al.*, 2004; Haltrich *et al.*, 1996). The enzyme is commercially  
40 used after extracting from many microorganisms especially fungal sources (Hanif *et al.*, 2004;  
41 Kang *et al.*, 2004) of mostly terrestrial origin but less from marine sources.

42 Therefore, in the present study, the enzyme production was attempted in a fungus, *Curvularia*  
43 *pallescens* isolated from textile effluent for maximizing its production under optimal conditions  
44 in submerged fermentation by using inexpensive substrate wheat bran.

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## 46 MATERIALS AND METHODS

### 47 Organism and culture condition

48 *Curvularia pallescens* was isolated from textile effluent using serial dilution and spread plate  
49 method (Graca *et al.*, 1997).

50 All the enzyme production studies were carried out under submerged conditions in a medium  
51 contain wheat bran 4.5%, yeast extract 1.5%, glucose 1%, NH<sub>4</sub>Cl 0.25%, Thiamine dichloride  
52 0.05%, KH<sub>2</sub>PO<sub>4</sub> 0.2%, MgSO<sub>4</sub>.7H<sub>2</sub>O 0.05%, CaCl<sub>2</sub> 0.01%, KCl 0.05%.. Ten (10) agar plugs of  
53 8mm diameter of the fungus grown for 7 days on PDA culture plates were inoculated in 100ml  
54 of the medium. The flasks were incubated at 28<sup>o</sup>C under shaker conditions at 120 rpm. Cultures  
55 were harvested on 5<sup>th</sup> day and assayed for cellulase activity.

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### 57 Optimization of the medium

58 Standardization of the optimum condition for the growth of the isolated organism as well as for  
59 cellulase production was determined by varying temperature, and initial pH of the medium,  
60 carbon and nitrogen sources, inoculum size, incubation period, mechanical shaking with different  
61 speed during incubation.

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### 63 Cellulase assay

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65 The test fungus was assayed for total cellulolytic activity by filter paper assay (FPA) (Mandel *et*  
66 *al.*, 1976); endoglucanase (Cx) activity by carboxymethyl assay (CMC), cellobiohydrolase (C1)  
67 activity by cotton assay and  $\beta$ -glucosidase activity by using p-nitrophenyl- $\beta$ -D-  
68 pyranoglucosidase (PNPG) method (Rosenberg *et al.*, 1975).

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70 1 unit of FPA , CMC<sub>case</sub> and cotton activity was defined as the amount of enzyme that releases 1  
71 micromole of glucose from the substrate per minute and 1 unit of  $\beta$ -glucosidase was defined as  
72 the amount of enzyme required to liberate 1 micromole of 4-nitrophenol per minute.

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77 **RESULTS**

78 Glucose favoured cellobiohydrolase and endoglucanase activity in *C. pallescens* (30.35 and  
79 21.24U/ml respectively) whereas sucrose and fructose proved to be best for FPA (61.35 U/ml)  
80 activity and  $\beta$ -glucosidase (6.97 U/ml) activity respectively (figure-1).

81 Organic nitrogen sources used for optimization were peptone, malt and yeast. *C. pallescens*  
82 showed maximum FPA activity 38.59 U/ml, cellobiohydrolase activity 30.35 U/ml,  $\beta$ -  
83 glucosidase activity 3.08 U/ml in the presence of yeast whereas endoglucanase activity 33.71  
84 U/ml reported higher with malt extract (figure-2).

85  $(\text{NH}_4)_2\text{SO}_4$  was reported as a best inorganic nitrogen source for cellobiohydrolase,  
86 endoglucanase and  $\beta$ -glucosidase activities at 57.18, 56.82 and 6.77 U/ml respectively. FPA  
87 activity was shown highest at 87.59 U/ml with  $\text{NaNO}_3$  (figure-3).

