

1 **THE IMMUNO-MODULATORY AND THROMBOCYTOPAENIC EFFECTS OF THE**  
2 **VARYING CONCENTRATIONS OF THE AQUEOUS LEAF EXTRACT OF MORINGA**  
3 **OLEIFERA IN MALE ALBINO WISTAR RATS.**

4  
5 **Abstract**

6 **Introduction:** *Moringa oleifera* and related species are commonly used in folk medicine for  
7 various human diseases.

8 **Aim:** The study was undertaken to establish the thrombocytopenic effect of the aqueous leaf  
9 extract of *moringa oleifera* and to find the utilization of platelet parameters in determining the  
10 cause of the thrombocytopenia.

11 **Methodology:** Fresh leaves of moringa were dried and extracted with water. Thirty (30) male  
12 albino Wistar rats, weighing between 150-250g, which were kept under uniform laboratory  
13 conditions, were randomly divided into five (5) groups (A-E), based on their weights. The  
14 control group (group A) was orally given 0.5ml of distilled water while the treatment groups  
15 (groups B to E) were orally given 250mg/kg, 450mg/kg, 650mg/kg and 850mg/kg body weight  
16 respectively of the extract, once a day, for 56 days and then sacrificed. At the end of the  
17 administration, blood samples were collected from each rat and examined for platelet indices.  
18 The effects of treatment with aqueous leaf extract of *moringa oleifera* on the platelet parameters  
19 were compared with the control group.

20 **Result:** The rats treated with the extract, showed a decrease in platelet count and platelet crit  
21 while there was a significant increase in the platelet distribution width, mean platelet volume and  
22 immature platelet fraction, concerning the control.

23 **Conclusion:** The aqueous leaf extract of *moringa oleifera* is therefore shown to modulate the  
24 immune system and cause thrombocytopenia, through platelet destruction.

25  
26 **Keywords: Moringa, Platelet, Thrombocytopenia, Blood, Immuno-modulatory.**

27  
28 **INTRODUCTION**

29 Thrombocytopenia (TCP) refers to a disorder in which there is a relative decrease of platelets,  
30 present in the blood<sup>[5]</sup>. A normal human platelet count ranges from 150,000 to 450,000 platelets  
31 per microliter of blood and thrombocytopenia is said to be a platelet count below 50,000 per  
32 microliter<sup>[8]</sup>. The causes of thrombocytopenia include decreased platelet production, increased  
33 platelet destruction and splenic sequestration/abnormal pooling, based upon the causative  
34 process<sup>[7, 8, 11]</sup>. It is one of the common causes of abnormal bleeding and characterized by  
35 spontaneous bleeding from the skin, arms, nose, gums and other mucous membranes<sup>[8,11]</sup>. In  
36 many cases of thrombocytopenia, large platelet was seen in the peripheral smear, this size and  
37 other platelet parameters were suggested to help in deciding the category of thrombocytopenia  
38<sup>[15]</sup>. The present study was undertaken to establish the thrombocytopenic effect of the aqueous

39 extract of moringa oleifera leaf and to find the utility of platelet parameters in determining the  
40 cause of the thrombocytopenia.

#### 41 **MATERIALS AND METHOD**

42 Thirty male albino Wistar rats weighing between 120g-250g were used for the experimental  
43 work. The animals were obtained from the animal care facility at the University of Portharcourt.  
44 The animals were housed in a wooden cage made of five (5) different compartments and the rats  
45 were placed in the cage and grouped into five (5) groups. The animals were allowed to  
46 acclimatize for fifteen days, to observe for any signs of illness before the experiment started.  
47 They were kept under standard laboratory conditions in a well ventilated standard housing  
48 condition and clean wooden rat cage with proper bedding (sawdust). The animals were properly  
49 fed with tap water and standard rat feed that contains groundnut, wheat brand, maize grains,  
50 palm kernel and fish meal, bought from the animal feed store in Choba. The feeding and water  
51 troughs were thoroughly cleansed daily to ensure proper hygiene and healthy living condition.  
52 The animal bedding was prepared with sawdust particles, obtained from a sawmill. These  
53 beddings were changed regularly to ensure a healthy environment for the animals.

