

Original Research Article

THE EFFECT OF AIR DRIED LEAF POWDER AND BURNT LEAF ASH OF DIFFERENT PLANTS ON ROOT-KNOT NEMATODE (*Meloidogyne* spp.) ON OKRA (*Abelmoschus esculentus* L. Moench)

ABSTRACT

The effect of air dried leaf powder and burnt leaf ash of different plants on the control of root-knot nematode (*Meloidogyne* spp.) on Okra was investigated in a pot experiment. The experiment was laid out in a Completely Randomized Design with nine treatments each replicated four times. The treatments included: burnt leaf ashes and air dried leaf powders of *Baphia nitida*, *Pentaclethra macrophylla* and *Delonix regia* respectively, a synthetic nematicide (Carbofuran 3G) and two control which included inoculated but untreated (control 1) and uninoculated and untreated (control 2) pots. The plants were inoculated with 1000 nematode eggs (*Meloidogyne* spp.) two weeks after germination. Three days after, the treatments were applied at the rate of 30 g each while the synthetic nematicide (Carbofuran 3G) at 3 g/per pot. Parameters recorded were: plant height, number of leaves, fresh and dry shoot weights, fresh root weight, and number and weight of pods per plant as plant growth parameters. Number of galls, number of nematode eggs in roots, and number of juveniles in soil as nematode parameters were considered. Results obtained indicated significant differences ($P \leq 0.05$) among the treatments in most parameters recorded and compared to controls. Generally, results from air-dried leaf powder of *Baphia nitida* and burnt leaf ash of *Delonix regia* were better and compared favourably with the nematicide treated plants.

Key words: Air dried leaf, Burnt leaf ash, Carbofuran, *Meloidogyne* spp., *Abelmoschus esculentus*

INTRODUCTION

Okra as a vegetable crop belongs to the genus *Abelmoschus*, Family *Malvaceae* and has two main species: *Abelmoschus esculentus* (L) Moench and *Abelmoschus caillei* (A. Chev) (Siemonsma, 1982). It is an annual crop mainly grown as a fruit and leafy vegetable in the tropics (Tamiyu *et al.*, 2012). It is widely distributed in the Tropical and Subtropical regions of Asia, Southern Europe, the Mediterranean countries and the Americas (Andres *et al.*, 2005). Nigeria is the second largest producer of Okra in the world after India and the largest producer in Africa (5.8 million tonnes), followed by Cote d'Ivoire, Ghana and others (FAOSTAT, 2011). In Nigeria, it is widely distributed and consumed in either fresh or dried forms (Farinde *et al.*, 2007).

Okra is an important constituent of most local dishes in West Africa. It is used as a soup thickener and may also be served with rice and other food types (Tamiyu *et al.*, 2012). In Nigeria, okra is grown across different ecological zones because it serves as a source of income to framers as well as a cheap source of protein, vitamins A, B complex and C and mineral (Ca, P, Fe and K) to many households (Adebisi *et al.*, 2007). It is a nutritious vegetable that contains 86.1% water, 2.2% protein, 0.2% fat, 9.7% carbohydrates, 1.0% fiber and 0.8% ash (Saifullah and Rabbani, 2009), hence it plays vital role in the human diet. Okra seeds contain greenish yellow edible oil which is also suitable for use as a bio-fuel (Farroq, *et al.*, 2010).