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89 The test fungus achieved maximum FPA activity followed by cellobiohydrolase, endoglucanase  
90 and  $\beta$ -glucosidase activity at 46.76, 42.06, 26.94 and 3.56 U/ml respectively at pH 5.5 (figure-  
91 4). The temperature of 28<sup>o</sup>C produced maximum cellulase activity. Highest activity recorded was  
92 of FPA (38.94 U/ml), followed by cellobiohydrolase (30.29 U/ml), endoglucanase (22.41 U/ml),  
93 and  $\beta$ -glucosidase (3.98 U/ml) (figure-5). FPA activity 38.65 respectively was obtained  
94 maximum for *C. pallescens* after 168 hrs whereas cellobiohydrolase, endoglucanase and  $\beta$ -  
95 glucosidase activities 40.29, 57.41 and 2.98 U/ml respectively were recorded highest at 120 hrs  
96 of incubation (figure-6).

97 Media containing various amounts of inocula were used for studying the effect of inoculum size  
98 on lignocellulolytic activity. Results are shown in figure-7. Reported maximum FPA,  
99 Cellobiohydrolase, endoglucanase and  $\beta$ -glucosidase activities 37.94, 30.01, 22.24 and 3.98  
100 U/ml by inoculation 10 disc of 8mm size in the production medium. *C. pallescens* also gave  
101 maximum cellulase production at 120rpm. Endoglucanase activity was observed highest  
102 followed by FPA, cellobiohydrolase and  $\beta$ -glucosidase activities as 38.59, 30.35, 27.41 and 1.91  
103 U/ml respectively (figure-8).

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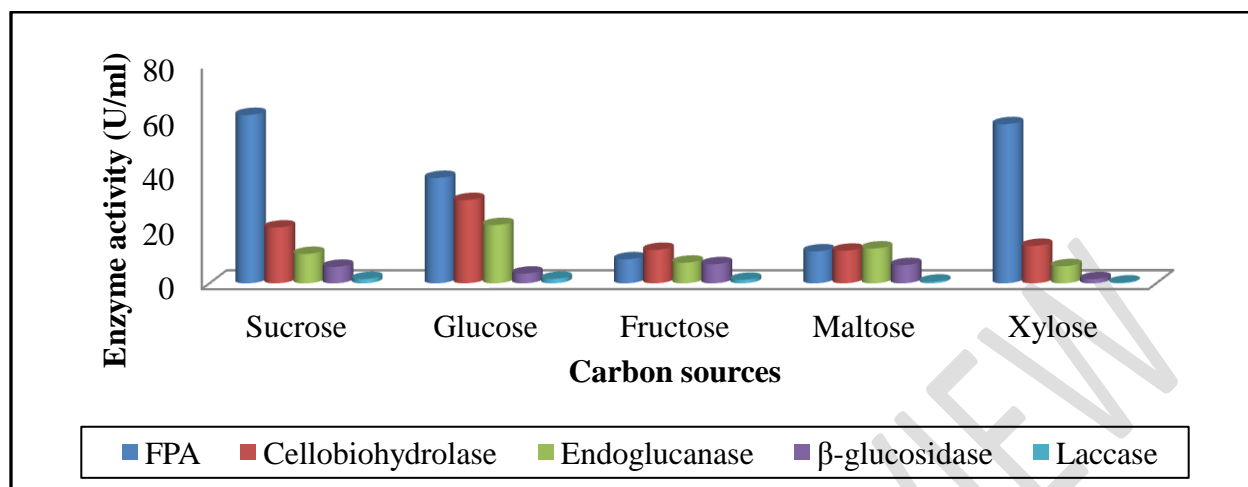
## 105 DISCUSSION

106 **Medium optimization is an important** aspect to be considered in the development of fermentation  
107 technology. The production of primary metabolites by microorganisms is highly influenced by  
108 their growth, which is determined by the availability of the nutrients in the substrates. Garcia *et*  
109 *al.*, (2002) reported that submerged fermentation for aerobic microorganisms is a well known  
110 and widely used method for the production of cellulase and xylanase. Chellapandi and Jani  
111 (2009) reported enhanced endoglucanase production by soil isolates of *Fusarium* sp. and  
112 *Aspergillus* sp. through the submerged fermentation process. Papinutti and Lechner (2008)  
113 studied the influence of the carbon source on the growth and lignocellulolytic enzyme production  
114 by *Morchella esculenta*. Arora and Sehgal (2010) reported the production of cellulase and  
115 xylanase by *Scopulariopsis acremonium* through submerged fermentation using shake flask  
116 cultivation media. The effect of process parameters such as the effect of temperature, pH and  
117 inoculum size was investigated. Maximum cellulase and xylanase having an enzyme activity of  
118 694.45 and 931.25 IU, respectively, were produced at 30°C incubation temperature. The pH  
119 optimum to achieve these enzyme activities was 5.5 with an inoculum size of  $1 \times 10^5$  spores ml<sup>-1</sup>  
120 of tween – 80.