54 The rats were randomly grouped into five (5) groups (groups A-E), comprising six rats in each  
55 group. A calculated amount of the aqueous extract of moringa was constituted in 20mls of  
56 distilled water to give doses of 250mg/kg to 850mg/kg body weight. Administration of the  
57 aqueous leaf extract of moringa oleifera was performed orally once daily, between 7.30 am and  
58 9.30 am, using a 2ml syringe. The various groups were administered as follows:

59 A) Group A served as the control, with no extract being administered; instead, 2ml of distilled  
60 water was given

61 B) Groups B, C, D and E received 2ml of the moringa extract, using a syringe from a 250mg/kg  
62 for group B, 450mg/kg for group C, 650mg/kg for group D and 850mg/kg for group E.

63 These administrations were carried out in the space of 56 days after which the animals were  
64 sacrificed and the blood samples collected in an EDTA bottle. The blood samples were collected,  
65 using the method of cardiac puncture, after each rat has been anaesthetized in a desiccator, using  
66 diethyl ether.

67 The *Moringa* leaves were shaded, dried at a warm temperature (not directly under the sun),  
 68 before tasking it for the preparation of the extract. The leaves were separately rinsed in clean  
 69 water to remove dirt, dried at room temperature for 14 days. 500g of the plant material was  
 70 introduced into an extraction jar. 1.2 litres of sterile distilled water was added into it and corked,  
 71 kept at room temperature and shaken at an interval of 30 minutes (with a mechanical shaker). It  
 72 was filtered after 24 hours, the discarded material and the filtrate concentrated using the rotary  
 73 evaporator in a vacuum. The paste collected and air-dried and weighed. The percentage yield  
 74 was calculated and the extract stored at -4°C in the refrigerator for photochemical studies on the  
 75 animals. Chemical tests were carried out on the aqueous extract and on the powdered specimens  
 76 using standard procedures, to identify the constituents as described by Sofowara (1993), Trease  
 77 and Evans (1919) and Harborne (1973).

## 78 STATISTICAL ANALYSES

79 The results were subjected to statistical analysis using statistical package for social sciences  
 80 (SPSS) version 20.0. Data are presented as mean  $\pm$  SEM. The difference of means was  
 81 considered significant at P value less than 0.05.

## 82 RESULTS

83 The results of the qualitative phytochemical analysis indicate that alkaloids were most  
 84 abundantly present while tannins, saponins, Salkowski, free anthraquinone, flavonoids were  
 85 moderately present. Steroids, Phlobatanins, combined anthraquinone, Lieberman's and Keller  
 86 kiliani were only slightly present while Cyanogenetic glycosides were observed to be absent  
 87 (Table 1).

88 **Table 1: Qualitative phytochemical analysis of aqueous leaf extract of *Moringa oleifera* leaf**  
 89 **in Wistar rats.**

Phytochemical	Observation	Inference
Alkaloid	+++	Heavily present
Tannins	++	Moderately present
Saponins	++	Moderately present
Flavonoids	++	Moderately present
Steroids	+	Slightly present

Phlobatanins	+	Slightly present
Combined anthraquinone	+	Slightly present
Free anthraquinone	++	Moderately present
Cyanogenetic glycosides	-ve	Absent
Salkowski	++	Moderately present
Liebermanns	++	Slightly present
Keller kiliani	+	Slightly present

91 +, slightly present; ++, moderately present; +++, heavily present; -ve, absent; \*, Significant  
 92 at P < 0.05 when compared to control.

93  
 94 The results of the quantitative phytochemical analysis indicate the presence of Polyphenols,  
 95 flavonoids, tannins, alkaloids and glycosides in the following percentages: 2.70, 4.10, 8.00,  
 96 15.00, and 2.50%, respectively (Table 2). The effects of the graded doses of moringa oleifera on  
 97 platelet indices were also determined (Table 3)

98 **Table 2: Quantitative phytochemical analysis of aqueous leaf extract of *Moringa oleifera***  
 99 **leaf in Wistar rats.**

Phytochemical	Percentage abundance (%)
Polyphenols	2.70
Flavonoids	4.10
Tannins	8.00
Alkaloids	15.00
Glycoside	2.50
Polyphenols	2.70

\*, Significant at P < 0.05 when compared to control.