43 Okra is notorious for its susceptibility to root-knot nematodes (Noling, 2012). *Meloidogyne* spp.
44 are responsible for about 70-90% of the yield losses in okra (Saufiuddin *et al.*, 2011). Almost all
45 the vegetables in tropical and warm temperate regions are severely attacked by plant-parasitic
46 nematodes particularly root-knot nematodes (*Meloidogyne* spp.) (Sikora and Fernandez, 2005).
47 *Meloidogyne* spp. cause wilting, chlorosis, stunted growth, the formation of galls in roots often
48 leading to destruction of roots, poor growth, yield and crop failure when nematode population
49 exceeds economic threshold level (Fourie *et al.*, 2001; Sikora and Fernandez, 2005). The
50 symptoms for identifying disease caused by *Meloidogyne* spp. is the presence of galls, wilting,
51 loss of vigor, yellowing of leaves (Kayani *et al.*, 2012). They have caused up to 80% yield losses
52 in heavily infested soils (Kaskavalci, 2007). However, the extent of damage is influenced by the
53 cultivar, nematode species, level of soil infestation and environment (Ononuju, 1999). It is,
54 therefore, necessary to control root-knot nematodes in order to avoid or minimize yield losses in
55 okra.

56 Management of root-knot nematodes with synthetic nematodes can be very effective (Sikora and
57 Fernandez, 2005; Adegbite and Agbaje, 2007). However, there is public outcry against synthetic
58 nematicides due to their toxicity, persistence and hazards posed to non-target species and
59 agriculturists (Oka *et al.*, 2014). As a result, there is a growing preference for plant products
60 which are less harmful, effective, easily degraded, pollution free, leave no harmful residues,
61 cheaper and not toxic to host plants and humans (Amadioha, 2003). These plants if explored for
62 nematicidal activity might be alternatives to synthetic nematicides (Siji *et al.*, 2010).

63 The objective of this study is to determine the effect of air-dried leaf powder and burnt leaf ash
64 of *Baphia nitida*, *Pentaclethra macrophylla* and *Delonix regia* on the population of root-knot
65 nematode (*Meloidogyne* spp.) on the growth and yield of okra, gall formation and also to
66 compare the effect of these plant materials with the synthetic nematicide (Carbofuran 3G).

67 MATERIALS AND METHODS

68 LOCATION OF EXPERIMENTAL SITE

69 The experiment was carried out in the Department of Plant Health Management, Michael Okpara
70 University of Agriculture, Umudike, Abia State, Nigeria located on latitude 5⁰2¹N and longitude
71 7⁰33¹E. It lies in the humid tropical rain forest zone with annual rainfall of 1916 mm per annum,
72 the altitude of 112m above sea level and relative humidity of 76% with temperature range of 19-
73 35⁰C (N.R.C.R.I, 2010).

74 EXPERIMENTAL DESIGN

75 The experiment was laid out in a Completely Randomized Design (CRD) on a platform in an
76 open field using plastic pots. Okra variety *Pusa suwaan* was planted. There were nine treatments
77 replicated four times including the control, giving a total of 36 pots.

78 EXPERIMENTAL MATERIALS

79 **Source of seed:** Seeds of okra variety *Pusa suwaan* were obtained from National Root Crop
80 Research Institute Umudike, Abia State, Nigeria.

81 **Treatments:** The materials used were: burnt leaf ash and air dried leaf powder of *Baphia nitida*,
82 *Pentaclethra macrophylla* and *Delonix regia* and a synthetic nematicide (Carbofuran 3G).

83 **Treatments' sources and preparation:** The plant materials (leaves and/or roots) were obtained
 84 within the school premises while the synthetic nematicide (Carbofuran 3G) was obtained from a
 85 chemical store. The plant materials were collected and washed under a running tap to remove
 86 soil particles and reduce contamination by microorganisms. They were then spread on a clean
 87 platform for air drying. This was done to reduce the moisture content after which each material
 88 was divided into two parts: one part was ground into powder using an electric grinder while the
 89 other part was burnt to ash in a controlled environment and other parts were put in a clean plastic
 90 container ready for application.

91 **Soil preparation and sterilization:** Top soil was collected and sifted to remove large soil
 92 particles, stones and plant debris. The soil was poured into a cut metal drum. It was moistened as
 93 a source of heat applied underneath. It was sterilized at 80°C for 20 minutes after which it was
 94 left to cool (Ononju *et al.*, 2014).