121 Gupta *et. al.*, (1990) studied microbial proteins and cellulase production from cellulosic  
122 materials by *Coprinus atramentarius* and reported the optimum pH for protein production and  
123 extracellular enzymes (cellulase and xylanase) by *C. atramentarius*, utilizing cellulose to be 6  
124 and optimum temperature 30°C. The resulting enzyme activities were endoxylanase as 7.2 IU ml<sup>-1</sup>  
125 <sup>1</sup>, exoglucanase as 1.0 IU ml<sup>-1</sup> and xylanase as 5 IU ml<sup>-1</sup>. Li *et al.*, (2006) reported pH of 4.14  
126 was reported to be optimum for the production of endoxylanase production by *Aspergillus*  
127 *awamori* under submerged fermentation which gave an enzyme activity of 28.25 U ml<sup>-1</sup>.

128 Shear stress within the medium, which is directly related to the stirrer speed, has also been  
129 shown to have a marked influence on enzyme production by *Thermomyces lanuginosus* SSBP  
130 (Reddy *et. al.*, 2002; Singh *et. al.*, 2000). Acharya *et. al.*, (2008) reported maximum cellulase  
131 production by *Aspergillus niger* in submerged fermentation at 120 rpm. However, Ojumu *et. al.*,  
132 (2003) observed maximum cellulase production by *Aspergillus flavus* Linn isolate NSPR 101 at  
133 the agitation of rate 200 rpm.

134 **Figure-1: Optimization of carbon source for lignocellulases production by *Curvularia pallescens***