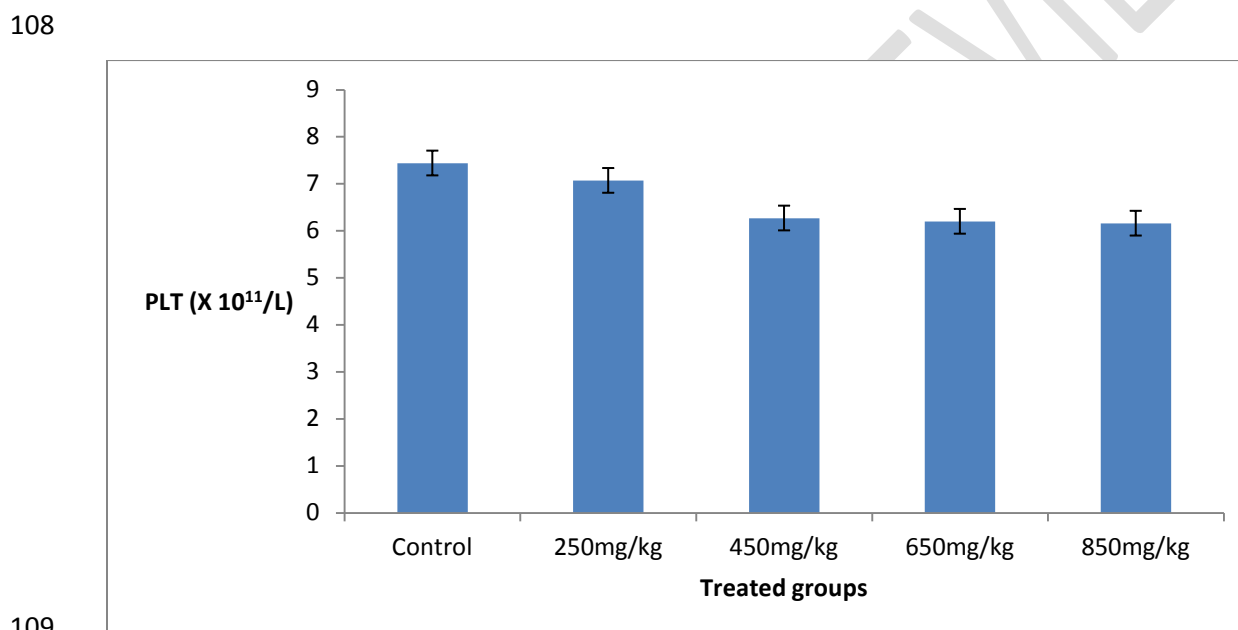
100 **Table 3: Effects of the graded doses of *Moringa oleifera* on platelet indices**

Groups	PLT(x10 <sup>11</sup> /l± sem)	PCT (ml/L ± sem)	MPV (fl ± sem)	PDW (%± sem)	IPF (%± sem)
Control	7.44 ± 0.33	5.20 ± 0.15	7.02 ± 0.22	15.08 ± 0.09	1.48 ± 0.36
250mg/kg	7.07 ± 0.56	5.20 ± 0.44	7.25 ± 0.10	15.17 ± 0.08	1.70 ± 0.21

<b>450mg/kg</b>	6.27 ± 0.31	4.77 ± 0.22	7.50 ± 0.10*	15.22 ± 0.07	2.08 ± 0.18
<b>650mg/kg</b>	6.20 ± 0.50	4.80 ± 0.33	7.37 ± 0.12	15.13 ± 0.06	2.35 ± 0.21*
<b>850mg/kg</b>	6.16 ± 0.42*	5.05 ± 0.28	7.83 ± 0.02*	15.12 ± 0.04	2.28 ± 0.15