95 **Extraction of nematode eggs from inoculum:** Eggs of root-knot nematodes were extracted
 96 from the heavily galled roots of *Basella alba* (Ceylon spinach). The galled roots were washed in
 97 tap water and cut into pieces of 2cm long and placed in a beaker containing 200 mls of 0.5%
 98 NaOCl (Sodium hypochloride) solution and shaken vigorously for about 5minutes (Hussey and
 99 Barker, 1973). This was done to prevent egg damage while the gelatinous matrix is being
 100 removed. The solution was poured over two nested sieves of 75µm mesh 25µm mesh to collect
 101 the eggs. Eggs in the 25µm mesh sieve were rinsed with cold water and washed into a beaker.
 102 The cut roots in the original beaker were washed twice with water to obtain additional eggs. The
 103 number of eggs in 1ml of water was estimated by counting four samples of a milliliter each using
 104 Doncaster's counting dish under a stereomicroscope and the average was taken (Hussey and
 105 Barker, 1973).

106 **Sowing of seeds:** Okra seeds were sown at two seeds per hole in plastic pots with diameter
 107 22.5cm containing 6kg sandy loam soil mixture that has been sterilized. Two weeks after, the
 108 seedlings were thinned down to a healthy plant per pot.

109 **Inoculation of plants with nematode eggs:** Three weeks after emergence, the plants were
 110 inoculated with a calculated volume of 1000 eggs of *Meloidogyne* spp.. Eggs were applied near
 111 the base of plant by making holes around the Okra plants.

112 **Application of treatments:** Thirty (30g) each of the air-dried leaf powder and burnt leaf ash of
 113 *B. nitida*, *P. macrophylla* and *D. regia* were applied evenly on the surface of the soil in each
 114 bucket according to the treatments and their replications three days after inoculation. A synthetic
 115 nematicide (Carbofuran 3G) at 3.0g ai per plant was applied. The controls include plants whose
 116 soils were inoculated but untreated (control 1) and un-inoculated and untreated (control 2)
 117 respectively.

118 The treatment combinations were as follows:

- 119 1. T1 Nematode + Burnt leaf ash of *Baphia nitida*
- 120 2. T2 Nematode + Air dried leaf ash of *Baphia nitida*
- 121 3. T3 Nematode + Burnt leaf ash of *Pentaclethra macrophylla*
- 122 4. T4 Nematode + Air dried leaf ash of *Pentaclethra macrophylla*
- 123 5. T5 Nematode + Burnt leaf ash of *Delonix regia*
- 124 6. T6 Nematode + Air dried leaf ash of *Delonix regia*
- 125 7. T7 Nematode + synthetic nematicide (Carbofuran 3G).
- 126 8. T8 Inoculated but untreated (Control 1)

127 9. T9 Un-inoculated and untreated (Control 2)
 128 (NPK 15:15:15) was applied at the rate of 100kg/ha to give a rate of 0.4g per bucket. Water was
 129 applied to the plant as required.

130

131 DATA COLLECTION

132 Data collected at the end of the experiment after twelve weeks included:

- 133 • Plant height- measured using a meter rule in centimeter.
- 134 • Number of leaves-.
- 135 • Fresh and dry shoot and fresh root weights- measured using a digital laboratory weighing
 136 balance in grams (g).
- 137 • Number of eggs in root and larva in soil- extracted using the modified Baermann
 138 technique (Hooper, 1969), and counted using Domncaster's counting dish under an
 139 electronic stereomicroscope.
- 140 • Number of galls in roots-.
- 141 • Number of pods-.
- 142 • Weight of pods- measured in grams (g) using a digital laboratory weighing balance.

143 STATISTICAL ANALYSIS

144 The data collected were subjected to Analysis of Variance (ANOVA) and means were separated
 145 using Least Significant Difference (LSD) at 5% probability level ($P < 0.05$) by using computer
 146 software "Genstat Discovery Edition 4".