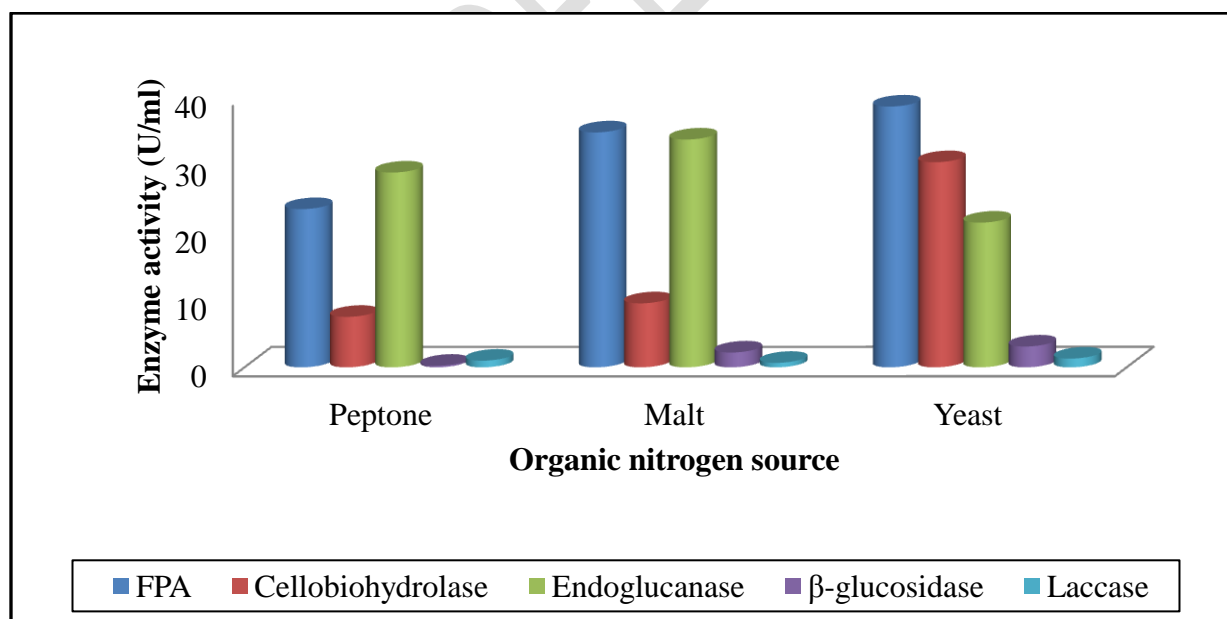
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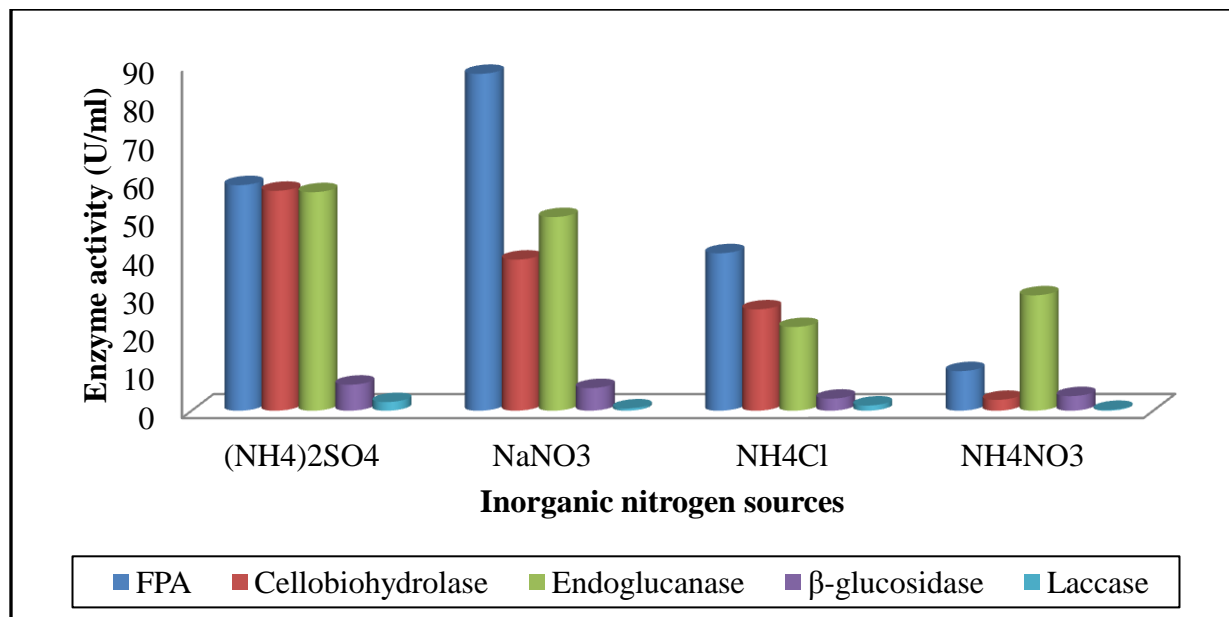
139 **Figure-2: Optimization of nitrogen source (organic) for lignocellulases production by *Curvularia***  
140 ***pallescens***



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143 **Figure-3: Optimization of nitrogen source (inorganic) for lignocellulases production by *Curvularia***  
144 ***pallescens***