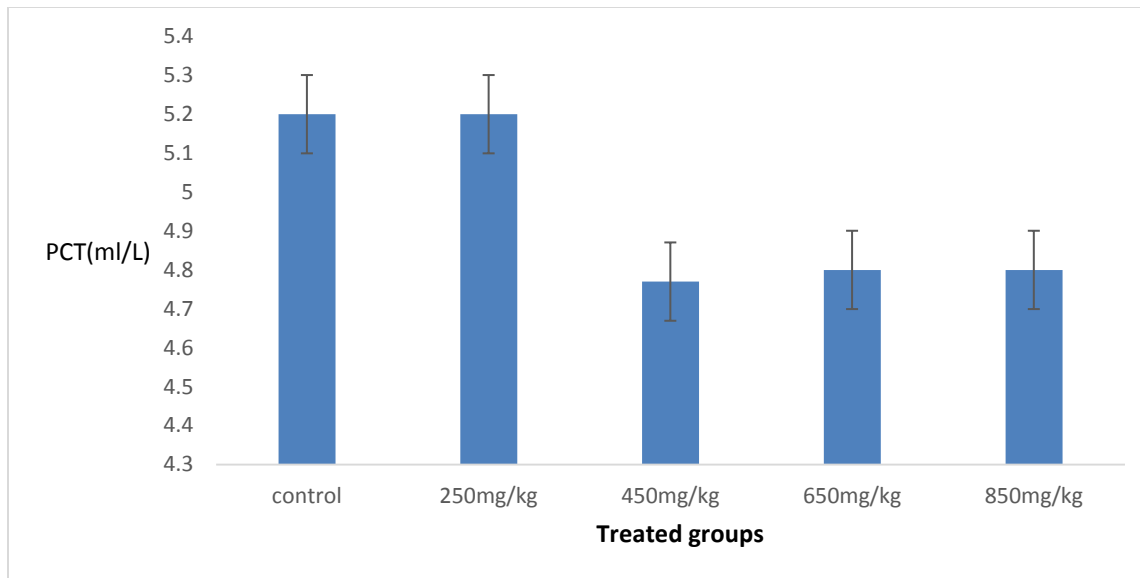
101 **All values are expressed as Mean ± S.E.M. n = 6, p < 0.05. \*=statistically significant when**  
 102 **compared to the control**

103  
 104 The result of *M. oleifera* extract on platelet indices shows a significant ( $P < 0.05$ ) decrease in the  
 105 level of platelet count for the high dose administered when compared with the control. A non-  
 106 significant reduction in the level of platelet count was seen for the 250, and 450 and 650 mg/kg  
 107 doses administered when compared with the control (Fig. 1).



109  
 110 Fig.1: Effect of *Moringa oleifera* leaf extract on platelet count (PLT)

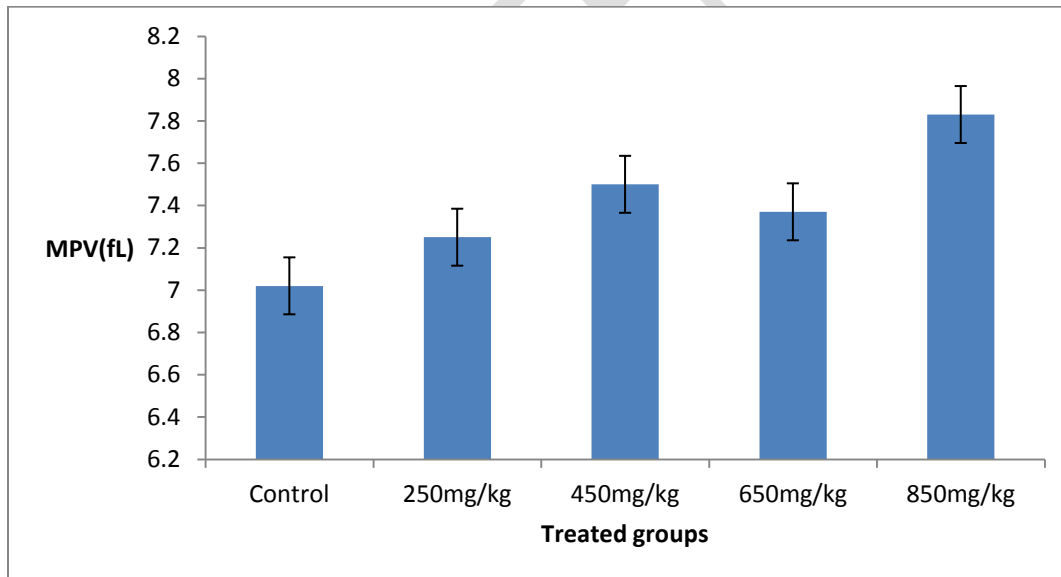
111  
 112 A dose-dependent non-significant ( $P > 0.05$ ) decrease in the level of Plateletocrit (PCT) was  
 113 observed following the administered doses when compared with the control. Following the  
 114 administration of the extract, a non-significant decrease was recorded in the level of PCT for the  
 115 doses of 450, 650 and 850 mg/kg when compared to the control (Fig 2). A significant ( $P < 0.05$ )  
 116 increase in the level of mean platelet volume (MPV) for the 450 mg/kg and 850mg/kg  
 117 administered dose when compared to the control. However, a dose-dependent but non-significant  
 118 increase in the level of mean platelet volume (MPV) was observed for the 450mg/kg and  
 119 850mg/kg doses administered (Fig 3).