147 RESULTS AND DISCUSSION

148 RESULTS

149 The effect of treatments on plant height and number of leaves are shown in Table 1. As for plant
 150 height, significant differences ($P < 0.05$) were observed between control 2 (un-inoculated and
 151 untreated) (27.17cm) and all the treatments were less than control 2. There was also a significant
 152 difference between the plants treated with air-dried leaf powder of *B. nitida* (18.77cm) and
 153 control 1 (6.65cm). As for number of leaves, there was no significant difference recorded among
 154 the treatments including the controls. It is worthy noticing that the highest number of leaves
 155 (15.00) was recorded for the plants treated with burnt leaf ash of *B. nitida*, compared to the
 156 lowest (6.00cm) was recorded for control 1 (inoculated but untreated).

157 **Table 1: Effect of treatments on the plant height and number of leaves of okra infected**
 158 **with a root-knot nematode (*Meloidogyne* spp.).**

Treatments	Plant height(cm)	No. of leaves per plant
Burnt leaf ash of <i>Baphia nitida</i>	11.20	15.00
Air dried leaf powder of <i>Baphia nitida</i>	18.77	13.00
Burnt leaf ash of <i>Pentaclethra macrophylla</i>	12.62	8.00
Air dried leaf powder of <i>Pentaclethra macrophylla</i>	12.00	10.00
Burnt leaf ash of <i>Delonix regia</i>	11.07	13.00

Air dried leaf powder of <i>Delonix regia</i>	7.25	10.00
Synthetic nematicide (Carbofuran 3G)	10.25	12.00
Inoculated but untreated (control 1)	6.65	6.00
Un-inoculated and untreated (control 2)	27.17	11.00
LSD _(0.05)	6.25	NS

159 Where, NS = No Significant Difference show +/- values for each measure; provides idea of
160 variability between samples

161

162 Table 2 shows the effect of treatments on number and weight of pods. There was no significant
163 difference (P<0.05) between the treatments and the controls on the number of pods. It is worthy
164 noticed that the highest number of pods was recorded by control 2 (11) and equals to that
165 recorded to plants treated with synthetic nematicide (11), with the least number of pods (5.0)
166 recorded for plants that were inoculated but untreated (control 1). Significant differences
167 (P<0.05) were recorded on the weight of pods. All the treatments differed significantly from
168 control 2 (34.9g), except the plants treated with synthetic nematicide (28.3g). Also, there were
169 significant differences between all treatments and control 1. Nevertheless, plants treated with air-
170 dried leaf powder of *B. nitida* (23.4g) were compared favourably with the synthetic nematicide

171 **Table 2: Effect of treatments on the number and weight of Okra pods.**

Treatments	No. of pods per plant	Weight of pods (g)
Burnt leaf ash of <i>Baphia nitida</i>	7.0	21.3
Air dried leaf powder of <i>Baphia nitida</i>	10.0	23.4
Burnt leaf ash of <i>Pentaclethra macrophylla</i>	7.0	15.6
Air dried leaf powder of <i>Pentaclethra macrophylla</i>	8.0	21.4
Burnt leaf ash of <i>Delonix regia</i>	10.0	22.0
Air dried leaf powder of <i>Delonix regia</i>	6.0	16.1
Synthetic nematicide (Carbofuran 3G)	11.0	28.3
Inoculated but untreated (control 1)	5.0	6.1
Un-inoculated and untreated (control 2)	11.0	34.9
LSD _(0.05)	NS	9.75

172 Where, NS = No Significant Difference

173 The effect of treatments on the fresh shoot and dry shoot and fresh root weights as illustrated in
174 Table 3. As for fresh shoot weight, air-dried leaf powder of *B. nitida* (480g) which differed
175 significantly compared favourably with control 1 (205g) and burnt leaf ash of *Baphia nitida*
176 (133g), but did not with control 2 and other treatments. Regarding dry shoot weight, there was no
177 significant difference recorded among the treatments including the controls. Although the highest
178 dry shoot weight (85.2g) was recorded for the plants treated with burnt leaf ash of *P.*
179 *macrophylla* compared to the lowest (30g) was recorded for the inoculated but untreated
180 plants(control 1). Regarding fresh root weight all the treatments significantly differed from
181 control 2 (371g). However, the same treatments did not differ significantly from control 1 and
182 from each other.