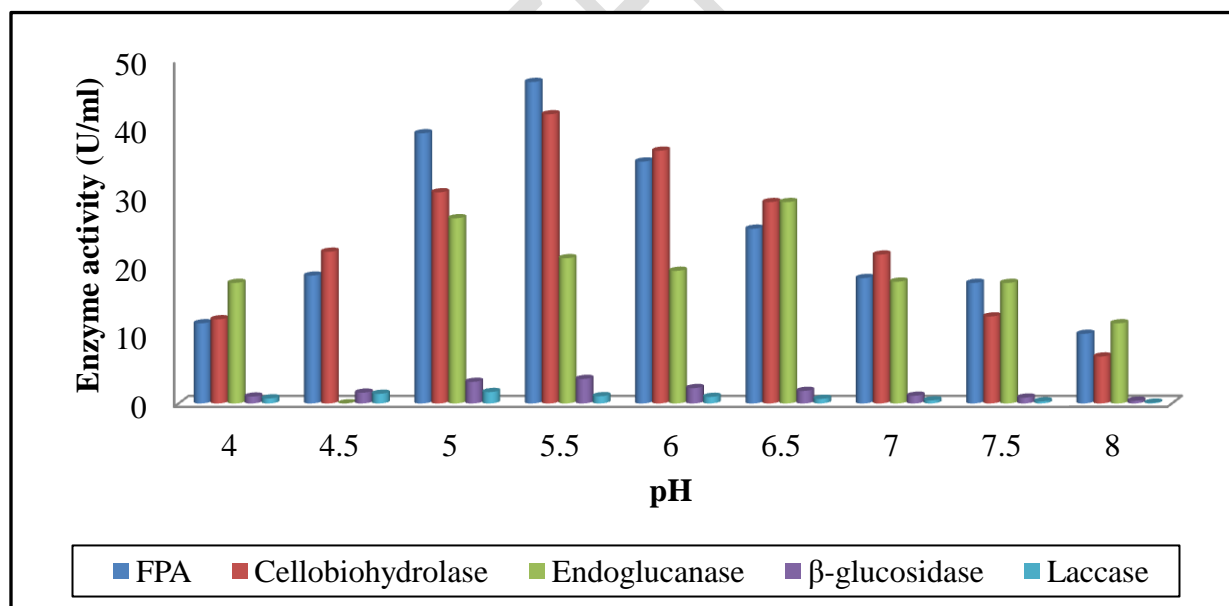
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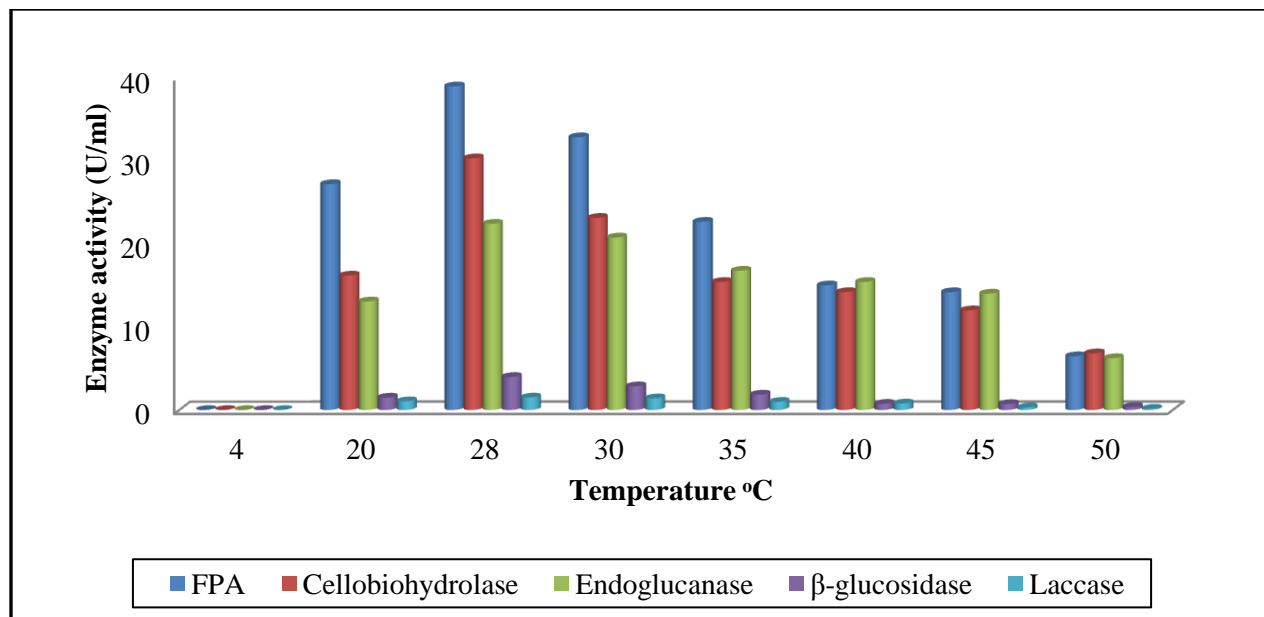
Figure-4: Optimization of pH for lignocellulases production by *Curvularia pallescens*