120

121 Fig.2: Effect of *Moringa oleifera* leaf extract on platelet crit (PCT)

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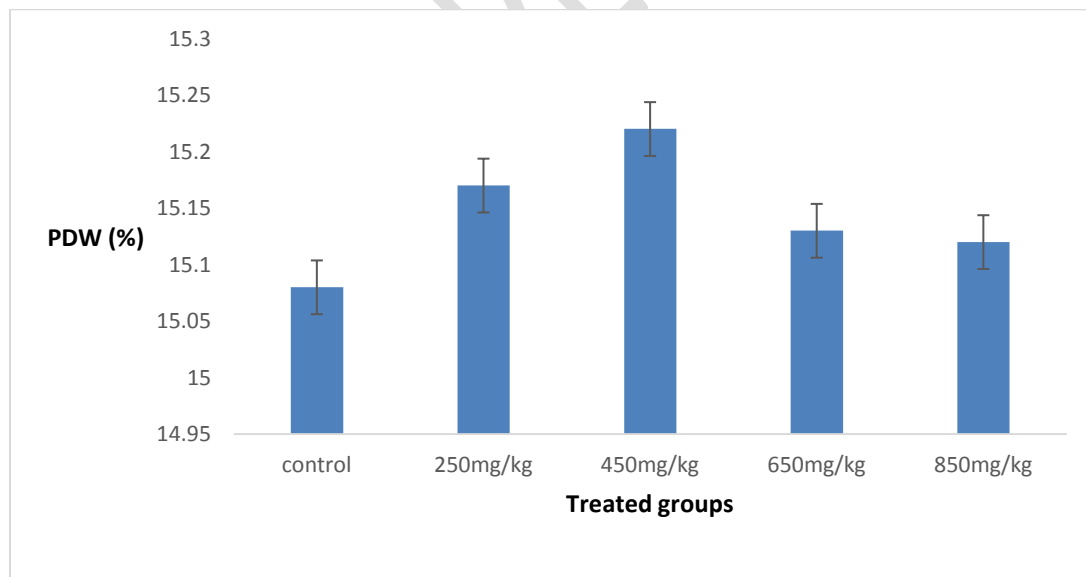
124 Fig.3: Effect of *Moringa oleifera* leaf extract on the mean platelet volume (MPV)

125 A dose-dependent non-significant ( $P > 0.05$ ) increase in the level of Platelet  
126 distribution width (PDW) was observed following the administered doses when  
127 compared with the control (Fig 4). A significant ( $P < 0.05$ ) increase in the level of  
128 immature platelet fraction (IPF) for the 650 mg/kg administered dose when  
129 compared to the control. However, a dose-dependent but non-significant increase  
130 in the level of immature platelet fraction (IPF) was observed for the 250mg/kg,  
131 450mg/kg and 850mg/kg doses administered (Fig 5).