183 **Table 3: Effect of treatments on fresh shoot, dry shoot and fresh root weights of Okra**
184 **infested with root-knot nematode (*Meloidogyne* spp.)**

Treatments	Fresh shoot weight (g)	Dry shoot weight (g)	Fresh root weight (g)
Burnt leaf ash of <i>Baphia nitida</i>	133.0	32.8	66
Air dried leaf powder of <i>Baphia nitida</i>	480.0	76.8	187
Burnt leaf ash of <i>Pentaclethra macrophylla</i>	335.0	85.2	118
Air dried leaf powder of <i>Pentaclethra macrophylla</i>	326.0	46.2	57
Burnt leaf ash of <i>Delonix regia</i>	308.0	64.0	136
Air dried leaf powder of <i>Delonix regia</i>	389.0	63.5	165
Synthetic nematicide (Carbofuran 3G)	362.0	70.0	132
Inoculated but untreated (control 1)	205.0	30.0	122
Un-inoculated and untreated (control 2)	629.0	68.2	371
LSD _(0.05)	267.1	NS	133.2

185 Where, NS = No Significant Difference

186 Effect of treatments on the number of nematode eggs and galls in roots, galls in roots, juveniles
 187 in soil are shown in Table 4. As for a number of galls in roots, all the treatments significantly
 188 compared favourably with control 1, but the treatments did not differ significantly reduced from
 189 each other. Similar observations were made on the number of eggs in root. As for number larva
 190 juveniles in soil, the treatments and the controls did not differ significantly from each other.
 191 However, apart from control 2 (000), plants treated with nematicide had the least number of
 192 juveniles (126) while the highest number (475) was recorded for plants treated with burnt leaf
 193 ash of *P. macrophylla*.

194 **Table 4: Effect of treatments on the population of *Meloidogyne* spp. in roots and soil.**

Treatments	No. of galls in roots	No. of eggs in roots	No. of larva juveniles in soil
Burnt leaf ash of <i>Baphia nitida</i>	0.75	126.00	350
Air dried leaf ash of <i>Baphia nitida</i>	0.25	102.00	250
Burnt leaf ash of <i>Pentaclethra macrophylla</i>	0.50	102.00	475
Air dried leaf ash of <i>Pentaclethra macrophylla</i>	1.75	202.00	351
Burnt leaf ash of <i>Delonix regia</i>	1.00	158.00	425
Air dried leaf ash of <i>Delonix regia</i>	0.75	126.00	300
Synthetic nematicide (Carbofuran 3G)	0.00	78.00	126
Inoculated but untreated (control 1)	5.00	575.00	425
Un-inoculated and untreated (control 2)	0.00	000.00	000
LSD _(0.05)	2.23	272.5	NS

195 Where, NS = No Significant Difference

196 DISCUSSION

197 The results on the effect of the treatments on plant height agreed with the findings of Bawa *et al.*,
 198 (2014) in which the plant extracts used had a significant effect on the height of tomato. They are
 199 still in agreement with the findings of Kayani *et al.*, (2012) who confirmed that the symptoms for
 200 identifying diseases caused by *Meloidogyne* spp. is the presence of galls, wilting, loss of vigor,
 201 yellowing of leaves. Also, Perry *et al.*, (2009) confirmed that the symptoms also include,
 202 yellowing of leaves and stunting, loss of vigor, wilting due to lack of moisture, decay of tissue
 203 due to secondary infection and yield loss. The growth reduction in crops such as Okra due to

204 nematode infestation varies depending on the population density of nematode, level of cultivar
 205 susceptibility and environmental condition to which the host is subjected to (Ononuju and
 206 Fawole, 2000).