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Figure-5: Optimization of temperature for lignocellulases production by *Curvularia pallescens*



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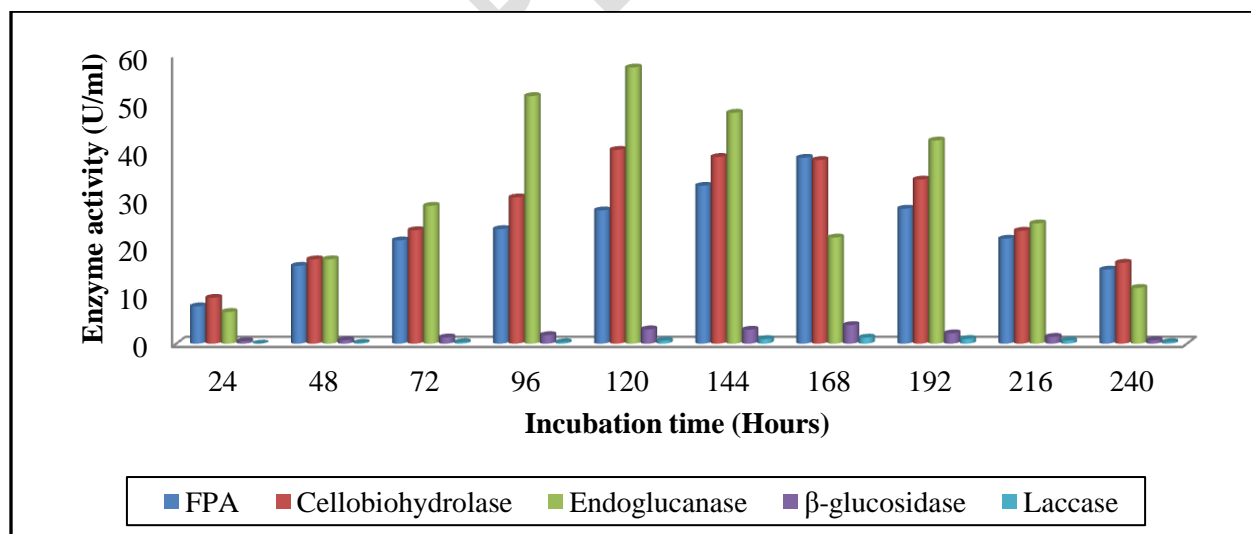
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Figure-6: Optimization of incubation time for lignocellulases production by *Curvularia pallescens*