132

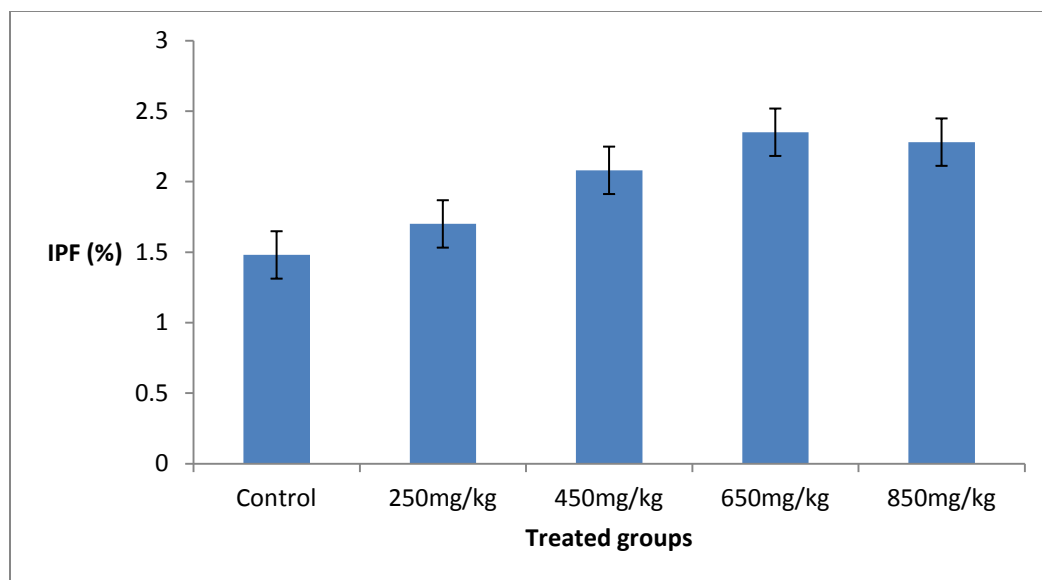
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136 Fig.4: Effect of *Moringa oleifera* leaf extract on platelet distribution width (PDW)



137  
 138 Fig.5: Effect of *Moringa oleifera* leaf extract on immature platelet fraction (IPF)

139 **DISCUSSION**

140 Table 1 above, reveals that the extract contains various types of phytochemicals in different  
 141 concentrations. Table 2 shows that Alkaloids are the most abundant phytochemicals (have the  
 142 highest percentage abundance) while Glycosides are the least abundant (in terms of percentage  
 143 abundance). The level of alkaloids in the aqueous leaf extract of *M. oleifera* (tables 1 and 2), may  
 144 suggest that the extract has immunomodulatory activity, since some bitter alkaloids (tropane  
 145 alkaloids) are metabolized in the liver into dimethylxanthine and finally methyl uric acid by  
 146 cytochrome p<sub>450</sub> oxygenase systems. Methyl uric acid in the liver stimulates the expression of  
 147 tumour necrosis factor (in the endothelial cells of the liver by macrophages) which modulates the  
 148 immune system<sup>[16]</sup>. Also, saponins, which was moderately present, are implicated in the  
 149 modulation of the immune system by serving as an adjuvant ( saponins – cholesterol –  
 150 phospholipid complexes) at low concentrations that stimulate the cell-mediated immune system  
 151 by inducing the production of interleukins, especially by the antigen-presenting cells in most  
 152 cells<sup>[13,17]</sup>. The presence of phenolic compounds in the extract may help among others, in  
 153 preventing oxidative stress by scavenging free radicals and bioactivation of carcinogens for  
 154 excretion in the liver

155 Phenolic compounds are also known to scavenge directly nitric oxide molecule, thereby  
 156 preventing the oxidation of LDL-C and tissue oxidative damage<sup>[14]</sup>. Nitric oxide is constitutively



157 produced in endothelial cells to maintain the dilation of blood vessels and relaxation of smooth  
158 muscles <sup>[9]</sup>. Flavonoids were reported to decrease also, the immobilization and adhesion of  
159 leukocytes to endothelial walls, and degranulation of neutrophils without affecting superoxide  
160 production, thereby regulating inflammatory responses in tissue injury and immune responses <sup>[6]</sup>.

161 The result of the effect of the extract on platelet count (PLT) can be seen in figure 1. The figure  
162 shows a significant dose-dependent decrease in platelet count, concerning the control. There was  
163 a statistically significant decrease in platelet count at the 850mg/kg dose of the extract, for the  
164 control. This result corroborates with other works which report a decrease in platelet count with  
165 the leaf extract of moringa oleifera<sup>[11]</sup>. This decrease in platelet parameters suggests that the  
166 extract contains some phytoconstituents which might have can destroyed the blood platelets  
167 (thrombocytopenia) and consequently causing excessive bleeding from wounds and negatively  
168 interfere with the normal coagulation process. A normal human platelet count ranges from  
169 150,000 to 450,000 platelets per microliter of blood and thrombocytopenia is said to be a platelet  
170 count below 50,000 per microliter

171 Figure 2 shows the effect of the varying concentrations of the extract on Plateletocrit. It shows  
172 that the 250mg/kg concentration of the extract has no effect on the value of the parameter, for the  
173 control but there was an observable sharp decrease in the parameter at higher concentrations of  
174 the extract. Plateletocrit is a measure of the total platelet mass. Its value depends on the mean  
175 platelet volume resulting in the overlap between normal platelets, thrombocytopenia and  
176 thrombocytosis <sup>[15]</sup>.