207 The reduction in the fresh shoot and root weight could be due to the slow rate of action of the
 208 active ingredients in the plant materials (Barman and Das, 1996), or due to poor nutrients and
 209 water flow as a result of giant galls formed on the plant roots (Ploeg, 2001) which inhibit the
 210 normal growth and development of shoots and roots of plants affected.

211 The reduced number of nematode population and number of galls compared to control 1 agrees
 212 with the observations of Adegbite and Adesiyani (2001) who indicated that the higher the
 213 nematode population as the higher the number of galls and the higher the amount of damage
 214 occurred to plants which hinders the transportation of water and mineral nutrients from roots to
 215 aerial parts of the plant. The results obtained confirmed the findings of Ozores-Hampton (2002)
 216 that the use of organic amendments suppressed soil phyto-parasitic nematode populations.
 217 Abulusoro *et al.*, (2004) reported that susceptible tomato plants infected with root-knot
 218 nematodes (*Meloidogyne* spp.) show stunted growth, yield loss and conspicuous root galls, but
 219 that a number of plants are thought to contain biologically active ingredients which when applied
 220 in the soil reduce the incidence of plant-parasitic nematodes. Sikora and Fernandez (2005) also
 221 found that application of sesame seed extract reduced the incidence of root-knot nematodes and
 222 the severity of galling on okra roots. According to Akhtar and Mahmood (1993), sesame seed
 223 extracts have a systemic activity against nematodes which may have accounted for the lower
 224 number of galls and lower mean population in treated plants. Also, Vats *et al.*, (1996) reported a
 225 reduction of galls and egg masses when some *M. javanica* infected tomato plants were treated
 226 with leaf extracts of *Azadirachta indica* and *Eucalyptus tereticornis*.

227 The reduced number and weight of pods agree with Bolles *et al.*, (2012) who reported that
 228 nematode (*Meloidogyne* spp.) greatly decreased and delayed the production of pods. Also,
 229 Adesiyani, (1990) reported that some crop varieties could be effective hosts of root-knot
 230 nematode but suffer no statistically significant damage. However, the extent of damage is
 231 generally influenced by the cultivar, nematode species, level of soil infestation and environment
 232 (Ononuju, 1999). According to Gommers (1981) active compounds with nematicide activity
 233 which have been found to limit the activities of plant-parasitic nematodes in plants includes
 234 alkaloids, diterpenes, fatty acids, glucosinolates, isothiocyanates, phenols, polyacetylenes,
 235 sesquiterpenes and thienyls. These compounds were extracted from different plants part such as
 236 roots, leaves and so on which could be responsible for the positive effects of air-dried leaf ash
 237 and burnt leaf ash of these plants on the population of nematode in the soil and root as well as in
 238 pod formation.

239 CONCLUSION AND RECOMMENDATION

240 The different treatments significantly decreased the population of root-knot nematode
 241 (*Meloidogyne* spp.). There were significant differences between the controls and the treatments
 242 in some of the parameters measured, while there were no significant differences in others.
 243 Synthetic nematicide (Carbofuran 3G) appeared to be more effective in the control of root-knot
 244 nematode. However, the performance of the synthetic nematicide was not significantly different
 245 from the results obtained from air-dried powder of *Baphia nitida* and burnt leaf ash of *Delonix*
 246 *regia* at the rate applied. From the findings of this study, air-dried powder of *B. nitida* and burnt
 247 leaf ash of *D. regia* appeared to have significantly performed better than other treatments and is
 248 being recommended as an alternative for the synthetic nematicide (Carbofuran 3G) in the control

249 of root-knot nematode. Notwithstanding, further research work both in green house and field
250 trials at different rates are needed in order to authenticate these findings.

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UNDER PEER REVIEW