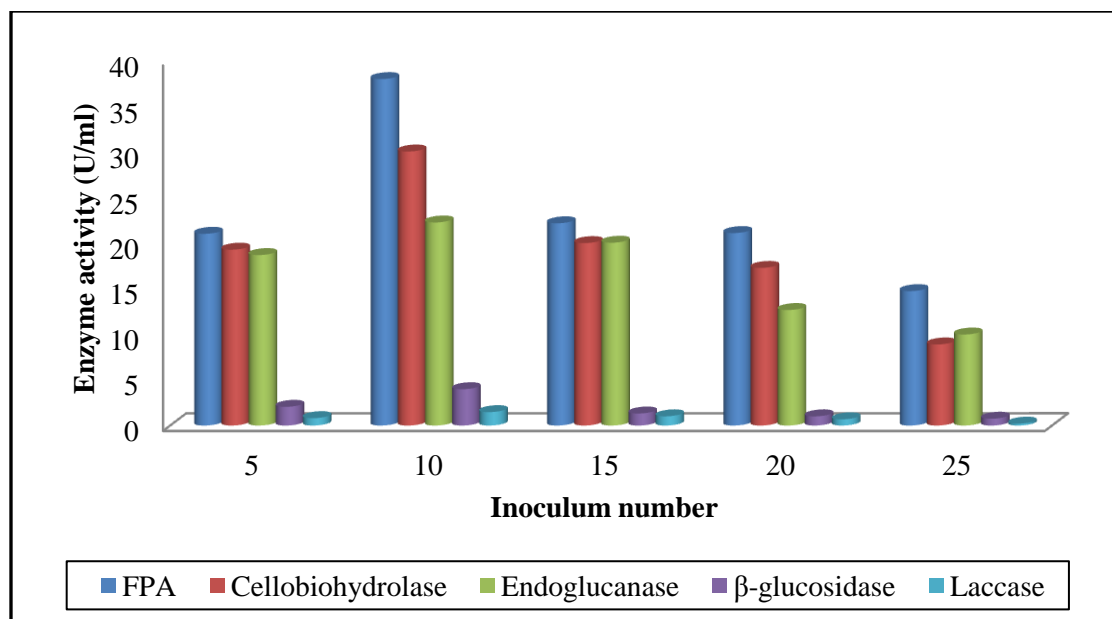


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Figure-7: Optimization of inoculum size for lignocellulases production by *Curvularia pallescens*



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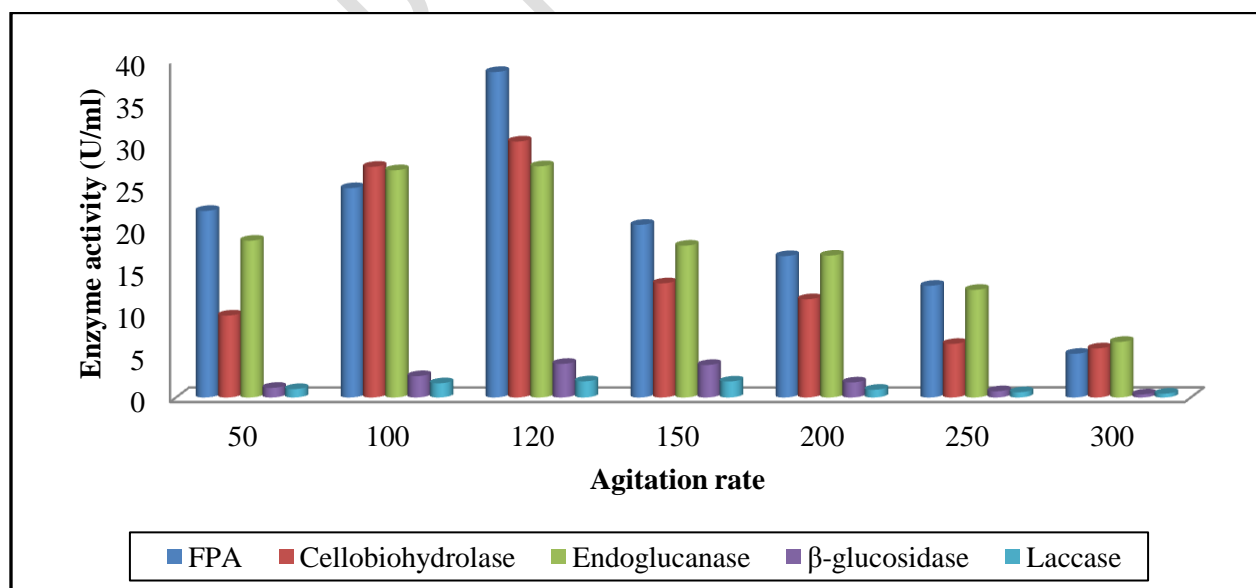
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Figure-8: Optimization of agitation rate for lignocellulases production by *Curvularia pallescens*



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169 **Conclusion**

170 Effects of pH, temperature, incubation time, source of carbon and nitrogen were investigated in  
171 the submerged fermentation process in the production of cellulose by *Curvularia pallescens*  
172 isolated from textile effluent. The effect of process parameters such as the effect of temperature,  
173 pH and inoculum size was also investigated. The production of primary metabolites by  
174 microorganisms is highly influenced by their growth, which is determined by the availability of  
175 the nutrients in the substrates.

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