177 The effect of the varying concentrations of the extract on the mean platelet volume (MPV) can  
178 be seen in figure 3. There was an observable (though irregular) increase in the value of the mean  
179 platelet volume, for the control. There was a statistically significant increase ( $P < 0.05$ ) in the  
180 value of the parameter at 450mg/kg and 850mg/kg concentrations of the extract. The mean  
181 platelet volume is a machine-calculated measurement of the average size of platelets found in the  
182 blood and is typically included in the blood tests as part of the complete blood count (CBC). The  
183 normal range in Humans is given as 7.5fL-11.5fL. It reflects the average size of platelets in a  
184 person's sample of blood. Larger platelets are usually relatively young and more recently  
185 released from the bone marrow, while smaller platelets may be older and have been in the  
186 circulation for a few days. The mean platelet volume test results can be used to make inferences  
187 about platelet destruction problems; it is generally higher when there is the destruction of

188 platelets, as seen in inflammatory bowel disease<sup>[12]</sup>. Unusual low MPV values have been found  
189 to correlate primarily with thrombocytopenia when it is due to impaired production as in aplastic  
190 anaemia<sup>[12]</sup>. A large number of large platelets (a large MPV) in a person with a low platelet  
191 count, therefore, suggests that the bone marrow is producing platelets and releasing them into the  
192 circulation rapidly.

193 Figure 4 showed that the varying concentrations of the extract cause a dose-dependent increase  
194 in the Platelet distribution width (PDW) of the experimental animals. The platelet distribution  
195 width measures the heterogeneity of platelet volume; it reflects how uniform the platelets are in  
196 size. The heterogeneity of platelet volume is considered to be due to ageing of platelets or due to  
197 the heterogeneous demarcation of megakaryocytes. It has been found that the increase in platelet  
198 distribution width (increased platelet heterogeneity) is associated with thrombocytopenia caused  
199 by platelet destruction<sup>[2]</sup>.

200 The effect of the varying concentrations of the extract on the immature platelet fraction (IPF) is  
201 shown in figure 5. There was an observable dose-dependent increase in the immature platelet  
202 fraction, for the control. The immature platelet fraction (IPF, %) is a measure of reticulated  
203 platelets (RPs), which represents the state of thrombopoiesis<sup>[10]</sup>. It is obtained from an automated  
204 haematology analyzer as one of the platelet parameters. It is an index of thrombopoiesis and can  
205 help to determine the mechanism of thrombocytopenia. An increased IPF in the presence of  
206 thrombocytopenia is indicative of platelet destruction or consumption, while a decreased or low  
207 IPF value is indicative of a decreased bone marrow production of platelets<sup>[3]</sup>. Patients with  
208 decreased platelet production, including those undergoing cytotoxic chemotherapy, have been  
209 found to have IPF either in the low or normal range<sup>[3]</sup>. There has also been found a significant  
210 inverse correlation of platelet count with IPF, such that the lower the platelet count, the higher  
211 the IPF<sup>[3]</sup>.

## 212 **CONCLUSION**

213 Based on this study, it is therefore concluded that the aqueous extract of moringa oleifera leaf is  
214 immuno-modulatory and thrombocytopenic in action. Since platelets in addition to other  
215 functions have been implicated in boosting the immune system, the extract likely enhances the  
216 immune system through other mechanisms and not through an increase in platelet production.  
217 Again, the thrombocytopenic action of the extract is probably mediated through the actual  
218 destruction of the platelets and not an interruption of the platelet production at the bone marrow.

219 This is because the increase in platelet distribution width (PDW), Mean platelet volume (MPV)  
220 and Immature platelet fraction (IPF) observable from the study have been associated with actual  
221 platelet destruction.

## 222 **COMPETING INTEREST / CONFLICT OF INTEREST:**

223 The authors have declared that no competing interests exist.

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